

1 Genotype–structure–phenotype 2 correlations define divergent natural history 3 in early-onset spastic paraplegia type 4

4 Julian E. Alecu,^{1,2,3} Luca M. Schierbaum,¹ Amy Tam,¹ Vicente Quiroz,¹ Katerina Bernardi,¹
5 Kathryn Yang,¹ Johanna R. Roller,^{2,3} Marion Döbler-Neumann,⁴ Tim W. Rattay,^{2,3,5} Carelis
6 González-Salazar,⁶ Elena A. Zehr,⁷ Daniils Skorohodovs,¹ Josh Rong,¹ Siofra Carty,¹ Nicole
7 Battaglia,¹ Seungbok Lee,^{8,9} Jangsup Moon,^{8,9} Michelle Christie,^{10,11} Antonina Roll-Mecak,⁷
8 Marcondes C. França Jr,⁶ Rebecca Schüle,¹² Ludger J. Schöls,^{2,3} and Darius Ebrahimi-
9 Fakhari^{1,13}

10 Abstract

11 Hereditary spastic paraplegia type 4 (SPG4), caused by variants in *SPAST*, is the most
12 common form of HSP and exhibits a remarkable phenotypic heterogeneity ranging from
13 late-onset pure presentations to severe, early-onset complex disease. Robust genotype–
14 phenotype correlations and detailed natural history data are lacking, limiting clinical trial
15 readiness.

16 We analyzed 206 patients with genetically confirmed SPG4 enrolled across seven
17 international centers, complemented by high-quality literature-derived cases. Deep
18 phenotyping included standardized motor scales, spasticity ratings, developmental
19 milestones, and patient-reported outcomes. We developed an extended essentiality-
20 mapping framework to classify *SPAST* missense variants by integrating *in silico*
21 pathogenicity predictions, evolutionary constraint, physicochemical residue connectivity,
22 and variant enrichment within the human spastin hexamer structure. Plasma
23 neurofilament light chain (pNfL) using was quantified using Simoa in 26 patients and 101
24 controls.

25 We identified 136 distinct *SPAST* variants, including 10 novel variants. Variant class
26 segregated strongly by inheritance, with *de novo* cases enriched for missense variants and
27 inherited cases showing a variety of variant classes with enrichment for truncating variants.
28 Longitudinal analysis revealed two latent trajectories: a rapidly progressive severe
29 subgroup enriched for *de novo* missense variants, and a biphasic moderate subgroup
30 enriched for inherited truncating variants. Patient stratification integrating spastin
31 essentiality mapping (missense variants affecting essential, neutral, or context-dependent

1 residues) with established genetic modifiers (biallelic pathogenic variants or modifier
2 variants *in trans*) classified patients into predicted severe and moderate subgroups with
3 divergent age at onset and clinical disease progression. The severe subgroup showed early
4 developmental delays, rapid loss of ambulation, and declining quality of life, while the
5 moderate subgroup displayed delayed but accelerating disease progression. pNfL levels
6 were elevated in both subgroups, most pronounced in severe early disease.

7 This study provides the most detailed natural history of SPG4 to date and introduces a
8 biologically informed stratification framework that links variant class and location to
9 divergent clinical trajectories. These data establish clinically meaningful benchmarks and
10 offer a genotype-based framework to improve anticipatory care and optimize trial design
11 for SPG4.

12

13 **Author affiliations:**

14 1 Movement Disorders Program, Department of Neurology and F.M. Kirby Neurobiology
15 Center, Boston Children's Hospital, Harvard Medical School, Boston 02115, MA, USA

16 2 Department of Neurodegenerative Diseases, Hertie-Institute for Clinical Brain Research,
17 and Center of Neurology, University of Tübingen, 72076 Tübingen, Germany

18 3 German Center of Neurodegenerative Diseases (DZNE), 72076 Tübingen, Germany

19 4 Department of Pediatric Neurology, University Children's Hospital, 72076 Tübingen,
20 Germany

21 5 Department of Neurology and Center for Rare Neurological Diseases (ZSNE), University
22 Hospital Schleswig-Holstein (UKSH), Campus Kiel, 24105 Kiel, Germany

23 6 Department of Neurology, University of Campinas (UNICAMP), Campinas, São Paulo,
24 13083-970, Brazil

25 7 Cell Biology and Biophysics Unit, National Institute of Neurological Disorders and Stroke,
26 Bethesda, MD 20892, USA

27 8 Department of Neurology, Seoul National University, Seoul, 03080, South Korea

