

Neuroimaging in hereditary spastic paraplegias: from qualitative cues to precision biomarkers

Grainne Mulkerrin, Marcondes C. França Jr, Jasmin Lope, Ee Ling Tan and Peter Bede

QUERY SHEET

This page lists questions we have about your paper. The numbers displayed at left are hyperlinked to the location of the query in your paper.

The title and author names are listed on this sheet as they will be published, both on your paper and on the Table of Contents. Please review and ensure the information is correct and advise us if any changes need to be made. In addition, please review your paper as a whole for typographical and essential corrections.

Your PDF proof has been enabled so that you can comment on the proof directly using Adobe Acrobat. For further information on marking corrections using Acrobat, please visit <http://journalauthors.tandf.co.uk/production/acrobat.asp>; <https://authorservices.taylorandfrancis.com/how-to-correct-proofs-with-adobe/>

The CrossRef database (www.crossref.org/) has been used to validate the references. Changes resulting from mismatches are tracked in red font.

AUTHOR QUERIES

- Q1** Please note that the ORCID for Peter Bede has been created from information provided through CATS. Please correct if this is inaccurate.
- Q2** The abstract is currently too long. Please edit the abstract down to no more than 200 words.
- Q3** Please note that funding information has been taken from information supplied with your manuscript submission. Please confirm if the funding information is accurate.
- Q4** References were not cited sequentially. Hence, references have been renumbered from Ref.141. Please check.
- Q5** The PubMed (<http://www.ncbi.nlm.nih.gov/pubmed>) and CrossRef (www.crossref.org/) databases have been used to validate the references. Mismatches between the original manuscript and PubMed or CrossRef are tracked in red font. Please provide a revision if the change is incorrect. Do not comment on correct changes.

REVIEW



Neuroimaging in hereditary spastic paraplegias: from qualitative cues to precision biomarkers

Q1 Grainne Mulkerrin^a, Marcondes C. França Jr^b, Jasmin Lope^c, Ee Ling Tan^c and Peter Bede ^{a,c}

5 ^aDepartment of Neurology, St James's Hospital, Dublin, Ireland; ^bDepartment of Neurology, the State University of Campinas, São Paulo, Brazil; ^cComputational Neuroimaging Group, Trinity College Dublin, Ireland

ABSTRACT

Introduction: Hereditary spastic paraplegias (HSP) include a clinically and genetically heterogeneous group of conditions. Novel imaging modalities have been increasingly applied to HSP cohorts, which helps to quantitatively evaluate the integrity of specific anatomical structures and develop monitoring markers for both clinical care and future clinical trials.

Areas covered: Advances in HSP imaging are systematically reviewed with a focus on cohort sizes, imaging modalities, study design, clinical correlates, methodological approaches, and key findings.

Expert opinion: A wide range of imaging techniques have been recently applied to HSP cohorts. Common shortcomings of existing studies include the evaluation of genetically unconfirmed or admixed cohorts, limited sample sizes, unimodal imaging approaches, lack of postmortem validation, and a limited clinical battery, often exclusively focusing on motor aspects of the condition. A number of innovative methodological approaches have also been identified, such as robust longitudinal study designs, the implementation of multimodal imaging protocols, complementary cognitive assessments, and the comparison of HSP cohorts to MND cohorts. Collaborative multicenter initiatives may overcome sample limitations, and comprehensive clinical profiling with motor, extrapyramidal, cerebellar, and neuropsychological assessments would permit systematic clinico-radiological correlations. Academic achievements in HSP imaging have the potential to be developed into viable clinical applications to expedite the diagnosis and monitor disease progression.

ARTICLE HISTORY

Received 8 June 2022
Accepted 24 August 2022

KEYWORDS

Hereditary spastic paraplegia; HSP; MRI; PET; biomarker; cognition

1. Introduction

Hereditary spastic paraplegia (HSP) is an umbrella term for a group of genetically and phenotypically diverse inherited neurodegenerative disorders, characterized by progressive spasticity and weakness primarily affecting the lower limbs. As proposed by Harding's criteria [1], HSP can be clinically divided into 'pure' and 'complicated' forms. Pure-HSP (pHSP) presents with insidious onset progressive spasticity of the lower limbs, hyperreflexia, and upper motor neuron (UMN) sign. Weakness of the lower limbs can be mild, with a characteristic discordance between the severity of spasticity and relatively preserved power. In complicated-HSP (cHSP), spastic paraparesis is accompanied by additional neurological signs. Optic atrophy, extrapyramidal features, cerebellar signs, cognitive impairment, sensorineural deafness, and epilepsy are some of the additional neurological manifestations associated with cHSP. Initial symptoms are typically subtle; gait disturbance, stiffness and falls, and disease progression is often slow, but can lead to significant disability [2].

Over 100 loci/88 genes have been implicated in the pathogenesis of HSP to date [3]. Inheritance can be autosomal dominant (AD), autosomal recessive (AR), X-linked, mitochondrial, or sporadic [4]. The genetic classification for HSP is based on sequential numbering of genes, using

a spastic paraplegia gene (SPG) prefix. The most common genotypes seen in clinical practice are SPG4 and SPG11. SPG4 is a pHSP that exhibits AD-inheritance of the spastin (SPAST) gene and accounts for up to 60% of all HSP cases. SPG11 is the most prevalent AR-inherited HSP accounting for approximately 8% of all HSP cases and categorized as a cHSP with additional features such as cognitive impairment, peripheral neuropathy, ataxia, and urinary symptoms. SPG3A is a clinically pure HSP phenotype and the second most common AD-inherited HSP, accounting for 5–10% of AD HSPs that are negative for SPAST mutation. SPG15 is inherited in an AR manner and phenotypically similar to SPG11. Finally, SPG7 that tends to manifest in the fourth decade with spastic paraparesis and cerebellar signs, is a cHSP inherited in an AR manner [5].

There is a striking paucity of postmortem studies on HSP. The 'dying-back axonopathy' phenomenon is traditionally considered as the pathological basis of the condition [6] with length-dependent degeneration of the corticospinal tracts, fasciculus gracilis, and spinocerebellar tracts [6]. Recent postmortem studies seem to support the 'dying-back' theory by describing axonal degeneration of the corticospinal tracts, with distal axons more severely affected [7].

The clinical indication for neuroimaging in HSP is to rule out alternative structural, vascular neoplastic, and

Article highlights

- Hereditary spastic Paraplegias (HSP) include a clinically and genetically heterogeneous group of conditions with diverse disability profiles.
- Radiological cues, such as the 'ear of the lynx' sign, thinning of the corpus callosum, cerebellar atrophy, high signal along the corticospinal tracts, and optic nerve hypoplasia have limited specificity to HSP genotypes.
- The practical relevance of robust, prospective computational imaging studies stems from their ability to ascertain HSP-specific imaging signatures, the prospect of expediting the diagnosis, and the development of viable monitoring markers for both clinical care and future pharmaceutical trials.
- A wide range of quantitative **gray** and white matter techniques have been recently applied to HSP cohorts including cortical thickness measurements, morphometric approaches, cortical and subcortical volumetry, spinal cord protocols, tractography, spectroscopy, **PET**, and **SPECT**.
- Stereotyped shortcomings of existing imaging studies include the evaluation of genetically unconfirmed or genetically admixed cohorts, limited sample sizes, unimodal imaging approaches, and a limited clinical battery often exclusively focusing on the motor aspects of the condition.
- Collaborative initiatives with **harmonized** protocols may overcome sample limitations; comprehensive clinical profiling with motor, extra-pyramidal, cerebellar, **cognitive**, and **behavioral** assessments would permit systematic clinico-radiological correlations, and multi-timepoint longitudinal study designs will allow the nuanced **characterization** of anatomical propagation patterns.

75 neuroinflammatory diagnoses, such as spinal spondylosis, spinal or cerebral AVMS, Chiari malformation, **and** primary progressive multiple sclerosis. Metabolic causes such as B12 vitamin deficiency or infectious causes such as HIV-myelopathy, **HTLV1**, or neurosyphilis are also potential differential diagnoses to consider during the initial investigations. Given the relatively low incidence of the condition, insidious onset, slow progression, the heterogeneity of initial presentations, and wide range of differential diagnoses, patients often face a protracted diagnostic journey. Diagnostic delay, misdiagnoses, and unnecessary interventions are not uncommon [8]. The cost of testing for large panels of HSP-associated genes may be prohibitive in some jurisdictions, family history may not be readily available in some cases, and clinical overlap with other neurodegenerative conditions such as primary lateral sclerosis (PLS) may also contribute to diagnostic delay [9]. Patients often seek second opinions **and** may attend multiple specialists before the diagnosis is confirmed. Patients may be initially referred to **orthopedic** services, **neurosurgeons**, or physiotherapists before review in a tertiary neurology service. Establishing the diagnosis in a timely manner is hugely important so that unnecessary interventions can be avoided, multidisciplinary interventions can be promptly initiated, **and** genetic **counseling** can be **organized**. In the absence of effective disease-modifying therapies, supportive care is the mainstay of HSP management. A multidisciplinary approach **comprising** pharmacological management of spasticity, fall prevention, physiotherapy for rehabilitation, and occupational therapy for disability are key strategies to maintain independence. No effective disease-modifying therapies have been confirmed to **date**, and clinical trials for new agents are limited by the lack of a reliable biomarker for the disease.

Computational neuroimaging is increasingly **recognized** as a powerful **noninvasive** tool in low-incidence neurodegenerative conditions [10–13], which readily capture disease-specific patterns of pathology in vivo based on regional integrity metrics. Quantitative neuroimaging is also increasingly **utilized** to track disease burden patterns longitudinally and has a viable biomarker role in a multitude of neurodegenerative conditions [14]. A novel frontier of neuroimaging is quantitative spinal cord imaging, which has successfully captured segmental **gray** and white matter alterations in motor neuron diseases, spinal muscular atrophy, **and** spinal and bulbar muscular atrophy [15–19]. Given the considerable progress in MR technology, the emergence of high-field and ultra-high field scanners, the availability of robust image processing pipelines and open-source analysis suites, we sought to systematically review the imaging literature in HSP.

2. Body

2.1. Methods

A formal literature review was conducted on PubMed between October 2021 and December 2021 using the keywords 'hereditary spastic paraplegia' and 'hereditary spastic paraparesis' in combination with the individual search terms: 'imaging,' 'neuroimaging,' 'magnetic resonance imaging,' 'magnetic resonance spectroscopy,' 'positron emission tomography,' and 'diffusion tensor imaging.' Only articles published in English were included. No exclusion criteria were set based on **the year** of publication. Additional articles were also included based on references **to** relevant review articles. Review articles, opinion **pieces**, **and** editorials were excluded. Identified original papers were systematically reviewed for sample size, genetic screening, choice of controls (healthy/disease), qualitative versus quantitative imaging, primary imaging methods (structural, diffusion, spectroscopy, functional, metabolic, **spinal**, etc.), unimodal versus multimodal imaging, image analysis software, **postmortem** validation, clinical assessment (scales, scores, motor, **and** cognitive), clinico-radiological **correlations**, and main study findings.

2.2. Results

Based on the above search criteria, a total of 108 original research papers were identified. **Forty-one** of these described qualitative findings, primarily on MRI. Sixty-seven studies **utilized** quantitative imaging such as diffusion tensor imaging (DTI – 30 studies), MR spectroscopy (MRS – 16 studies), PET-CT (18 studies) and functional MRI (fMRI – 3 studies). The vast majority of studies focused on cerebral imaging. Spinal alterations were specifically appraised by 23 studies, but only 4 **utilized** quantitative approaches.

The majority of imaging studies **on** HSP suffer from grave sample size limitations. Only two imaging studies were identified with greater than 50 HSP patients [20,21]. Forty-five studies included less than 10 participants, and a further 18 studies were single patient case-reports. The vast majority of identified studies were cross-sectional in design, **while** longitudinal imaging follow-up was only performed in

eight studies. Some clinical assessment was performed in almost all studies, and disease severity, mainly SPRS, was reported in 23 studies. Clinico-radiological correlations were explored in 23 studies, most commonly with disease duration [20,22–29], cognition [22,30–35] and spasticity [20–27,29,34,36–38]. We identified no studies that correlated in vivo imaging patterns to postmortem pathological findings.

2.3. Typical findings on visual inspection – qualitative imaging

The diagnosis of HSP can be challenging, especially given the clinical and genetic heterogeneity of the condition. While MRI may look normal on visual inspection [39–46], stereotyped cerebral and spinal cord alterations are commonly appreciated (Figure 1).