28 9 Department of Genomic Medicine, Seoul National University, Seoul, 03080, South Korea

29

30 10 Department of Neurology and Rehabilitation Medicine, Texas Scottish Rite Hospital,
31 University of Texas Southwestern Medical Center, Dallas, TX 75219, USA

1 11 Spastic Paraplegia - Centers of Excellence Research Network (SP-CERN), Dallas, TX
2 75219, USA

3 12 Division of Neurodegenerative Diseases and Movement Disorders, Department of
4 Neurology, Heidelberg University Hospital and Faculty of Medicine, 69120 Heidelberg,
5 Germany

6 13 Spastic Paraplegia - Centers of Excellence Research Network (SP-CERN), Boston 02115,
7 MA, USA

8

9 Correspondence: Dr Darius Ebrahimi-Fakhari

10 Movement Disorders Program

11 Department of Neurology, Boston Children's Hospital

12 300 Longwood Avenue, Boston, MA 02215, USA

13 E-mail: darius.ebrahimi-fakhari@childrens.harvard.edu

14

15 **Running title:** Natural history of early-onset SPG4

16 **Keywords:** spastic paraplegia, spg4, natural history, SPAST, genotype-phenotype
17 correlation, clinical trial readiness

18

19 **ORCID iDs**

Julian E. Alecu	0000-0002-1420-9775
Amy Tam	0000-0003-2072-3436
Vicente Quiroz	0009-0005-9212-0943
Katerina Bernardi	0009-0004-3387-7623
Kathryn Yang	0009-0004-6892-5189
Johanna R. Roller	0009-0006-9751-4180
Tim W. Rattay	0000-0002-5456-7761
Carelis González-Salazar	0000-0003-2300-682X
Elena A. Zehr	0000-0002-9768-5792
Nicole Battaglia	0009-0008-9751-4580
Jangsup Moon	0000-0003-1282-4528
Seungbok Lee	0000-0002-6620-6269
Michelle Christie	0000-0002-1329-8008
Antonina Roll-Mecak	0000-0003-2621-7307

Marcondes C. França Jr 0000-0003-0898-2419
Rebecca Schüle 0000-0002-7781-2766
Ludger Schöls 0000-0001-7774-5025
Darius Ebrahimi-Fakhari 0000-0002-0026-4714

1

2 Introduction

3 The hereditary spastic paraplegias (HSP) are an evolving group of over 90 monogenic
4 disorders that primarily affect corticospinal motor neurons¹⁻³ and collectively represent the
5 most common cause of inherited spasticity and associated disability worldwide⁴⁻⁶.

6 Spastic paraplegia type 4 (SPG4, HSP-*SPAST*, OMIM #182601) is the most prevalent HSP
7 subtype across diverse populations, accounting for 22% to 41% of HSP cases⁷⁻¹³.

8 Heterozygous variants in *SPAST* cause SPG4, although a small number of patients with
9 biallelic variants have also been reported. Both autosomal dominant inheritance and *de*
10 *novo* occurrence are observed, with *SPAST* variants accounting for the majority of
11 autosomal dominant^{7,14-16} and approximately one-fifth of all *de novo* HSP cases^{17,18}.

12 *SPAST* encodes the AAA (ATPases Associated with diverse cellular Activities) ATPase
13 spastin. Alternative translation initiation generates two major isoforms with distinct
14 functions¹⁹. M87-spastin, translated from the second start codon and lacking the N-
15 terminal hydrophobic domain^{20,21}, forms homo-hexameric complexes that act as
16 microtubule (MT) severing enzymes²²⁻²⁷, governing the balance of MT depolymerization and
17 regrowth^{28,29}, essential for numerous cellular processes. Both the MT-binding and AAA
18 ATPase domain are indispensable for severing: spastin engages the polyglutamylated C-
19 terminus of β -tubulin³⁰ within the hexamer's pore formed by the AAA ATPase domains, and
20 ATP hydrolysis drives MT lattice disruption^{24,26,27}. Depending on cellular context, severing
21 can lead to microtubule depolymerization or microtubule amplification by generating new
22 seeds for microtubule polymerization and promoting lattice repair^{28,29}. This is essential for
23 cortical neuron development and survival, where intact MT dynamics are prerequisite for
24 the morphogenesis and maintenance of long projecting axons³¹⁻³⁶. By contrast, M1-spastin
25 has broader context-dependent roles, including regulation of endoplasmic reticulum (ER)
26 morphology, ER-organelle contact sites, Ca²⁺ signaling, endosomal trafficking, and lipid-
27 droplet biology³⁷⁻⁴¹. Both isoforms are expressed in brain and spinal cord²⁵, but M1 levels
28 are tightly regulated through restricted translation initiation^{19,20} and targeted degradation⁴²⁻
29 ⁴⁴, resulting in low abundance in the brain²⁵.