2.3.1. Spinal cord atrophy

A large study of genetically unconfirmed pHSP and cHSP detected considerable cervicothoracic spinal cord atrophy compared to demographically matched controls [47]. The same study found no difference between the pHSP and cHSP subgroups. In another group of clinically confirmed HSP patients, a reduced cervical spine cross-sectional area was observed in patients in comparison to controls [34]. The anteroposterior (AP) diameters of the thoracic spine at T3 and T9 were significantly lower than controls in another qualitative

study looking at clinically confirmed pHSP [48]. In genetically confirmed HSP, spinal cord atrophy may be observed in SPG5 [41], SPG6 [42], SPG7 [49]. Interestingly, a syrinx was observed in two SPG54 patients [50]. However, cord atrophy is not a consistent finding; in other studies, MRI of the spinal cord looked relatively normal in SPG3A [51,52], SPG4 [39], SPG7 [53,54], SPG11 [55], SPG15 [56] and SPG46 [57]. This inconsistency highlights the limitations of visual inspection, disease-associated radiological cues, and qualitative assessments. None of the above articles describing ante mortem qualitative spinal changes provided descriptions of postmortem cord changes.

2.3.2. The ‘ears of the lynx’ sign

The ‘ears of the lynx’ sign refers to T2/FLAIR hyperintensity on axial views at the tip of the frontal horn of the lateral ventricles, named after the tufts of hair at the tips of the ears of a lynx cat. This sign is typically associated with SPG11 and SPG15 mutations [58]. The ears of the lynx sign is found to be relatively sensitive (78.8–97%) and rather specific (90.9–100%) to SPG11 and SPG15 in a blinded study with four different raters [58].

2.3.3. Thin corpus callosum

Thin corpus callosum (TCC) is a common finding in HSP. Prior to the discovery of the many genetic subtypes of HSP, autosomal recessive HSP with thin corpus callosum (AR-HSP-TCC)

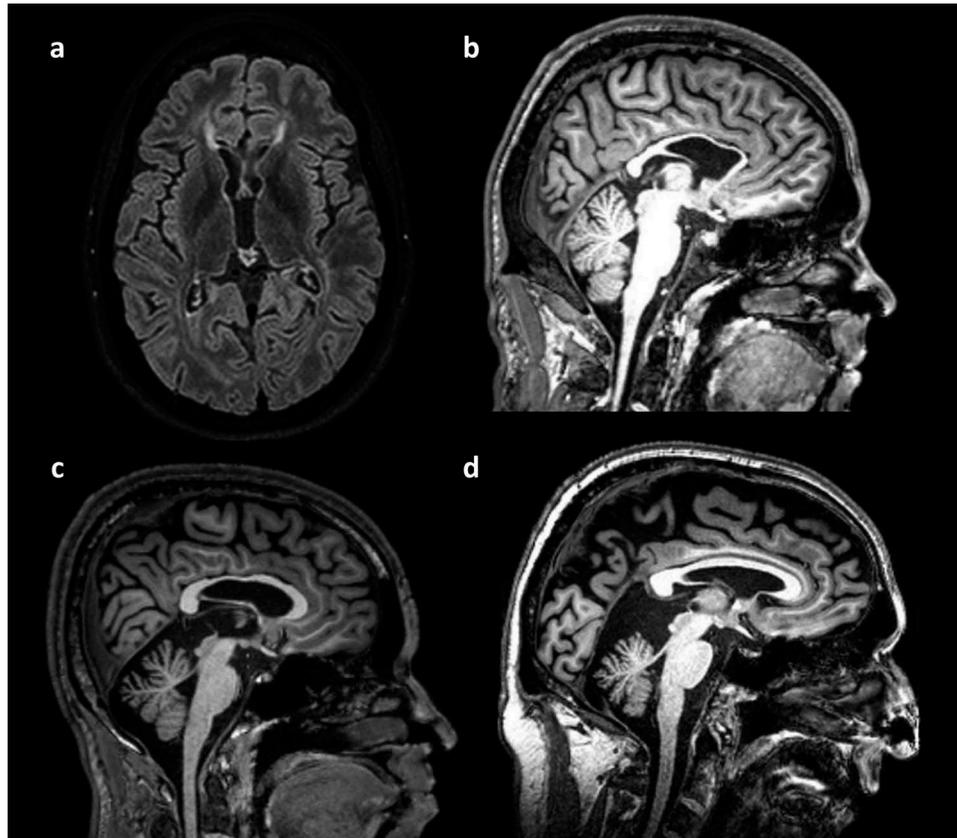


Figure 1. Illustrative examples of radiological changes commonly observed in HSP on visual inspection. a: ‘Ears of the lynx’ sign on axial FLAIR imaging in a 21 year-old female patient with SPG11. b: thin corpus callosum on sagittal T1-weighted imaging in a 25 year-old male patient with SPG11. c: Cerebellar atrophy on sagittal T1-weighted imaging in a 37 year-old male patient with SPG15. d: Cerebellar atrophy and arachnoid cyst on sagittal T1-weighted imaging in a 35 year-old male patient with SPG7.

210 was a commonly used terminology to describe what is now
 a probably genetically diverse subgroup. HSP-TCC is most
 commonly reported in association with SPG11 [55,59–66]
 and SPG15 [56,59,63,67–69]. However, TCC may also be
 215 a feature in SPG3A [51,52], SPG21 [70], SPG 28 [71], SPG30
 [62,72], SPG35 [63,73], SPG45 [74], SPG46 [57,75], SPG47 [76],
 SPG48 [63], SPG52 [62], SPG54 [50,62,77–80] and SPG78 [81].
 TCC is not a consistent finding in many of these subtypes, for
 example, in a study of SPG3A it was only noted in 1 of the 18
 220 patients imaged [51]. In a small study (n = 2) which imaged
 patients with SPG11 **longitudinally**, TCC progressed over
 a **5-year** interval [55]. In **contrast**, however, another case report
 of AR-HSP-TCC detected no progressive thinning of the corpus
 callosum over a 4 years interval [82]. It is noteworthy that thin
 corpus callosum (CC) is by no means specific to HSP and is
 225 commonly observed in ALS, PLS, **CBS**, and other neurodegenerative
 conditions [83–87].

2.3.4. Cerebellar atrophy

Cerebellar atrophy has been described in SPG7 [54,59,88–90],
 SPG21 [70], SPG30 [62,91], SPG31 [62], SPG35 [62,63,92], SPG39
 230 [59], SPG 46 [57,75], SPG50 [93], SPG78 [81,94], and SPG54 [80].
 Cerebellar atrophy is thought to be progressive on longitudinal
 imaging in a small (n = 2) SPG30 study [72], and in a single
 patient with SPG78 who was followed longitudinally [94]. In
 contrast, other case reports of SPG 30 [91] and SPG47 [76]
 235 observed relatively stable cerebellar volumes over time. Other
 posterior fossa abnormalities, such as retro-cerebellar fluid
 collection and cerebellar hypoplasia in SPG4 [95] and uncom-
 plicated arachnoid cysts of the cerebellopontine angle in SPG4
 [96] were also reported. In some cases, cerebellar atrophy is
 240 accompanied by brainstem hypoplasia/atrophy, e.g. SPG35
 may present with **pontocerebellar** hypoplasia [92]. Similar to
 the above qualitative cues, cerebellar atrophy is far from
 unique to HSP, and is commonly observed in motor neuron
 disorders and other neurodegenerative conditions [97–100].

2.3.5. Cerebral atrophy

245 Cerebral atrophy on routine brain MRI has been commonly
 observed in HSP. In a group of genetically **heterogeneous** HSP
 patients, cortical atrophy was reported in 25/58 patients
 imaged [21]. **Likewise**, in clinically (but not genetically) con-
 250 firmed HSP, mild brain atrophy was observed in 4 of 9 patients
 [101]. In studies that looked at those with a causative gene for
 HSP, cerebral or cortical atrophy was noted in SPG4 [32], SPG7
 [49], SPG11 [60,61], SPG 21 [70], SP30 [62], SPG56 [62], and
 SPG78 [81]. In a subset of SPG4 patients with cognitive impair-
 255 ment, frontotemporal atrophy **was** correlated with the degree
 of cognitive impairment [102]. As with the above signs, cere-
 bral volume loss is notoriously **nonspecific** and has a limited
 role in swaying a diagnostic dilemma.

2.3.6. White matter hyperintensities

260 White matter hyperintensities (WMH) are **nonspecific** findings
 associated with numerous pathological processes. WMH has
 been reported in SPG2 [103], SPG4 [62], SPG5 [104], SPG11 [61–
 63,105], SPG15 [63,67,68], SPG21 [70], SPG31 [62], SPG35
 [62,63,73,92,106], SPG45 [74], SPG47 [107], SPG48 [63], SPG52

[62] and SPG54 [50,62,77]. In a case series of **two** related SPG2 265
 patients, the index patient was initially misdiagnosed with primary
 progressive multiple sclerosis based on presentation and imaging
 findings [103]. In one patient with SPG2 who was imaged longi-
 tudinally, the WMH was stable on repeat imaging [108].

2.3.7. Subcortical alterations

270 While deep brain structure abnormalities have been observed
 on routine brain MRI in certain HSP subtypes, these findings
 are inconsistent in the literature. In SPG35, mild hypointensity
 of the globus pallidus was noted in 77% of cases on the
 retrospective review of MR images [92]. Hypothalamic atrophy 275
 was observed in SPG11 cases compared with controls in
 another study [24]. Atrophy of the caudate nucleus was
 noted in an imaging study of three related patients with
 genetically confirmed SPG78 exhibiting prominent extrapyra-
 midal signs [81]. Hyperintensity of the dentate nucleus was 280
 observed both in SPG7 [89] and KIF1A [109,110]. Subcortical
 iron deposition has been observed in SPG28 [71]. Iron deposi-
 tion has also been described in SPG35 cases [106] **but** not
 confirmed by a **larger** series (n = 19) on the same gene [92].

2.3.8. Optic nerve atrophy

285 Optic nerve atrophy or hypoplasia has been described in
 several HSPs, including SPG30/KIF1A [109,110], SPG7 [111]
 and SPG54 [62]. The spectrum of ophthalmological abnormal-
 ities observed in HSP [112] comprises other changes such as
 yellow retinal ‘flecks’ in SPG11 and SPG15, progressive external 290
 ophthalmoplegia in SPG7 [113], **and** supranuclear gaze palsy
 in SPG7 [49] **but** the discussion of these manifestations and
 macular optical coherence tomography (OCT) findings is
 beyond the scope of this review.

2.4. Computational neuroimaging – quantitative imaging

2.4.1. Spinal cord imaging

295 There is a striking scarcity of quantitative spinal imaging studies
 on HSP. Existing studies have either performed morphometric
 [38] **or** diffusivity analyses [114]. Spinal cord cross-sectional area 300
 reductions were captured in SPG11 patients in comparison to
 controls [26] which correlated inversely with both disease dura-
 tion and severity. Spinal cord atrophy, involving both **gray** and
 white matter **components**, has been observed in SPG11 and
 SPG4, but not in SPG3A or SPG7 [28]. The **gray** matter area of 305
 the spinal cord was inversely correlated with **the disease** dura-
 tion in the SPG4 group, but no significant correlation was seen
 in the other genetic subtypes [28]. One study ascertained CST FA
 reductions at the cervical level [37], which correlated with dis-
 ease duration, but not symptom severity as assessed by the 310
 SPRS. A recent SPG4 study detected FA reductions and increased
 radial diffusivity (RD) at cervical and thoracic levels **in both** the
 lateral and dorsal columns. Key study characteristics of imaging
 studies evaluating cord alterations are **summarized** in Table 1.

2.4.2. Structural, morphometric, and volumetric studies

315 Cerebral, **cerebellar**, and corpus callosum volume reductions
 are commonly detected by quantitative HSP studies [115].

Table 1. Imaging studies commenting on spinal alterations.

First author, year of publication	Study design Imaging modality	Study groups	Main conclusions
Hsu, 2021 [51]	Cross-sectional, 2 center s, retrospective descriptive, qualitative MRI brain and spinal cord	SPG3A (n = 18) No controls	MRI brain: thin corpus callosum in one patient, otherwise normal Normal cervicothoracic spine
List, 2019 [37]	Cross-sectional, single- center ; quantitative imaging, clinico-radiological correlation Brain and spinal cord DTI, TBSS, and ROI analyses	SPG4, SPG5, SPG31 Disease (n = 20) Control (n = 17)	TBSS revealed corticospinal tract degeneration in pHSP. Reduced FA and increased AD in CC, CST, optic radiation, frontal lobe, parieto-occipital regions, temporal lobe, and brainstem. No cerebellar FA changes. Significant negative correlation between disease duration and C Spine FA. No correlation found between imaging findings and SPRS.
Nielsen, 1998 [39]	Single- center ; cross-sectional, qualitative MRI brain and spinal cord	SPG4 (n = 16) imaged, age- and sex-matched healthy controls	No obvious brain or spinal cord MRI abnormalities compared with controls
Lan, 2014 [41]	Single- center ; cross-sectional, qualitative, no clinico-radiological correlations, MRI brain and spine	SPG5 (n = 5) HSP (n = 25)	Normal MRI brain in SPG5. Thoracolumbar atrophy (not quantified) was present in 3/5 patients with SPG5.
Agosta, 2015 [34]	Single- center ; cross-sectional, quantitative, clinico-radiological correlations, MRI brain and cervical cord, DTI, VBM, TBSS	No controls pHSP (n = 20) cHSP (n = 24) Healthy controls (n = 7)	VBM: no difference between HSP and controls. TBSS: pHSP and cHSP FA reduction in CST, CC, external capsule, and most association WM tracts compared to controls. Reduced cervical spinal cord cross-sectional area in HSP compared to controls. WM alterations correlate with spasticity and cognitive impairment.
Sperfeld, 2005 [47]	Single- center ; Cross-sectional, qualitative, no clinico-radiological correlations, MRI cervical and thoracic cord	HSP (n = 30) Not genetically confirmed Controls (n = 96)	Significant cervical and thoracic spinal cord atrophy in HSP compared to controls. No difference between pHSP and cHSP.
Krabbe, 1997 [48]	Single- center ; cross-sectional, qualitative, no clinico-radiological correlations, MRI brain and whole spine	Not genetically confirmed Controls (n = 15)	The patients had thinner corpus callosum than controls. The anteroposterior diameters of the spinal cord at T3 and T9 were significantly reduced in patients compared to controls
Faber, 2018 [26]	Single- center ; cross-sectional, quantitative clinico-radiological correlations, DTI, cortical thickness, deep gray matter volumes, spinal cord morphology	SPG11 (n = 25) Genetically confirmed age- and sex-matched controls (n = 25)	Reduced SC area was in SPG11 compared with controls. Cortical thickness reductions in bilateral motor cortices, limbic structures, cingulate gyri, left parahippocampal gyrus, bilateral superior temporal sulcus, left precuneus, and superiorparietal cortices. SPRS negatively correlated with cortical thickness in precentral and paracentral cortices bilaterally. Reduced deep gray matter structure volumes. Widespread and diffuse WM alterations. SPRS and disease duration correlated inversely with spinal cord area.
Servelhere, 2021 [28]	Single- center ; cross-sectional, quantitative clinico-radiological correlations, MRI cervical spine: T2*weighted 3D imaging	SPG3A (n = 7), SPG4 (n = 12), SPG7 (n = 10), SPG11 (n = 8), healthy controls (n = 21)	No spinal cord atrophy identified in SPG3A and SPG7. In SPG4 & SPG11: spinal cord atrophy compared to controls, in both GM and WM. Inverse correlation between GM area and disease duration in in SPG4.
Warnecke, 2007 [49]	Cross-sectional, single- center ; descriptive, no clinico-radiological correlations, MRI brain & cervicothoracic spine, DTI Brain (FA)	Controls (n = 50)	Qualitative descriptions: normal cervicothoracic MRI, cerebellar atrophy and mild frontal cerebral atrophy. DTI: reduced WM integrity in CST, the frontal lobes, and the midbrain
Guo Liu, 2008 [42]	Single- center ; cross-sectional, qualitative, no clinico-radiological correlations, MRI brain & cervicothoracic cord	SPG6 (n = 6) Healthy controls (does not quantify)	Spinal cord atrophy observed in all affected patients. Reduced anteroposterior diameters, transverse diameters and cross-sectional areas at the C2, C3, C7, T1, T2, T3, and T4 levels. MRI brain: normal
Winner, 2004 [55]	Single- center ; longitudinal (imaging 5 years apart), qualitative and quantitative, no clinico-radiological correlations, MRI brain & spine, PET CT brain	SPG11 (n = 2) No controls	MRI brain: progressive CC thinning, symmetrical white matter lesions, not progressive. MR spine: normal. PET-CT: hypometabolism in frontotemporal and temporal cortices. Progressive in 1 of 2 patients.
Orlacchio, 2011 [52]	Single- center ; cross-sectional, qualitative no clinico-radiological correlations, MRI brain and spinal cord	SPG3A (n = 1)	MRI brain: thin corpus callosum, MRI spine: normal
Hourani, 2009 [101]	Single- center ; cross-sectional, qualitative no clinico-radiological correlations, MRI brain and whole spine	HSP (n = 9) Clinically confirmed (Harding's criteria) No controls	White matter hyperintensities in 6/9 patients, mild brain atrophy in in 4/9 patients, thin corpus callosum in 4/9, mild spinal cord atrophy in 3/9
Cao, 2013 [124]	Single- center ; cross-sectional, qualitative and quantitative, no clinico-radiological correlations, MRI brain & spinal cord DTI	SPG11 (n = 2) controls (n = 11)	MRI brain: thin CC, MRI spinal cord: normal, DTI: increased MD and decreased FA in CC, frontal WM, temporal lobe WM compared to healthy controls
Rezende, 2015 [38]	Single- center ; cross-sectional, clinico-radiological correlations, volumetric analyses, TBSS, and tractography	SPG4 (n = 11) Healthy controls (n = 23)	Spinal cord volumetric analysis: atrophy in HSP group. Brain volumetric analyses: no abnormality. TBSS: reduced FA in CST, cingulate gyri and splenium of the CC. FA of the CC and CST correlated with disease severity (SPRS). No correlation between tractography and SPRS.
Navas-Sánchez [114]	Cross-sectional, clinico-radiological correlations, volumetric analyses, FA and RD, assessments	SPG4 (n = 12) Controls (n = 14)	Reduced cervical cord cross-sectional area with anteroposterior flattening. Reduced FA and increased RD at cervical and thoracic levels in both the lateral and dorsal columns. FA changes correlate with disease severity.

However, in some studies, volumetric analyses have not captured brain atrophy in SPG4-associated HSP [38,44]. In genetically confirmed HSP, reduced cortical thickness has been observed in SPG4 [22], SPG11 [20,26], and SPG8 [20]. Conversely, the lack of cortical thickness reduction was highlighted by some SPG4 and SPG3A studies [20]. Voxel-based morphometry often captures gray matter alterations in the prefrontal cortex [25] and the thalami [23,25,26,29]. Brain parenchymal fractions also reveal significant brain atrophy in HSP patients compared with controls [31,116]. Basal ganglia atrophy is a common finding in HSP patients and has been reported both in SPG4 [20,22,23] and SPG11 [20,117]. In line with subjective and qualitative observations, cerebellar atrophy has also been detected in SPG11 [20], SPG7 [20] and SPG4 [23].

2.4.3. Diffusion tensor imaging (DTI)

Tract-based spatial statistics (TBSS) typically reveals corticospinal tract degeneration in HSP [23,25,34,37,38,118]. In genetically confirmed HSP, corticospinal white matter alterations were specifically observed in SPG4 [22,23,38,44], SPG7 [40,49] and SPG11 [64,119]. White matter changes are also commonly observed in other brain regions, often with limited change over time on longitudinal follow-up [25]. Specific anatomical locations that exhibit reduced white matter integrity in HSP include the corpus callosum [25,34,37,38,116,118,120], the brainstem [37], the optic radiation [37], the frontal lobes [37], fronto-occipital regions [25], parieto-occipital regions [37], the external capsule [34], the temporal lobes [37], the longitudinal fasciculi [25,118], the cingulate gyri [25,38] and the thalamic radiations [118]. Anatomically widespread, 'generalized,' white matter degeneration was observed in SPG11 with reference to controls [26,121]. The corpus callosum is affected in SPG4 [22,23], SPG7 [20,122] and SPG11 [20,64,119,121–124]. Extra-motor fractional anisotropy (FA) reductions are commonly observed in SPG11 including the thalamus [123], the frontal and peritrigonal white matter [122], periventricular white matter [125] and the cerebellum [123]. In SPG7, the corticospinal tract [40,49], frontal lobes [40,49], and midbrain [49] exhibit reduced white matter integrity. Multiple SPG4 studies describe particularly widespread white matter degeneration, in particular in the corticospinal tracts [23,44], thalamus [23], brainstem [23] and cerebellum [23]. The distinguishing WM signature of HSP in comparison to PLS and ALS is poorly characterized [126–128]. The main study characteristics of structural and diffusivity studies are summarized in Table 2.

2.4.4. PET-CT

PET-CT, SPECT, and DAT studies on HSP suffer from considerable sample size limitations. Nonetheless, these studies provided important academic insights with regard to metabolic signatures in specific HSP subtypes and the likely substrate of the motor and extra-motor deficits observed clinically. Thalamic hypometabolism is routinely observed in SPG11 on FDG-PET-CT [60,65,66]. This may be progressive in some patients over a 10-year follow-up [60]. In another study of genetically unconfirmed HSP-TCC, progressive thalamic hypoperfusion was observed on SPECT over 4 years [82]. Reduced striatal density has also been observed in clinically confirmed

HSP [129], SPG11 [27], SPG78 [94], SPG7 [54]. This correlated with SPRS and disease duration in the SPG11 group [27]. Cortical hypometabolism may also be a feature of HSP. This has been reported in the frontal cortex of SPG3A [130], and in the frontotemporal cortical regions in SPG11 [55]. On SPECT, hypoperfusion of the frontotemporal cortices in SPG4 was linked to cognitive impairment [32]. Other studies noted reduced regional cerebral blood flow in the left frontotemporal cortex in SPG4, without direct correlations with cognitive profiles [35]. Cerebellar hypometabolism is also commonly observed in HSP; it has been described both in SPG11 [131] and SPG5 [104]. On SPECT, cerebellar hypoperfusion has also been observed in SPG4 [132].

Similar to initiatives in other MNDs such as ALS [133,134], functional reorganization in response to structural degeneration is increasingly investigated. Cortical activation patterns in SPG4-linked HSP were evaluated by rCBF PET-CT during affected and unaffected lower limb tasks [135]. During ankle movement, patients exhibited increased rCBF in the primary motor cortices (PMC), the supplementary motor areas (SMAs), and the premotor cortex compared to controls; which may be indicative of motor cortical reorganization or adaptation to degenerative change. Conversely, there are also reports of normal PET-CT imaging HSP, specifically in SPG2 [108] and SPG7 [54].

2.4.5. MR spectroscopy (MRS)

Sixteen studies utilized magnetic resonance spectroscopy to evaluate the underlying biochemical and metabolic processes determining the clinical phenotypes of HSP. Metabolic abnormalities may be a consistent feature of HSP, however the inconsistency between studies with regard to region-of-interest (ROI) makes the systematic integration of study findings challenging. Investigated ROIs include the centrum semiovale [60,108,136,137], basal ganglia [50,77,78,138], corpus callosum [105,119], the frontal white matter [30,136], occipitoparietal regions [46,60], occipito-frontal region [108], mid-occipital region [108], parietal region [136], the cerebellum [119], supraventricular white matter [65], periventricular white matter [105], precentral white matter [21], the internal capsule [60], the corona radiata [105] and the brainstem [136]. The most common MRS finding in HSP is N-acetyl-aspartate (NAA) level reduction in various brain regions, but a large study (n = 70) identified normal precentral metabolite levels [21]. In SPG11, NAA reductions were reported in the frontal white matter [60], corona radiata [105], periventricular white matter [65,105], the centrum semiovale [136], and corpus callosum [119]. In genetically diverse HSP, reduced NAA/Cr levels were observed in the right prefrontal region [25]. However, the findings are inconsistent; NAA was reduced in the occipito-frontal white matter in only one of the two patients with SPG2 in another study [108], and increased NAA levels were also reported in the centrum semiovale [137]. NAA/Cr reductions in the corona radiata in SPG11 inversely correlated with disability scores [105]. Myoinositol (MI) changes are the second most common MRS alterations reported in HSP. Increased ml/Cr ratio has been described in occipito-parietal regions in SPG5A [46], occipito-frontal areas

Table 2. Structural and diffusion studies.