30 Clinically, SPG4 presents with remarkable heterogeneity despite classically being
31 considered a rather pure form of HSP⁴⁵. Phenotypes range from a complex infantile-onset

1 neurological syndrome with global developmental delay, seizures, and early spastic
2 tetraparesis, to early-onset lower limb spastic paraparesis with mild or absent
3 developmental impairments, to late-onset pure HSP, all following distinct patterns of
4 disease progression⁴⁶⁻⁵⁰. Despite multiple efforts, robust genotype-phenotype correlations
5 have not been established^{46,48-51}. A notable exception is the severe early-onset phenotype
6 associated with biallelic *SPAST* variants, either a pathogenic allele *in trans* with a non-
7 pathogenic intragenic modifier such as p.(Ser44Leu), or two pathogenic variants^{46,48,50,52,53}.

8 To date, no detailed natural history characterization of SPG4 incorporating both clinician-
9 and patient-reported outcome measures - critical for interventional trials - has been
10 available. This gap poses a major barrier as emerging therapeutic strategies advance
11 toward clinical testing^{44,54-56}. Given the small cohorts typical of rare disease trials,
12 biologically informed patient stratification and precise delineation of natural disease
13 progression are essential to maximize statistical power and detect treatment effects within
14 limited observational periods⁵⁷.

15 Here, we investigate genotype-phenotype correlations and the natural history of SPG4,
16 with a focus on divergent clinical trajectories. To maximize sensitivity for detecting
17 meaningful associations, we concentrated on the disease phase characterized by the
18 greatest phenotypic variability. Prior studies have demonstrated substantial heterogeneity
19 in age at onset and clinical presentation during childhood, adolescence, and early
20 adulthood, including a bimodal distribution of onset with peaks in childhood and the third
21 to fourth decade of life, evidence of prodromal and often subclinical pathological changes
22 in *SPAST* variant carriers preceding clinical self-recognition, and reports of early-onset,
23 severe phenotypes associated with specific variant classes. Accordingly, we enrolled
24 individuals younger than 40 years of age with genetically confirmed SPG4 from seven
25 academic centers across four continents, assembling a cohort of 206 patients with deep
26 clinical phenotyping and outcome measures prioritized for natural history studies and
27 future clinical trial design.

28

29 **Materials and methods**

30 **Study design, cohort and clinical characterization**

31 Patients were enrolled through the Registry and Natural History Study for Early-Onset
32 Hereditary Spastic Paraplegia (NCT04712812; Boston Children's Hospital IRB: P00033016)
33 with data contributed by seven collaborating centers across North America, South
34 America, Europe, and Asia. The study was conducted in accordance with the Declaration

1 of Helsinki and received ethical approval at all participating sites. Inclusion criteria were: (i)
2 a confirmed pathogenic or likely pathogenic *SPAST* variant, (ii) clinical signs of disease on
3 neurological examination, and (iii) age ≤ 40 years. All participants were evaluated by board-
4 certified neurologists with expertise in HSP using standardized examination protocols.
5 Variant pathogenicity was determined according to ACMG criteria.

6 Clinical assessments included the Spastic Paraplegia Rating Scale (SPRS)⁵⁸, SPATAX-
7 EUROSPA disability stage (SPATAX), Gross Motor Function Classification System (GMFCS),
8 and the modified Ashworth Scale to evaluate tone in upper limbs (elbow, wrist, thumb,
9 fingers) and lower limbs (quadriceps, hamstrings, gastrocnemius, soleus). The SPRS
10 spasticity subscore was derived from items 7-10. Developmental quotients and motor
11 milestone data were systematically collected. Patient-reported outcomes included the
12 Caregiver Priorities and Child Health Index of Life with Disabilities (CPCHILD), a validated
13 measure of health-related quality of life in children with neurodevelopmental disabilities.
14 No post hoc score calibration across centers was performed.