First author, year of publication	Study design Imaging modality	Study groups	Main conclusions
Servelhere, 2021 Servalhere, Rezende [20]	Single-center, cross-sectional, clinico-radiological correlations, DTI, cortical thickness, BG volumetry, cerebellar GM analyses, WM integrity analyses	SPG3A (n = 9), SPG4 (n = 27), SPG7 (n = 10), SPG8 (n = 9), SPG11 (n = 29), healthy controls (n = 84)	SPG3A: No brain abnormality, SPG4: Posterior WM changes, caudate and thalamic atrophy, SPG7: CC alterations, cerebellar atrophy, SPG8: cortical thinning at pre- and post-central gyri, pallidum atrophy, SPG11: Cerebral cortical thinning, cerebellar, BG atrophy, WM changes in CC, CR, and subcortical pathways. Clinico-radiological correlations only in SPG11 cases.
Lindig, 2015 [23]	Single-center, cross-sectional, retrospective, quantitative, clinico-radiological correlations, TBSS, VBM	SPG4 (n = 15), controls (n = 15)	Widespread WM change in the CC, medio-dorsal thalamus, parieto-occipital regions, upper brainstem, cerebellum, and corticospinal tract.
Kassubek, 2006 Kassubek, Sperfeld [31]	Single-center, cross-sectional, clinico-radiological correlations with cognitive domains, brain parenchymal fractions	p-HSP (n = 21), c-HSP (n = 12) not genetically confirmed, healthy controls (n = 30)	Imaging findings correlated with disease duration and severity (SPRS). "Global" brain atrophy in HSP, with cHSP more severely affected than pHSP. No significant correlation between MMSE and degree of atrophy.
Aghakhanyan, 2014 Aghakhanyan, Martinuzzi [118]	Single-center, cross-sectional, no clinico-radiological correlations, DTI-TBSS	pHSP (n = 12), SPG10 (n = 1), SPG3a (n = 1), SPG4 (n = 7), SPG7 + SPG4 (n = 1), SPG5 (n = 2), healthy controls (n = 12)	Widespread FA reductions, increased mean RD, MD, and AD compared to controls. Decreased FA in the bilateral anterior thalamic radiations, corticospinal tracts, corpus callosum with forceps major and minor, inferior and superior longitudinal fasciculi.
Martinuzzi, 2016 Martinuzzi, Montanaro [21]	Multicenter, cross-sectional, clinico-radiological correlations, DTI, MRS	SPG3a (n = 7), SPG4 (n = 32), SPG5 (n = 7), SPG7 (n = 4), SPG10 (n = 3), SPG11 (n = 11), SPG15 (n = 3), SPG31 (n = 1), SPG35 (n = 2), healthy controls (n = 22)	Standard MRI brain: WM alterations in 19/58, "ears of the lynx" sign in 9/58 (and 8/10 of SPG11), cortical atrophy in 25/58, CC abnormality in 15/58. SPRS – FA correlations. MRS: inconclusive.
Uttner, 2007 Uttner, Baumgartner [33]	Single-center, cross-sectional, clinico-radiological correlations, neuropsychological assessments, brain parenchymal fractions	pHSP (n = 20), cHSP (n = 9), not genetically confirmed	BPF – neuropsychological performance correlations. pHSP cohort: BPF correlated only with MMSE. cHSP cohort: BPF correlated only with CPM
Kassubek, 2007 Kassubek, Juengling [116]	Single-center, cross-sectional, no clinico-radiological correlations, DTI, VBM	pHSP (n = 21), cHSP (n = 12), age- and sex-matched controls (n = 30)	GM: moderate regional abnormalities in pHSP & significant pericentral GM changes in cHSP. WM: small regional reductions in WM seen in pHSP group & considerable volume reduction in cHSP, particularly in the CC
Montanaro, 2020 Montanaro, Vavla [25]	Single-center, longitudinal, clinico-radiological correlations, DTI – TBSS, VBM, MRS	HSP (n = 31) – follow-up (n = 23) genetically confirmed, controls (n = 36)	VBM: gray matter reductions in the thalami and right prefrontal cortex. TBSS: widespread WM alterations at baseline with no significant progression on longitudinal assessment. No relevant clinico-radiological correlations identified. MRS: reduced prefrontal NAA/Cr, increased Rt. pre-central Cho/Cr, and increased Lt. pre-central ml/Cr. No significant changes on follow-up.
Muller, 2012 Muller, Unrath [126]	Single-center, cross-sectional, no clinico-radiological correlations, DTI -tractography	pHSP (n = 20), cHSP (n = 12), ALS (n = 20), PLS (n = 20)	Reduced FA in the CST and CC of both pHSP and cHSP patients compared to controls; also seen in the ALS and PLS cohort.
Unrath, 2010 Unrath, Muller [127]	Single-center, cross-sectional, no clinico-radiological correlations, DTI tractography	HSP (n = 24), not genetically confirmed, PLS (n = 25), X-SBMA (n = 20), healthy controls (n = 24)	Reduced FA in the CST, CC, capsula extrema, brain stem and limbic system in HSP with reference to controls. Also seen in PLS patients and to a lesser extent in X-SBMA.
Navas-Sanchez, 2021 Navas-Sánchez, Fernández-Pena [29]	Single-center, cross sectional, clinico-radiological correlations, subcortical segmentation & volumetry, vertex analyses	SPG4 (n = 12), controls (n = 22)	Reduced thalamic volume bilaterally. No hippocampal, putamen, globus pallidus, or caudate volume reductions. Negative correlation between SPRS and thalamic volume. Negative correlation was found between the disease duration and the Rt thalamic volume.
Lin, 2020 Lin, Zheng [22]	Single-center, cross-sectional, clinico-radiological correlations, DTI TBSS, cortical thickness, subcortical segmentation	SPG4 (n = 7), healthy controls (n = 7)	CC volume reductions in SPG4. Reduced cortical thickness in the bilateral superior frontal gyri, inferior precentral cortices, central cortices, and inferior parietal cortices; Lt. inf. frontal triangular cortex, postcentral gyrus, and sup. temporal gyrus; and the Rt. middle cingulate cortex and sup occipital cortex. Cortical thickness of Rt inferior frontal cortex negatively correlated with SPRS. Subcortical gray matter atrophy which correlated with disease duration. Reduced FA & AD, increased MD & RD in CC-splenium and CST.
Garaci, 2014 Garaci, Toschi [122]	Single-center, cross-sectional, no clinico-radiological correlations, DTI TBSS	SPG11 (n = 4), SPG4 (n = 3), healthy controls (n = 16)	SPG11: increased MD and decreased FA in centrum semiovale, frontal WM, PLIC, and CC. SPG4: increased MD and decreased FA in centrum semiovale, PLIC, genu of CC.

(Continued)

Table 2. (Continued).

First author, year of publication	Study design Imaging modality	Study groups	Main conclusions
Chen, 2008 [123]	Single-center, cross-sectional, no clinico-radiological correlations, DTI	SPG11 ($n = 2$), healthy controls ($n = 20$)	Increased MD and decreased FA in CC, cerebellum, and thalamus in SPG11 patients compared with healthy controls
Pan, 2013 Pan, Huang [121]	Single-center, cross-sectional, no clinico-radiological correlations, DTI-tractography	SPG11 ($n = 5$), controls ($n = 10$)	Widespread FA reductions. Tractography: reduced integrity in commissural and association fibers in particular prefrontal and motor portions of the CC.
Schneider-Gold, 2017 Schneider-Gold, Dekomien [119]	Single-center, cross-sectional, no clinico-radiological correlation, GM volumetry, brain parenchymal fractions, DWI, MRS	SPG11 ($n = 2$), healthy controls ($n = 5$)	CC atrophy, BPF: brain atrophy, FA reduction & increased MD in CC. MRS: reduced NAA/Cr and NAA/ml in the CC in one of the two patients
Duning, 2010 Duning, Wamecke [44]	Single-center, cross-sectional, no clinico-radiological correlations, GM volumetry, DTI	SPG4 ($n = 6$), controls ($n = 32$)	Normal brain volumetry. Widespread FA reductions particularly in CST and temporal lobes
Oguz, 2013 [120]	Single-center, cross-sectional, no clinico-radiological correlations, DTI-TBSS	HSP-TCC $n = 4$ – not genetically confirmed, healthy controls ($n = 15$)	MRI brain: TCC in all patients. Mild to moderate cerebral atrophy. T2 hyperintensities in the periventricular white matter. DTI: Significantly lower FA in the corpus callosum, thalami, internal and external capsules, anterior thalamic radiations, bilateral cingulum, fornices, superior longitudinal fasciculi, uncinate fasciculi, sagittal stratum, brain stem, and the cerebellum in patients compared with controls.
Franca, 2012 França, Yasuda [117]	Single-center, cross-sectional, no clinico-radiological correlations, DTI – TBSS, VBM	SPG11 ($n = 5$), controls ($n = 15$)	VBM: significant gray matter atrophy in thalamus, lentiform and caudate nucleus in SPG11 compared with controls TBSS: Reduced FA involving symmetrically subcortical white matter of the temporal and frontal lobes, the cingulate gyrus, cuneus, striatum, corpus callosum and brainstem

in SPG2 [108], in the supraventricular white matter in SPG11 [65] and in pre-central regions in genetically diverse HSP [25]. The basal ganglia were specifically evaluated in four MRS studies. Abnormal basal ganglia lipid profiles were described in SPG54 [50,78,138], while levels of other metabolites in the basal ganglia may be normal [138]. In contrast, another study found normal lipid levels in the basal ganglia in SPG54 [77].

2.4.6. Functional MRI (fMRI)

Despite its advantages to characterize functional alterations and network integrity alterations noninvasively [10], very few studies utilized fMRI in HSP. One resting state study revealed widespread cerebral connectivity alterations [139]. Other fMRI studies implemented a motor task to assess disease-associated cerebral activation profiles [36,140]. One study identified increased cerebral activation [140], and the other found decreased activation [36]. Specific study characteristics of metabolic and functional studies are summarized in Table 3.

3. Conclusions

Our review illustrates that a wide variety of neuroimaging techniques have been used in HSP, often with strikingly inconsistent findings. The literature is dominated by case studies, case series, and studies of very small sample sizes. Unifying conclusions are difficult to draw from the available PET, MRS, and fMRI studies due to the considerable differences in study designs. In contrast, DTI studies show relatively consistent results, highlighting CST and CC degeneration as a relatively consistent feature of HSP. Despite sample limitations and methodological differences, spinal cord atrophy and spinal corticospinal degeneration are also a relatively consistent observation in HSP. Cerebellar involvement, thalamic abnormalities, and preferential corpus callosum atrophy are other commonly detected radiological features.

Clinical parameters of disease severity and disability scales (such as the widely used SPRS) may be of relatively limited value for the precision tracking of accruing disease burden. Functional scales are merely indirect proxies of pathological change in the central nervous system (CNS) and may be less likely to change during the typical duration of a clinical trial than a quantitative biomarker. This is in contrast to neuroimaging, which provides objective, observer-independent, anatomically coded quantitative integrity markers. An additional benefit of neuroimaging is that the vast majority of advanced imaging modalities are noninvasive and require no contrast administration. MR scanners are widely available at most institutions, and the vast majority of suspected HSP patients undergo cerebral and spinal imaging anyway as part of their diagnostic work-up. The inclusion of high-resolution structural and diffusion pulse sequences adds little additional time and data processing can be run on a number of open-source and user-friendly cloud-based applications.

The systematic review of imaging in HSP helps not only to identify stereotyped shortcomings, but also to outline desirable study designs for future studies. As with other low incidence MNDs, collaborative efforts, national repositories, and multi-site studies may help to boost cohort sizes and conduct larger imaging studies. Ideally, several genotypes should be

included in one study in an attempt to delineate genotype-specific imaging traits. Harmonized protocols have been extensively used in other neurodegenerative conditions such as AD, ALS, FTD, HD [143,144]. In other conditions, presymptomatic imaging has revealed considerable structural, metabolic, and functional changes long before symptom manifestations [18,145,146] this also is of considerable interest in HSP [147]. The evaluation of presymptomatic changes may help to clarify emerging biological concepts such as 'motor reserve,' factors leading up to phenoconversion, and the chronology of motor and extra-motor changes. From a practical standpoint, the characterization of presymptomatic changes may help to establish the optimal window for pharmacological intervention in future trials and the development of predictive models with regard to projected symptom onset. Longitudinal radiological changes are poorly characterized in HSP despite their relevance to anatomical propagation patterns, flooring- and ceiling-effects of MR metrics, etc. Longitudinal studies are not merely of academic importance, they are also crucial for the development of monitoring markers for clinical practice and markers to appraise response to therapy in future trials. Multi-timepoint and multi-modal studies with uniform follow-up intervals are ideally suited to characterize progressive changes. While machine-learning (ML) applications based on imaging variables have been extensively tested in other neurological disorders, these have not been implemented in HSP to date [148–153]. Given the long diagnostic journey most patients with HSP face, imaging-based patient categorization into specific subgroups may help to expedite the diagnosis. Cluster analyses based on imaging have also contributed important insights in motor neuron disorders but are yet to be applied to HSP data [154,155]. Another important trend in neuroimaging is the characterization of disease burden in single patients instead of describing group-level signatures, which helps the precision tracking of accruing pathology in individual patients [156,157]. Stereotyped limitations of identified HSP studies include the exclusive focus on cerebral imaging, small sample sizes, the assessment of genetically admixed groups, inclusion of genetically not confirmed patients, lack of correlations with clinical variables, lack of extra-motor assessments (cognition, behavior, apathy, language, extra-pyramidal features, etc.), lack of longitudinal follow-up, and the absence of postmortem validation. Desirable future study features would include multi-site/multicenter projects, harmonized imaging protocols, the genetic screening of each participant, in-depth clinical profiling, characterization of extra-motor features, spinal imaging, and the inclusion of asymptomatic mutation carriers. Clinical assessments accompanying imaging should ideally include a bladder questionnaire, as well as tests for executive function, language, memory, and social cognition. Delineation of radiological changes observed in primary lateral sclerosis (PLS) needs to be clarified as there is a considerable overlap in both clinical and radiological features. Imaging studies of PLS also invariably describe PMC, CST, CC, and BG changes similar to those observed in genetically confirmed HSP [158,159]. PLS studies should, therefore, meticulously screen their participants for known HSP mutations [160].

It is noteworthy that more recent white matter techniques such as neurite orientation dispersion and density imaging

Table 3. Metabolic and functional studies on HSP.

First author, year of publication	Study design Imaging modality	Study groups	Main conclusions
Doi 2014 [78]	Single-centre, cross-sectional, no clinico-radiological correlations, MRI brain, MRS: lipids in thalamus and BG	SPG54 (n = 2) Genetically confirmed No controls	MRI brain: thin corpus callosum, MRS: Increased lipid in basal ganglia
Erichsen 2009 [30]	Single-centre, cross-sectional, clinico-radiological correlations, MRS	SPG4 (n = 8), controls (n = 8)	MRS: Decreased total Cho & decreased Cho/Cr in PMC. Cho/Cr reduction correlated with verbal learning memory performance
Hehr 2007 [60]	Single-centre, longitudinal, no clinico-radiological correlations, MRI brain, FDG-PET-CT, MRS: frontal, parieto-occipital, centrum semiovale, IC	SPG11 (n = 20), MRS (n = 1), FDG-PET-CT (n = 3)	MRI brain: thin CC, WM hyperintensities. PET-CT: progressive cortical and thalamic hypometabolism. MRS: mild frontal NAA reduction
Liguori 2014 [141]	Single-centre, cross-sectional, no clinico-radiological correlations, MRS: CSF and cortex	SPG28 (n = 2), no controls	MRS: no cortical change. Lactate accumulation in ventricular CSF in one patient, not in the other
Lossos 2015 [137]	Single-centre, longitudinal (4-year follow-up), no clinico-radiological correlation, MRI brain, MRS: frontal, parieto-occipital	SPG75 (n = 3), no controls	MRI brain: WM hyperintensities, progressive CC and cerebellar atrophy. MRS: decreased NAA in the frontal WM
Orlen 2009 [65]	Single-centre, cross-sectional, no clinico-radiological correlations, MRI brain, PET-CT, MRS: supraventricular white matter	SPG11 (n = 4), healthy controls (n = 5)	MRI: TCC. MRS (n = 1): reduced NAA/Cr and elevated ml/Cr in supraventricular WM. PET-CT (n = 2): bilateral thalamic and sensorimotor cortex hypometabolism
Roos 2014 [46]	Single-centre, cross-sectional, no clinico-radiological correlations, MRI brain, MRS: occipito-parietal lobe	SPG5A (n = 4), no controls	MRI brain: WM hyperintensities in 1 of 4 patients. MRS (n = 2): Increased ml/Cr in occipito-parietal region
Schuurs-Hoeijmakers, 2012 [50]	Single-centre, cross-sectional, no clinico-radiological correlations, MRI brain, MRI C spine, MRS: basal ganglia and thalamus	SPG54 (n = 11), no controls	MRI brain: thin CC & periventricular WM hyperintensities. MRS (n = 5): abnormal lipid peak in the BG and thalamus. MRI spine (n = 2): syrinx in 2/2
Stromillo 2011 [105]	Single-centre, cross-sectional, no clinico-radiological correlations, MRI brain, MRS: CC, sup. corona radiata, periventricular WM	SPG11 (n = 10), controls (n = 10)	MRI brain: WM hyperintensities & volume loss. MRS: Decreased NAA/Cr in CR and periventricular WM. Cerebral volume and CR NAA/Cr correlates with disability
Svenstrup 2010 [108]	Single-centre, longitudinal, no clinico-radiological correlations, MRI brain, PET, MRS: occipito-frontal WM, occipital GM, centrum semiovale	SPG2 (n = 2)	MRI brain: periventricular WM hyperintensities – no progression longitudinally. MRS: Reduced WM NAA, increased WM Cho and MI in white matter (n = 2) PET CT normal
Estrada-Cuzzano, 2017 Estrada-Cuzzano, Martin [94]	Single-centre, cross-sectional, no clinico-radiological correlations, MRI brain, DAT	SPG78 (n = 5), no controls	MRI: cerebellar atrophy, predominantly vermian. DAT scan: decreased putaminal dopamine transporter density
Terada, 2013 Terada, Kono [130]	Single-centre, cross-sectional, no clinico-radiological correlations, PET-CT	SPG3a (n = 2), one control (n = 1)	Reduced frontal glucose metabolism
Samaranch, 2008 Samaranch, Riverol [66]	Single-centre, cross-sectional, no clinico-radiological correlations, MRI, PET-CT	SPG11 (n = 4), one control (n = 1)	MRI: TCC and WM hyperintensities. FDG-PET: reduced paracentral and thalamic metabolism
Anheim, 2009 Anheim, Lagier-Tourenne [142]	Single-centre, cross-sectional, clinico-radiological correlations, ¹²³ I-iodoflupane SPECT	SPG11 (n = 2), one control (n = 1)	Reduced striatal ligand uptake
Seon Kim, 2013 Kim, Kim [129]	Single-centre, cross-sectional, no clinico-radiological correlations, DAT-SPECT	HSP (n = 9), not genetically confirmed	Normal DAT scan in 5 patients, reduced striatal ligand uptake in 4/9 patients
Criscuolo, 2009 Criscuolo, Filla [104]	Single-centre, cross-sectional, no clinico-radiological correlations, MRI brain, PET CT	SPG5 (n = 6), no controls	MRI brain: WM hyperintensities, PET CT: cerebellar hypometabolism in one patient
Orlacchio, 2005 Orlacchio, Kawarai [32]	Single-centre, cross-sectional, clinico-radiological correlations, MRI brain, SPECT	SPG4 (n = 11)	Frontotemporal hypoperfusion correlated with cognitive impairment.
Scheuer, 2006 Scheuer, Nielsen [135]	Single-centre, cross-sectional, clinico-radiological correlations, rCBF PET CT	SPG4 (n = 13), controls (n = 13)	Increased PMC & SMA rCBF during motor task.
Scheuer, 2005 [35]	Single-centre, cross-sectional, clinico-radiological correlations, PET-CT	SPG4 (n = 18), controls (n = 18)	Decreased left frontotemporal rCBF. No correlations with cognitive impairment.

(Continued)

Table 3. (Continued).

First author, year of publication	Study design Imaging modality	Study groups	Main conclusions
Faber, 2018, Faber, Martinez [27]	Single-centre, cross-sectional, clinico-radiological correlations, DAT SPECT – TcTRODAT	SPG11 (n = 22), healthy controls (n = 19)	Decreased striatal density, inverse correlation with disease duration. Reduced caudal and putaminal tracer uptake. SPRS score DAT correlations
Liao, 2018 Liao, Huang [139]	Single-centre, cross-sectional, clinico-radiological correlations, rsfMRI	SPG4 (n = 12), healthy controls (n = 10)	Decreased low-frequency fluctuation amplitudes in left insular cortex and higher in right precentral and superior frontal gyrus. Increased right PMC values correlated negatively with SPRS scores. Increased functional connectivity between left middle frontal gyrus to the left middle orbitofrontal gyrus.
Koritnik, 2009 Koritnik, Azam [140]	Cross-sectional, single-centre, no clinico-radiological correlations, task fMRI	HSP (n = 12), not genetically confirmed, healthy controls (n = 12)	Increased PMC and posterior parietal cortex activation during finger tapping
Tomberg, 2012 Tomberg, Braschinsky [36]	Cross-sectional, single-centre, clinico-radiological correlations, task fMRI	SPG4 (n = 7), ADHSP not genetically confirmed (n = 2), controls (n = 14)	Activation clusters in HSP were smaller than in controls in the sensorimotor area during motor task, and more pronounced in the cerebellum. The degree of sensorimotor cortex was associated with spasticity.

(NODDI), diffusion kurtosis imaging (DKI), and high angular resolution diffusion imaging (HARDI) have not been applied to larger patient cohorts yet, despite their superior potential to characterize white matter integrity especially with regard to crossing fibers. Furthermore, novel multi-voxel whole-brain spectroscopy sequences that have been extensively used to other MNDs are yet to be applied to HSP [161,162]. Other imaging modalities, which have been extensively utilized in MNDs, such as connectomic approaches, quantitative susceptibility mapping (QSM), motor imagery-based fMRI, and texture analysis, are yet to be implemented in HSP [163–165]. Another stereotyped shortcoming of existing HSP imaging studies is the lack of relevant disease controls such as ALS or PLS, which would help to appraise the specificity of purported radiological signatures to HSP. Anatomically expanded activation patterns observed on task-based fMRI studies need to be interpreted with caution. There is a notion that slowly progressive neurodegenerative conditions and neurological syndromes with initial insult in childhood, such as post-polio myelitis syndrome, may be associated with compensatory processes and structural adaptation, which may be detected by advanced imaging methods [166]. The existence of such adaptive or compensatory processes needs to be clarified in HSP. Contrary to other MNDs, these mechanisms have not been studied in HSP to date.

4. Expert Opinion

Irrespective of the underlying genotype, hereditary spastic paraplegias are associated with significant diagnostic challenges, motor disability, and quality-of-life implications. HSP also exhibit considerable clinical heterogeneity with respect to extra-motor manifestations, progression rates, and age of onset. A unifying feature of HSPs is the lack of disease modifying therapies, scarcity of drug development initiatives, and lack of non-clinical quantitative monitoring markers. Key barriers to the effective management of this low-incidence condition include the lengthy diagnostic journey, limited understanding of the pathophysiology of the genetic subtypes, and a lack of disease modifying therapies.

MRI brain and whole spine are routinely performed in all patients presenting with spastic paraparesis to rule out the relevant disease mimics. While reviewing clinical scans for HSP-associated radiological cues such as thin corpus callosum, 'ears-of-the-lynx sign,' and cord atrophy may be helpful, the limited specificity of these qualitative findings to HSP is very important to recognize. Proposed associations between radiological cues and genotypes, such as thin corpus callosum in SPG11 and SPG4 and the 'ears-of-the-lynx sign' in SPG11 and SPG15 should be treated with caution and MR findings alone should not guide genetic screening. Postmortem studies in HSP are notoriously scarce [6,7] despite their importance in deciphering underlying processes and the preferential involvement of specific anatomical regions.