15 In addition, a systematic literature review (Table S1) identified 146 previously described
16 patients meeting the same inclusion criteria with high-quality patient-level data. These
17 cases were included exclusively in analyses of genetic spectrum and disease progression
18 using SPRS, SPATAX, and GMFCS. Literature-derived cases were included only if patient-
19 level clinical scores or clearly defined motor milestones were explicitly reported and could
20 be directly mapped to these instruments. Cases with ambiguous, aggregated, or non-
21 comparable data were excluded. Missing data were handled in an analysis-specific
22 manner, with patients contributing only to analyses for which the relevant variables were
23 available. No imputation of missing data was performed.

24 The dataset included 21 individuals carrying biallelic *SPAST* configurations (five from the
25 multicenter cohort and sixteen literature-derived cases), including patients with two
26 pathogenic variants as well as individuals carrying a pathogenic variant *in trans* with the
27 established modifier variants p.(Ser44Leu) or p.(Pro45Gln). These cases were retained in
28 the analyses given the well-established association of such configurations with severe
29 SPG4 phenotypes^{48,50,52,53,59}.

30 **Essentiality mapping of spastin residues**

31 To stratify *SPAST* missense variants by functional relevance, we adapted the Essential3D
32 framework by Iqbal and Brünger et al.^{60,61} which integrates evolutionary conservation,
33 missense constraint, and pathogenic variant enrichment in spatial context to identify
34 functionally essential protein regions.

1 We introduced three extensions: (i) composite pathogenicity scoring by integrating
2 predictions from 14 *in silico* tools for all possible amino acid substitutions per residue, with
3 the standard deviation across substitutions retained as a measure of context dependence;
4 (ii) unified evolutionary conservation scoring from nine conservation metrics across
5 multiple phylogenetic clades; and (iii) residue connectivity analysis using evolutionary
6 coupling and physicochemical interaction networks to validate functional
7 interdependence assumptions and reduce false positives in essential site identification.
8 Three-dimensional burden analysis compared pathogenic variants (ClinVar, HGMD) versus
9 population variants (gnomAD). Analyses were performed using the human wild-type
10 spastin hexamer structure (PDB: 6PEN)²². A detailed description of spastin essentiality
11 mapping is provided in the *Supplementary material, Methods*.

12 Neurofilament light Single-Molecule Array Measurements

13 Plasma neurofilament light (pNfL) levels were measured as previously described⁶². Briefly,
14 plasma samples were obtained by standard venipuncture, and NfL measurements in
15 duplicates using the Simoa HD-X Analyzer platform with the Simoa NF-light V2 Advantage
16 Kit (Quanterix, Cat#104073). Further details are provided in the *Supplementary material,*
17 *Methods*.

18 Statistical analysis

19 A full description of all statistical analyses is provided in the *Supplementary material,*
20 *Methods*.

21

22 Results

23 Demographic characteristics of the prospective cohort

24 A total of 206 patients with SPG4 were enrolled from seven centers across the USA,
25 Germany, Brazil, and South Korea. The cohort included 126 males (61.2%) and 80 females
26 (38.8%). Median age at enrollment was 11.2 years (IQR 19.3), and median age at last
27 follow-up was 12.9 years (IQR 20.6). Longitudinal data were available for 102 patients; 43
28 had ≥ 3 visits and 17 had ≥ 4 visits over a median interval of 15.6 months (IQR 14.4).

29

1 Molecular spectrum of early-onset SPG4

2 Across 206 patients, 136 distinct *SPAST* variants were identified, including ten not
3 previously reported (Table S2). Missense variants predominated (45.3%), followed by
4 deletions (16.1%), nonsense (13.1%), and splicing variants (13.1%) (Figure 1A). One patient
5 carried a pathogenic exon gain variant encompassing duplicated exons 5-16.

6 Inheritance was determined in 183 patients (88.8%): 90 had *de novo* variants (43.9%)
7 confirmed by trio testing, and 93 had inherited variants (44.9%). Parental samples were
8 unavailable for 23 patients, precluding definitive determination of inheritance. Variant
9 distributions differed by inheritance. Inherited cases showed the expected distribution of
10 coding impacts with truncating variants most frequent (nonsense 23.6%, deletions 22.5%,
11 frameshift 13.5%), followed by splicing (20.2%) and missense variants (19.1%). By
12 contrast, *de novo* cases were strongly enriched for missense variants (95.3%), consistent
13 with literature data (93.1% of reported cases, Figure S1).

14 Recurrent missense variants included p.(Arg499His) in 42 patients, p.(Met390Val) in 6
15 patients, p.(Arg499Cys) in 4 patients, and p.(Leu195Val) in 3 patients. Most missense
16 variants (90.2%) localized to the AAA ATPase domain, whereas truncating variants were
17 more evenly distributed (Figure 1B).