From all the quantitative imaging modalities explored in the academic setting, diffusion tensor-based white matter imaging yielded the most unifying imaging signature, consistently revealing bilateral corticospinal tract degeneration and corpus callosum changes. There are other important

lessons to consider from the advances in neuroimaging in MNDs [167]. Presymptomatic changes are not only important to characterize for academic reasons but also so that pharmacological intervention could be informedly timed. An additional caveat of existing HSP imaging studies is the limited scope of modalities; studies are either primarily structural, diffusion, metabolic, or functional. Ideally, multimodal studies should be conducted, so the comparative detection sensitivity of the various imaging methods can be juxtaposed. The design of the few longitudinal studies identified may also be improved. Ideally, enrollment into longitudinal studies should be relatively uniform with regard to symptom duration, and only multi-timepoint longitudinal studies are suitable for characterizing linear versus non-linear changes in integrity metrics and exploring possible ceiling and flooring effects. Only high-quality, multimodal, and multi-timepoint studies can adequately characterize natural disease trajectories, anatomical propagation patterns, and progression rates. Ultimately, only robust longitudinal studies can lead to the development of sensitive monitoring markers with practical utility in future clinical trials. In future studies, patients should be carefully stratified by the underlying genotype as admixed HSP cohorts are unsuitable for the development of imaging-based biomarkers. Existing imaging studies on HSP overwhelmingly perform group-level descriptive analyses to describe cohort-associated radiological traits. While this is of interest in the academic setting, future studies should capitalize on recent advances in machine-learning and focus on the categorization of individual patient scans into clinically relevant subgroups.

HSP imaging is an exciting and relatively underexplored frontier of MND research. There are ample methodological lessons to consider from imaging initiative in other neurodegenerative conditions. The implementations of novel imaging sequences (QSM, spinal cord, DKI, NODDI, etc.), pooled data from multiple sites, and meticulous stratification by genotype is likely to advance our understanding of pathophysiology in HSP. Ultimately, advanced neuroimaging techniques have the potential to be developed into viable clinical applications, curtailing the diagnostic delay and serving as noninvasive monitoring markers in future clinical trials.

Funding

Health Research Board (HRB EIA-2017-019 & JPND-Cofund-2-2019-1), the Irish Institute of Clinical Neuroscience (IICN), the Spastic Paraplegia Foundation (SPF), the EU Joint Programme – Neurodegenerative Disease Research (JPND), the Andrew Lydon scholarship, and the Iris O'Brien Foundation.

Declaration of Interest

Professor Bede is supported by the Health Research Board (HRB EIA-2017-019 & JPND-Cofund-2-2019-1), the Irish Institute of Clinical Neuroscience (IICN), the Spastic Paraplegia Foundation (SPF), the EU Joint Programme – Neurodegenerative Disease Research (JPND), the Andrew Lydon scholarship, and the Iris O'Brien Foundation.

The sponsors had no bearing on the opinions expressed herein. The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict

605

610

615

620

625

630

635

640

645

650

Q3

655

660 with the subject matter or materials discussed in the manuscript apart
from those disclosed.

Reviewers disclosure

Peer reviewers on this manuscript have no relevant financial relationships
or otherwise to disclose.

665 ORCID

Peter Bede  <http://orcid.org/0000-0003-0492-4772>

Q5 References

Papers of special note have been highlighted as either of interest (*) or of
considerable interest (***) to readers.

- 670 1. Harding AE. Classification of the hereditary ataxias and paraplegias.
Lancet. 1983;3218334:1151–1155.
- 675 2. Fink JK. Hereditary spastic paraplegia: clinico-pathologic features
and emerging molecular mechanisms. Acta Neuropathol. 2013;126
(3):307–328.
- 680 3. Elsayed LEO, et al. Insights into clinical, genetic, and pathological
aspects of hereditary spastic paraplegias: a comprehensive
overview. Front Mol Biosci. 2021;8:690899.
- 685 4. Boutry M, Morais S, Stevanin G. Update on the genetics of spastic
paraplegias. Curr Neurol Neurosci Rep. 2019;19(4):18.
- 690 5. Shribman S, et al. Hereditary spastic paraplegia: from diagnosis to
emerging therapeutic approaches. Lancet Neurol. 2019;18
(12):1136–1146.
- 695 6. Schwarz GA, Liu CN. Hereditary (familial) spastic paraplegia; further
clinical and pathologic observations. AMA Arch Neurol Psychiatry.
1956;75(2):144–162.
- 700 7. Deluca GC, Ebers GC, Esiri MM. The extent of axonal loss in the long
tracts in hereditary spastic paraplegia. Neuropathol Appl Neurobiol.
2004;30(6):576–584.
- 705 8. Koutsis G, et al. A novel ABCD1 mutation detected by next gen-
eration sequencing in presumed hereditary spastic paraplegia: a
30-year diagnostic delay caused by misleading biochemical
findings. J Neurol Sci. 2015;355(1–2):199–201.
- 710 9. Brugman F, et al. Differentiation of hereditary spastic paraparesis
from primary lateral sclerosis in sporadic adult-onset upper motor
neuron syndromes. Arch Neurol. 2009;66(4):509–514.
- 715 10. Proudfoot M, Bede P, Turner MR. Imaging cerebral activity in
amyotrophic lateral sclerosis. Front Neurol. 2018;9:1148.
- 720 11. Feron M, et al. Extrapyramidal deficits in ALS: a combined biome-
chanical and neuroimaging study. J Neurol. 2018;265(9):2125–2136.
12. Chipika RH, et al. “Switchboard” malfunction in motor neuron
diseases: selective pathology of thalamic nuclei in amyotrophic
lateral sclerosis and primary lateral sclerosis. Neuroimage Clin.
2020;27:102300.
13. Li Hi Shing S, et al. Increased cerebral integrity metrics in polio-
myelitis survivors: putative adaptation to longstanding lower
motor neuron degeneration. J Neurol Sci. 2021;424:117361.
14. Bede P, et al. Brainstem pathology in amyotrophic lateral sclerosis
and primary lateral sclerosis: a longitudinal neuroimaging study.
Neuroimage Clin. 2019;24:102054.
15. Querin G, et al. Biomarkers of spinal and bulbar muscle atrophy
(SBMA): a comprehensive review. Front Neurol. 2018;9:844.
16. Querin G, et al. The spinal and cerebral profile of adult
spinal-muscular atrophy: a multimodal imaging study.
Neuroimage Clin. 2019;21:101618.
17. El Mendili MM, et al. Spinal cord imaging in amyotrophic lateral
sclerosis: historical concepts-novel techniques. Front Neurol.
2019;10:350.
18. Querin G, et al. Presymptomatic spinal cord pathology in c9orf72
mutation carriers: a longitudinal neuroimaging study. Ann Neurol.
2019;86(2):158–167.
19. Bede P, et al. Spinal cord markers in ALS: diagnostic and biomarker
considerations. Amyotroph Lateral Scler. 2012;13(5):407–415.
20. Servelhere KR, et al., Brain damage and gene expression across
hereditary spastic paraplegia subtypes. Mov Disord. 2021;36(7):
1644–1653. 725
- **Quantitative imaging study of HSP genotypes.**
21. Martinuzzi A, et al. Clinical and paraclinical indicators of motor
system impairment in hereditary spastic paraplegia: a pilot study.
PloS one. 2016;11(4):e0153283. PLoS One. 730
- **Qualitative and quantitative radiological observations in
genetically classified HSP patients with clinico-radiological
correlations.**
22. Lin JZ, et al. Cortical damage associated with cognitive and motor
impairment in hereditary spastic paraplegia: evidence of a novel
spast mutation. Front Neurol. 2020; 11:399. •Imaging signatures of
SPG4 with clinico-radiological correlation 735
23. Lindig T, et al., Gray and white matter alterations in hereditary
spastic paraplegia type SPG4 and clinical correlations. J Neurol.
2015;262(8): 1961–1971.
- **A quantitative imaging study of SPG4 exploring clinico-
radiological correlations.** 740
24. Cardozo-Hernández ALC, Rezende TJR, França MC Jr. Hereditary
spastic paraplegia type 11 (SPG11) is associated with obesity and
hypothalamic damage. J Neurol Sci. 2020;416:116982.
25. Montanaro D, et al. Multimodal MRI longitudinal assessment of
white and gray matter in different SPG types of hereditary spastic
paraparesis. Front Neurosci. 2020;14:325. 745
- **A longitudinal imaging study of a relatively large sample (n =
31) of HSP patients, which utilizes computational imaging
techniques and comments on clinico-radiological associations.** 750
26. Faber I, et al. SPG11 mutations cause widespread white matter and
basal ganglia abnormalities, but restricted cortical damage.
Neuroimage Clin. 2018;19:848–857.
- **Brain and spinal cord abnormalities assessed in a large sample
of SPG11 patients (n = 25).** 755
27. Faber I, et al. SPG11-related parkinsonism: clinical profile, molecular
imaging and l-dopa response. Mov Disord. 2018;33(10):1650–1656.
28. Servelhere KR, et al., Spinal cord gray and white matter damage in
different hereditary spastic paraplegia subtypes. AJNR Am
J Neuroradiol. 2021;42(3): 610–615. 760
- **Quantitative imaging study confirming spinal cord atrophy in
multiple HSP genotypes.**
29. Navas-Sánchez FJ, et al., Thalamic atrophy in patients with pure
hereditary spastic paraplegia type 4. J Neurol. 2021;268(7):
2429–2440. 765
- **Volumetric analysis of the basal ganglia identifying thalamic
atrophy in patients with SPG4.**
30. Erichsen AK, et al. Proton magnetic resonance spectroscopy and
cognition in patients with spastin mutations. J Neurol Sci. 2009;277
(1–2):124–129. 770
31. Kassubek J, et al., Brain atrophy in pure and complicated hereditary
spastic paraparesis: a quantitative 3D MRI study. Eur
J Neurol.2006;13(8): 880–886.
- **Brain parenchymal fractions detect cerebral atrophy in HSP
patients compared with healthy controls.** 775
32. Orlacchio A, et al. Clinical and genetic study of a large SPG4 Italian
family. Mov Disord. 2005;20(8):1055–1059.
33. Uttner I, et al. Cognitive performance in pure and complicated
hereditary spastic paraparesis: a neuropsychological and neuroima-
ging study. Neurosci Lett. 2007;419(2):158–161. 780
34. Agosta F, et al., Hereditary spastic paraplegia: beyond clinical phe-
notypes toward a unified pattern of central nervous system
damage. Radiology. 2015;276(1): 207–218.
- **Large computational neuroimaging study (n = 44) demonstrat-
ing cerebral white matter alterations and spinal cord atrophy
in HSP patients compared with controls.** 785
35. Scheuer KH, et al. Reduced regional cerebral blood flow in
SPG4-linked hereditary spastic paraplegia. J Neurol Sci. 2005;235
(1–2):23–32.