18 Five patients carried biallelic *SPAST* variants: Two with intragenic modifier variant
19 p.(Ser44Leu) *in trans* with splicing variants, one patient with the same modifier variant *in*
20 *trans* with a frameshift variant, one with biallelic pathogenic missense variants, and one
21 with frameshift and splicing variants *in trans*.

22

23 Exploring genotype-phenotype associations

24 SPG4 demonstrates substantial phenotypic pleiotropy, posing challenges for trial design
25 and interpretation. Establishing generalizable genotype-phenotype associations is
26 therefore critical for effective patient stratification. Three observations motivated our
27 exploration: 1) *de novo* variants are associated with severe, early-onset, and complex
28 phenotypes^{47,49}, 2) *de novo* cases are strongly enriched for missense variants (confirmed
29 here)⁴⁹, and 3) although missense variants are linked to earlier onset and complex features,
30 prior studies have not consistently associated them with worse motor function⁴⁸.

31 Exploratory longitudinal analysis of the SPRS revealed two distinct longitudinal patterns
32 (Figure 1C). A rapidly progressive subgroup reached SPRS scores exceeding 20 points
33 before age 10 and approached the upper range of observed scores by the second to third
34 decade. A second apparent subgroup showed slower early progression with SPRS scores

1 remaining below 15 before age 10 and typically reaching values of 10-20 by the fourth
2 decade. The rapidly progressing subgroup was enriched for missense variants mostly
3 occurring *de novo*, the more slowly progressing subgroup was enriched for inherited
4 truncating variants, although relevant overlap existed across categories.

5 Among 95 distinct missense variants in the combined prospective and literature-derived
6 dataset, only three were shared between *de novo* and inherited cases, indicating near-
7 complete inheritance-based segregation (Figure 1D). This genetic divide supported the
8 hypothesis that different variants – particularly different missense variants – exert distinct
9 molecular effects on spastin function, potentially driving distinct clinical trajectories.

10 Classifications based solely on variant type or inheritance did not adequately predict
11 severity in this or prior studies^{21,46,48,50} (Figure 1C), prompting development of a biologically
12 informed stratification approach.

13

14 Spastin essentiality mapping

15 To classify *SPAST* missense variants by functional relevance, we adapted the Essential3D
16 framework^{60,61}, which integrates missense constraint, evolutionary conservation, and
17 pathogenic variant enrichment in protein structural context to identify functionally
18 essential residues. We introduced three extensions: First, we replaced single-metric
19 missense tolerance with composite scores from 14 *in silico* pathogenicity prediction tools,
20 generating average pathogenicity scores for all possible substitutions per residue while
21 retaining variability as a context-dependence measure. Second, we integrated nine
22 conservation metrics across phylogenetic depths into unified evolutionary constraint
23 scores. Third, we incorporated residue connectivity analyses using evolutionary coupling
24 and physicochemical interaction networks to validate functional interdependence
25 assumptions of three-dimensional residue neighborhoods. This framework was chosen
26 over other variant classification approaches because it integrates relevant complementary
27 data in the specific three-dimensional context of the protein of interest, making it
28 particularly well-suited for the interrogation of potential genotype-phenotype correlations.

29 Analyses used the human wild-type spastin hexamer structure (PDB: 6PEN)²² spanning
30 parts of the MT-binding and the full AAA domain (residues 318-610) where most missense
31 variants localize. Consensus classification proceeded hierarchically (Figure 1E): Residues
32 with minimal connectivity ($Z \leq -2$) were excluded from 3D analysis; remaining residues
33 were evaluated for pathogenic variant enrichment, then conservation and missense
34 constraint. Residues enriched for pathogenic variants ($OR > 1$, $p_{adj.} < 0.05$) with increased
35 conservation and missense constraint ($Z > 0$) were classified as Essential3D sites.

1 Residues not enriched ($p_{\text{adj.}} \geq 0.05$) or depleted ($\text{OR} < 1$, $p_{\text{adj.}} < 0.05$) of pathogenic variants
2 were classified as neutral sites. Residues enriched for pathogenic variants but not meeting
3 the conservation or missense constraint criteria were further evaluated for increased
4 variability in pathogenicity predictions ($Z > 0$), with high-variability residues classified as
5 context-dependent sites.

6 Of 292 residues, 2.4% were unsuitable due to minimal connectivity (mostly loop residues
7 at hexamer edges). The remainder were classified as neutral (42.3%), Essential3D (29.4%),
8 or context-dependent (25.9%). For the latter, all possible missense variants were classified
9 as either severe ($Z > 0$) or moderate ($Z \leq 0$) based on variant-specific pathogenicity
10 predictions.