- 790 36. Tomberg T, et al. Functional MRI of the cortical sensorimotor system in patients with hereditary spastic paraplegia. *Spinal Cord*. 2012;50(12):885–890.
37. List J, et al. Ascending axonal degeneration of the corticospinal tract in pure hereditary spastic paraplegia: a cross-sectional DTI study. *Brain Sci*. 2019; 9(10). [10.3390/brainsci9100268](https://doi.org/10.3390/brainsci9100268)
- 795 • **A robust quantitative imaging study describing both brain and spinal cord alterations in HSP compared to controls.**
38. Rezende TJ, et al. Multimodal MRI-based study in patients with SPG4 mutations. *PLoS one*. 2015;10(2):e0117666. *PLoS One*.
- 800 • **An elegant neuroradiology study which demonstrating both brain and spinal cord abnormalities in SPG4-associated HSP compared to controls.**
39. Nielsen JE, et al. Autosomal dominant pure spastic paraplegia: a clinical, paraclinical, and genetic study. *J Neurol Neurosurg Psychiatry*. 1998;64(1):61–66.
- 805 40. Warnecke T, et al. A novel splice site mutation in the SPG7 gene causing widespread fiber damage in homozygous and heterozygous subjects. *Mov Disord*. 2010;25(4):413–420.
41. Lan MY, et al. Clinical and genetic analysis of Taiwanese patients with hereditary spastic paraplegia type 5. *Eur J Neurol*. 2015;22(1):211–214.
- 810 42. Liu SG, et al. Clinical and genetic study of SPG6 mutation in a Chinese family with hereditary spastic paraplegia. *J Neurol Sci*. 2008;266(1–2):109–114.
- 815 43. Alber B, **Pernauer M, Schwan A**, et al. Spastin related hereditary spastic paraplegia with dysplastic corpus callosum. *J Neurol Sci*. 2005;236(1–2):9–12.
44. Duning T, et al. Specific pattern of early white-matter changes in pure hereditary spastic paraplegia. *Mov Disord*. 2010;25(12):1986–1992.
- 820 45. Masciullo M, et al. Hereditary spastic paraplegia: novel mutations and expansion of the phenotype variability in SPG56. *Eur J Paediatr Neurol*. 2016;20(3):444–448.
46. Roos P, et al. CYP7B1: novel mutations and magnetic resonance spectroscopy abnormalities in hereditary spastic paraplegia type 5A. *Acta Neurol Scand*. 2014;129(5):330–334.
- 825 47. Sperfeld AD, Baumgartner A, Kassubek J. Magnetic resonance investigation of the upper spinal cord in pure and complicated hereditary spastic paraparesis. *Eur Neurol*. 2005;54(4):181–185.
- **A large study (n = 30) assessing spinal cord atrophy in HSP.**
- 830 48. Krabbe K, et al. MRI of autosomal dominant pure spastic paraplegia. *Neuroradiology*. 1997;39(10):724–727.
49. Warnecke T, et al. A novel form of autosomal recessive hereditary spastic paraplegia caused by a new SPG7 mutation. *Neurology*. 2007;69(4):368–375.
- 835 50. Schuurs-Hoeijmakers JH, et al. Mutations in DDHD2, encoding an intracellular phospholipase A(1), cause a recessive form of complex hereditary spastic paraplegia. *Am J Hum Genet*. 2012;91(6):1073–1081.
51. Hsu SL, et al. Clinical and genetic characterization of hereditary spastic paraplegia type 3A in Taiwan. *Parkinsonism Relat Disord*. 2021;87: 87–91.
- 840 52. Orlacchio A, et al. Late-onset hereditary spastic paraplegia with thin corpus callosum caused by a new SPG3A mutation. *J Neurol*. 2011;258(7):1361–1363.
- 845 53. Mahoney CJ, et al. A novel phenotype of hereditary spastic paraplegia type 7 associated with a compound heterozygous mutation in paraplegin. *Muscle Nerve*. 2020;62(1):E44–e45.
54. Pedrosa JL, et al. SPG7 with parkinsonism responsive to levodopa and dopaminergic deficit. *Parkinsonism Relat Disord* 2018;47: 88–90.
- 850 55. Winner B, et al. Clinical progression and genetic analysis in hereditary spastic paraplegia with thin corpus callosum in spastic gait gene 11 (SPG11). *Arch Neurol*. 2004;61(1):117–121.
56. Hsu SL, et al. Investigating ZFYVE26 mutations in a Taiwanese cohort with hereditary spastic paraplegia. *J Formos Med Assoc*. 2021.
- 855 57. Coarelli G, et al. Novel homozygous GBA2 mutation in a patient with complicated spastic paraplegia. *Clin Neurol Neurosurg*. 2018;168:60–63.
58. Pascual B, et al. “Ears of the Lynx” MRI Sign Is Associated with SPG11 and SPG15 hereditary spastic paraplegia. *AJNR Am J Neuroradiol*. 2019;40(1):199–203. 860
59. Dong Y, et al. Genetic, clinical and neuroimaging profiles of sporadic and autosomal recessive hereditary spastic paraplegia cases in Chinese. *Neurosci Lett*. 2021;761: 136108.
- 865 60. Hehr U, et al. Long-term course and mutational spectrum of spatacsin-linked spastic paraplegia. *Ann Neurol*. 2007;62(6):656–665.
61. Stevanin G, et al. Mutations in SPG11 are frequent in autosomal recessive spastic paraplegia with thin corpus callosum, cognitive decline and lower motor neuron degeneration. *Brain*. 2008;131(3):772–784. 870
62. Dosi C, et al. Neuroimaging patterns in paediatric onset hereditary spastic paraplegias. *J Neurol Sci*. 2021;425:117441.
63. Pensato V, et al. Overlapping phenotypes in complex spastic paraplegias SPG11, SPG15, SPG35 and SPG48. *Brain*. 2014;137(Pt 7):1907–1920. 875
64. Wagner F, et al. Subtle imaging findings aid the diagnosis of adolescent hereditary spastic paraplegia and ataxia. *Clin Neuroradiol*. 2019;29(2):215–221.
65. Orlén H, et al. SPG11 mutations cause Kjellin syndrome, a hereditary spastic paraplegia with thin corpus callosum and central retinal degeneration. *Am J Med Genet B Neuropsychiatr Genet*. 2009;150(7):984–992. 880
66. Samaranch L, et al. SPG11 compound mutations in spastic paraparesis with thin corpus callosum. *Neurology*. 2008;71(5):332–336.
- 885 67. Goizet C, et al. SPG15 is the second most common cause of hereditary spastic paraplegia with thin corpus callosum. *Neurology*. 2009;73(14):1111–1119.
68. Özdemir TR, et al. A case of spastic paraplegia-15 with a novel pathogenic variant in ZFYVE26 gene. *Int J Neurosci*. 2019;129(12):1198–1202. 890
69. Chakrabarty S, et al. Spastizin mutation in hereditary spastic paraplegia with thin corpus callosum. *J Neurol*. 2016;263(10):2130–2132.
- 895 70. Simpson MA, et al. Maspardin is mutated in mast syndrome, a complicated form of hereditary spastic paraplegia associated with dementia. *Am J Hum Genet*. 2003;73(5):1147–1156.
71. Dard R, et al. Mutations in DDHD1, encoding a phospholipase A1, is a novel cause of retinopathy and neurodegeneration with brain iron accumulation. *Eur J Med Genet*. 2017;60(12):639–642.
- 900 72. Hotchkiss L, et al. Novel de novo mutations in KIF1A as a cause of hereditary spastic paraplegia with progressive central nervous system involvement. *J Child Neurol*. 2016;31(9):1114–1119.
73. Landouré G, et al. Hereditary spastic paraplegia type 35 in a family from Mali. *Am J Med Genet A*. 2019;179(7):1122–1125.
- 905 74. Elsaid MF, et al. NT5C2 novel splicing variant expands the phenotypic spectrum of Spastic Paraplegia (SPG45): case report of a new member of thin corpus callosum SPG-subgroup. *BMC Med Genet*. 2017;18(1):33.
75. Nakamura-Shindo K, et al. A novel mutation in the GBA2 gene in a Japanese patient with SPG46: a case report. *eNeurologicalSci*. 2020;19: 100238 910
76. Blumkin L, et al. A new locus (SPG47) maps to 1p13.2-1p12 in an Arabic family with complicated autosomal recessive hereditary spastic paraplegia and thin corpus callosum. *J Neurol Sci*. 2011;305(1–2):67–70. 915
77. Nicitá F, et al. Defining the clinical-genetic and neuroradiological features in SPG54: description of eight additional cases and nine novel DDHD2 variants. *J Neurol*. 2019;266(11):2657–2664.
- 920 78. Doi H, et al. Late-onset spastic ataxia phenotype in a patient with a homozygous DDHD2 mutation. *Sci Rep*. 2014;4(1):7132.
79. Gonzalez M, et al. Mutations in phospholipase DDHD2 cause autosomal recessive hereditary spastic paraplegia (SPG54). *Eur J Hum Genet*. 2013;21(11):1214–1218.
80. Zaidi SA, et al. The “broken wishbone” splenial sign: a diagnostic hallmark for SPG54 spastic ataxia. *J Neurol Sci*. 2019;403:114–116. 925

81. Odake Y, et al. Identification of a novel mutation in ATP13A2 associated with a complicated form of hereditary spastic paraplegia. *Neurol Genet.* 2020;6(5):e514.
- 930 82. Okubo S, et al. Neurological and neuroradiological progression in hereditary spastic paraplegia with a thin corpus callosum. *Acta Neurol Scand.* 2000;102(3):196–199.
83. Finegan E, et al. The clinical and radiological profile of primary lateral sclerosis: a population-based study. *J Neurol.* 2019;266(11):2718–2733.
- 935 84. Finegan E, et al. Evolving diagnostic criteria in primary lateral sclerosis: the clinical and radiological basis of “probable PLS.” *J Neurol Sci.* 2020;417:117052.
85. Bede P, Chipika RH. Commissural fiber degeneration in motor neuron diseases. *Amyotroph Lateral Scler Frontotemporal Degener.* 2020. p. 1–3.
- 940 86. Finegan E, et al. Extra-motor cerebral changes and manifestations in primary lateral sclerosis. *Brain Imaging Behav.* 2021;15(5):2283–2296.
- 945 87. Boelmans K, et al. Diffusion tensor imaging of the corpus callosum differentiates corticobasal syndrome from Parkinson’s disease. *Parkinsonism Relat Disord.* 2010;16(8):498–502.
88. Bogdanova-Mihaylova P, et al. Neurophysiological and ophthalmological findings of SPG7-related spastic ataxia: a phenotype study in an Irish cohort. *J Neurol.* 2021.
- 950 89. Hewamadduma CA, et al. Novel genotype-phenotype and MRI correlations in a large cohort of patients with SPG7 mutations. *Neurol Genet.* 2018;4(6):e279.
90. Klebe S, et al. Spastic paraplegia gene 7 in patients with spasticity and/or optic neuropathy. *Brain.* 2012;135(Pt 10):2980–2993.
- 955 91. Spagnoli C, et al. Long-term follow-up until early adulthood in autosomal dominant, complex SPG30 with a novel KIF1A variant: a case report. *Ital J Pediatr.* 2019;45(1):155.
92. Rattay TW, et al. FAHN/SPG35: a narrow phenotypic spectrum across disease classifications. *Brain.* 2019;142(6):1561–1572.
- 960 93. Bettencourt C, et al. Genotype-phenotype correlations and expansion of the molecular spectrum of AP4M1-related hereditary spastic paraplegia. *Orphanet J Rare Dis.* 2017;12(1):172.
94. Estrada-Cuzcano A, et al. Loss-of-function mutations in the ATP13A2/PARK9 gene cause complicated hereditary spastic paraplegia (SPG78). *Brain.* 2017;140(2):287–305.
- 965 95. Scuderi C, et al. Posterior fossa abnormalities in hereditary spastic paraparesis with spastin mutations. *J Neurol Neurosurg Psychiatry.* 2009;80(4):440–443.
96. Orlacchio A, et al. A new SPG4 mutation in a variant form of spastic paraplegia with congenital arachnoid cysts. *Neurology.* 2004;62(10):1875–1878.
- 970 97. Chipika RH, et al. Cerebellar pathology in motor neuron disease: neuroplasticity and neurodegeneration. *Neural Regen Res.* 2022;17(11):2335–2341.
- 975 98. Bede P, et al. Genotype-associated cerebellar profiles in ALS: focal cerebellar pathology and cerebro-cerebellar connectivity alterations. *J Neurol Neurosurg Psychiatry.* 2021;92(11):1197–1205.
99. McKenna MC, et al. Infratentorial pathology in frontotemporal dementia: cerebellar grey and white matter alterations in FTD phenotypes. *J Neurol.* 2021;268(12):4687–4697.
- 980 100. Finegan E, et al. Cerebellar degeneration in primary lateral sclerosis: an under-recognized facet of PLS. *Amyotroph Lateral Scler Frontotemporal Degener.* 2022. p. 1–12.
101. Hourani R, et al. MR imaging findings in autosomal recessive hereditary spastic paraplegia. *AJNR Am J Neuroradiol.* 2009;30(5):936–940.
- 985 102. Orlacchio A, et al. Silver syndrome variant of hereditary spastic paraplegia: a locus to 4p and allelism with SPG4. *Neurology.* 2008;70(21):1959–1966.
- 990 103. Rubegni A, et al. SPG2 mimicking multiple sclerosis in a family identified using next generation sequencing. *J Neurol Sci.* 2017;375:198–202.
104. Criscuolo C, et al. Two novel CYP7B1 mutations in Italian families with SPG5: a clinical and genetic study. *J Neurol.* 2009;256(8):1252–1257.
- 995 105. Stromillo ML, et al. Structural and metabolic damage in brains of patients with SPG11-related spastic paraplegia as detected by quantitative MRI. *J Neurol.* 2011;258(12):2240–2247.
106. Pedroso JL, et al. Fatty acid 2-hydroxylase deficiency: clinical features and brain iron accumulation. *Neurology.* 2015;84(9):960–961. 1000
107. Ebrahimi-Fakhari D, et al. Clinical and genetic characterization of AP4B1-associated SPG47. *Am J Med Genet A.* 2018;176(2):311–318.
108. Svenstrup K, et al. Hereditary spastic paraplegia caused by the PLP1 ‘rumpshaker mutation.’ *J Neurol Neurosurg Psychiatry.* 2010;81(6):666–672. 1005
109. Vecchia SD, et al. Monoallelic KIF1A-related disorders: a multicenter cross sectional study and systematic literature review. *J Neurol.* 2022;269(1):437–450.
110. Della Vecchia S, et al. Correction to: monoallelic KIF1A-related disorders: a multicenter cross sectional study and systematic literature review. *J Neurol.* 2022;269(1):451. 1010
111. van Gassen KL, et al. Genotype-phenotype correlations in spastic paraplegia type 7: a study in a large Dutch cohort. *Brain.* 2012;135(Pt 10):2994–3004.
112. de Freitas JL, et al. Ophthalmological changes in hereditary spastic paraplegia and other genetic diseases with spastic paraplegia. *J Neurol Sci.* 2020;409:116620. 1015
113. Pfeffer G, et al. Mutations in the SPG7 gene cause chronic progressive external ophthalmoplegia through disordered mitochondrial DNA maintenance. *Brain.* 2014;137(Pt 5):1323–1336. 1020
114. Navas-Sánchez FJ, et al., Tract-specific damage at spinal cord level in pure hereditary spastic paraplegia type 4: a diffusion tensor imaging study. *J Neurol.* 2022;269(6): 3189–3203.
- **A robust computational radiology study highlighting spinal cord and cerebral white matter abnormalities in SPG4.** 1025
115. França MC Jr., et al. Prospective neuroimaging study in hereditary spastic paraplegia with thin corpus callosum. *Mov Disord.* 2007;22(11):1556–1562.
116. Kassubek J, et al., Different regional brain volume loss in pure and complicated hereditary spastic paraparesis: a voxel-based morphometric study. *Amyotroph Lateral Scler.* 2007;8(6): 328–336.
- **A quantitative imaging study of cerebral alterations in a relatively large sample (n = 33) of genetically unclassified HSP patients.** 1030
117. França MC Jr., et al. White and grey matter abnormalities in patients with SPG11 mutations. *J Neurol Neurosurg Psychiatry.* 2012;83(8):828–833. 1035
118. Aghakhanyan G, et al., Brain white matter involvement in hereditary spastic paraplegias: analysis with multiple diffusion tensor indices. *AJNR Am J Neuroradiol.* 2014;35(8): 1533–1538. 1040
- **A robust quantitative radiology study of a large sample (n = 22) of genetically confirmed HSP patients.**
119. Schneider-Gold C, et al. Monozygotic twins with a new compound heterozygous SPG11 mutation and different disease expression. *J Neurol Sci.* 2017;381:265–268. 1045
120. Oğuz KK, et al. Tract-based spatial statistics of diffusion tensor imaging in hereditary spastic paraplegia with thin corpus callosum reveals widespread white matter changes. *Diagn Interv Radiol.* 2013;19(3):181–186.
121. Pan MK, et al. Microstructural integrity of cerebral fiber tracts in hereditary spastic paraparesis with SPG11 mutation. *AJNR Am J Neuroradiol.* 2013;34(5):990–6, s1. 1050
122. Garaci F, et al. Diffusion tensor imaging in SPG11- and SPG4-linked hereditary spastic paraplegia. *Int J Neurosci.* 2014;124(4):261–270.
123. Chen Q, et al. Diffusion tensor imaging of two unrelated Chinese men with hereditary spastic paraplegia associated with thin corpus callosum. *Neurosci Lett.* 2008;441(1):21–24. 1055
124. Cao L, et al. Novel SPG11 mutations in Chinese families with hereditary spastic paraplegia with thin corpus callosum. *Parkinsonism Relat Disord.* 2013;19(3):367–370. 1060
125. Dreha-Kulaczewski S, et al. Cerebral metabolic and structural alterations in hereditary spastic paraplegia with thin corpus callosum assessed by MRS and DTI. *Neuroradiology.* 2006;48(12):893–898.
126. Muller HP, et al., Neuroanatomical patterns of cerebral white matter involvement in different motor neuron diseases as studied by 1065

diffusion tensor imaging analysis. *Amyotroph Lateral Scler.* 2012;13 (3): 254–264.

• **An elegant quantitative imaging study contrasting cerebral white matter abnormalities in HSP to other motor neuron diseases.**

127. Unrath A, et al., Whole brain-based analysis of regional white matter tract alterations in rare motor neuron diseases by diffusion tensor imaging. *Hum Brain Mapp.* 2010;31(11): 1727–1740.

• **An innovative computational radiology study comparing white matter diffusivity abnormalities in HSP with other motor neuron diseases.**

128. Tahedl M, et al. Evaluation and categorisation of individual patients based on white matter profiles: single-patient diffusion data interpretation in neurodegeneration. *J Neurol Sci.* 2021;428:117584.

129. Kim JS, et al. Striatal dopaminergic functioning in patients with sporadic and hereditary spastic paraplegias with parkinsonism. *J Korean Med Sci.* 2013;28(11):1661–1666.

130. Terada T, et al. SPG3A-linked hereditary spastic paraplegia associated with cerebral glucose hypometabolism. *Ann Nucl Med.* 2013;27(3):303–308.

131. Ma J, et al. Novel mutations c.[5121_5122insAG]+[6859C>T] of the SPG11 gene associated with cerebellum hypometabolism in a Chinese case of hereditary spastic paraplegia with thin corpus callosum. *Parkinsonism Relat Disord.* 2014;20(2):256–259.

132. Nielsen JE, et al. Hereditary spastic paraplegia with cerebellar ataxia: a complex phenotype associated with a new SPG4 gene mutation. *Eur J Neurol.* 2004;11(12):817–824.

133. Bede P, et al. Degenerative and regenerative processes in amyotrophic lateral sclerosis: motor reserve, adaptation and putative compensatory changes. *Neural Regen Res.* 2021;16(6):1208–1209.

134. Abidi M, et al. Adaptive functional reorganization in amyotrophic lateral sclerosis: coexisting degenerative and compensatory changes. *Eur J Neurol.* 2020;27(1):121–128.

135. Scheuer KH, et al. Motor activation in SPG4-linked hereditary spastic paraplegia. *J Neurol Sci.* 2006;244(1–2):31–39.

136. Fradakis MJ, et al. Novel compound heterozygous spatacsin mutations in a Greek kindred with hereditary spastic paraplegia spg11 and dementia. *Neurodegener Dis.* 2016;16(5–6):373–381.

137. Lossos A, et al. Myelin-associated glycoprotein gene mutation causes Pelizaeus-Merzbacher disease-like disorder. *Brain.* 2015;138 (Pt 9):2521–2536.

138. Thabet F, Tlili-Graïss K, Tabarki B. Distinct neuroimaging features of DDHD2 gene-related spastic paraplegia, a mimicker of cerebral palsy. *Arch Dis Child.* 2020;105(5):482.

139. Liao X, et al. Resting state fMRI studies in SPG4-linked hereditary spastic paraplegia. *J Neurol Sci.* 2018;384:1–6.

140. Koritnik B, et al. Functional changes of the cortical motor system in hereditary spastic paraparesis. *Acta Neurol Scand.* 2009;120(3):182–190.

141. Liguori R, et al. Impairment of brain and muscle energy metabolism detected by magnetic resonance spectroscopy in hereditary spastic paraparesis type 28 patients with DDHD1 mutations. *J Neurol.* 2014;261(9):1789–1793.

142. Anheim M, et al. SPG11 spastic paraplegia. A new cause of juvenile parkinsonism. *J Neurol.* 2009;256(1):104–108.

143. Muller HP, et al. A large-scale multicentre cerebral diffusion tensor imaging study in amyotrophic lateral sclerosis. *J Neurol Neurosurg Psychiatry.* 2016;87(6):570–579.

144. Holland D, McEvoy LK, Dale AM. Unbiased comparison of sample size estimates from longitudinal structural measures in ADNI. *Hum Brain Mapp.* 2012;33(11):2586–2602.

145. Chipika RH, et al. The presymptomatic phase of amyotrophic lateral sclerosis: are we merely scratching the surface? *J Neurol.* 2021;268 (12):4607–4629.

146. Lulé DE, et al. Deficits in verbal fluency in presymptomatic C9orf72 mutation gene carriers: a developmental disorder. *J Neurol Neurosurg Psychiatry.* 2020;91(11):1195–1200.

147. Rattay TW, et al., The prodromal phase of hereditary spastic paraplegia type 4: the preSPG4 cohort study. *Brain.* 2022.

•• **An innovative and conceptually original study characterising presymptomatic radiological changes in SPG4.**

148. Grollemund V, et al. Development and validation of a 1-year survival prognosis estimation model for amyotrophic lateral sclerosis using manifold learning algorithm UMAP. *Sci Rep.* 2020;10 (1):13378.

149. Grollemund V, et al. Manifold learning for amyotrophic lateral sclerosis functional loss assessment: development and validation of a prognosis model. *J Neurol.* 2021;268(3):825–850.

150. Bede P, Murad A, Hardiman O. Pathological neural networks and artificial neural networks in ALS: diagnostic classification based on pathognomonic neuroimaging features. *J Neurol.* 2021.

151. Schuster C, Hardiman O, Bede P. Development of an automated MRI-based diagnostic protocol for amyotrophic lateral sclerosis using disease-specific pathognomonic features: a quantitative disease-state classification study. *PLoS One.* 2016;11(12): e0167331.

152. Bede P, Chang KM, Tan EL. Machine-learning in motor neuron diseases: prospects and pitfalls. *Eur J Neurol.* 2022;29(9):2555–2556.

153. McKenna MC, et al. The changing landscape of neuroimaging in frontotemporal lobar degeneration: from group-level observations to single-subject data interpretation. *Expert Rev Neurother.* 2022;22 (3):179–207.

154. Bede P, et al. Clusters of anatomical disease-burden patterns in ALS: a data-driven approach confirms radiological subtypes. *J Neurol.* 2022;269(8):4404–4413.

155. Dukic S, et al. Resting-state EEG reveals four subphenotypes of amyotrophic lateral sclerosis. *Brain.* 2022;145(2):621–631.

156. Tahedl M, et al. Cortical progression patterns in individual ALS patients across multiple timepoints: a mosaic-based approach for clinical use. *J Neurol.* 2021;268(5):1913–1926.

157. Tahedl M, et al. Propagation patterns in motor neuron diseases: individual and phenotype-associated disease-burden trajectories across the UMN-LMN spectrum of MNDS. *Neurobiol Aging*2021;109. 78–87.

158. Finegan E, et al. Widespread subcortical grey matter degeneration in primary lateral sclerosis: a multimodal imaging study with genetic profiling. *Neuroimage Clin.* 2019;24:102089.

159. Finegan E, et al. Thalamic, hippocampal and basal ganglia pathology in primary lateral sclerosis and amyotrophic lateral sclerosis: evidence from quantitative imaging data. *Data Brief.* 2020;29. 105115.

160. Bede P, et al. Primary lateral sclerosis: clinical, radiological and molecular features. *Rev Neurol (Paris).* 2021;178(3):196–205.

161. Christidi F, et al. Neurometabolic alterations in motor neuron disease: insights from magnetic resonance spectroscopy. *J Integr Neurosci.* 2022;21(3):87.

162. Govind V, et al. Comprehensive evaluation of corticospinal tract metabolites in amyotrophic lateral sclerosis using whole-brain 1H MR spectroscopy. *PLoS One.* 2012;7(4):e35607.

163. Meier JM, et al. Connectome-based propagation model in amyotrophic lateral sclerosis. *Ann Neurol.* 2020;87(5):725–738.

164. Abidi M, et al. Neural correlates of motor imagery of gait in amyotrophic lateral sclerosis. *J Magn Reson Imaging.* 2021;53 (1):223–233.

165. Conte G, et al. Amyotrophic lateral sclerosis phenotypes significantly differ in terms of magnetic susceptibility properties of the precentral cortex. *Eur Radiol.* 2021;31(7):5272–5280.

166. Li Hi Shing S, et al. Cerebellar remodelling decades after spinal cord insult: neuroplasticity in poliomyelitis survivors. *J Integr Neurosci.* 2021.

167. McKenna MC, et al. frontotemporal pathology in motor neuron disease phenotypes: insights from neuroimaging. *Front Neurol.* 2021; 12. 723450.

1135

1140

1145

1150

1155

1160

1165

1170

1175

1180

1185

1190

1195