11

12 Essential3D sites are enriched for functionally relevant structural 13 features

14 Essential3D residues were significantly enriched for functionally critical structural features
15 (Figure 2A). Using manual annotations from *D. melanogaster* homologue studies^{23,63} and
16 automated annotations derived from residue interaction networks (RIN), Essential3D sites
17 were significantly enriched for pore loop 1 ($\text{OR} = \text{Inf}$ [95%-CI 4.4-Inf], $p_{\text{adj.}} = 1.0\text{e-}3$) and pore
18 loop 2 ($\text{OR} = 18.1$ [95%-CI 2.3-822.9], $p_{\text{adj.}} = 0.011$) residues, mostly corresponding to
19 microtubule-interacting sites ($\text{OR} = \text{Inf}$ [95%-CI 2.3-Inf], $p_{\text{adj.}} = 0.019$). Nucleotide/ligand
20 binding residues were overrepresented (RIN-based: $\text{OR} = 7.0$ [95%-CI 2.5-23.0], $p_{\text{adj.}} = 5.3\text{e-}$
21 4), as were residues of the Walker B motif crucial for ATP hydrolysis^{23,64-66} ($\text{OR} = \text{Inf}$ [95%-CI
22 3.0-Inf], $p_{\text{adj.}} = 9.0\text{e-}3$). Manually annotated nucleotide binding sites ($p_{\text{adj.}} = 0.056$) and
23 Walker A motif residues ($p_{\text{adj.}} = 0.060$) showed nominal but not FDR-corrected significance.
24 Residues at protomer interfaces, presumed to mediate hexamerization, were not enriched
25 among essential sites.

26

27 Recurrent variants at Arg499 exemplify context-dependent essentiality

28 Essentiality mapping revealed that residue 499, the arginine finger crucial for ATP
29 hydrolysis²³ and most commonly affected in our cohort, was classified as context-
30 dependent, while other residues crucial for ATP hydrolysis Lys388 and Glu442 (part of the
31 Walker A and B motifs) were Essential3D sites (Figure 2B). Given substantial clinical
32 heterogeneity reported for Arg499 variants^{50,67-71}, we systematically explored severity
33 subclassifications for all potential substitutions.

1 Variants p.(Arg499Pro), p.(Arg499Leu), and p.(Arg499His) were predicted context-
2 dependent severe, while p.(Arg499Gly), p.(Arg499Cys), and p.(Arg499Ser) were moderate
3 (Figure 2C). Proline substitutions were absent in our cohort and the literature; histidine,
4 leucine, and cysteine substitutions occurred in our cohort, and glycine in prior reports.

5 Longitudinal SPRS analysis revealed complete separation between severity subgroups
6 (Figure 2D). Patients with p.(Arg499Cys) had consistently lower scores, whereas
7 p.(Arg499His) and p.(Arg499Leu) showed higher scores and rapid progression.

8 Given the paucity of individual SPRS or comparable clinical scores in published reports, we
9 analyzed age at symptom onset as an orthogonal and consistently documented variable to
10 confirm these trends. Combined data of 95 patients with Arg499 variants derived from the
11 literature and our cohort revealed significant onset differences across substitutions ($p =$
12 $2.6e-10$). Moderate variants p.(Arg499Gly) (median = 22 years, $p_{adj.} = 0.01$) and
13 p.(Arg499Cys) (median = 13.5 years, $p_{adj.} = 2.9e-10$) showed later onset than p.(Arg499His)
14 (median = 11.5 months) (Figure 2E), with larger onset variability for p.(Arg499Cys) (IQR 28.5
15 years) versus p.(Arg499His) (IQR 9.0 months).

16 Across all missense variants, Essential3D and context-dependent sites were associated
17 with earlier onset than neutral sites, and context-dependent moderate variants with later
18 onset than severe variants ($p < 1e-16$) (Figure 2A).

19

20 Patient stratification integrates essentiality mapping with established 21 genotype-phenotype correlations

22 Next, we stratified the entire combined cohort ($n = 352$: 206 study patients, 146 literature
23 patients) by integrating the novel essentiality classification of missense variants with
24 established genetic factors known to influence SPG4 severity: biallelic pathogenic *SPAST*
25 variants and intragenic modifier variants *in trans*^{48,50,53,59,72}.

26 Two predicted phenotypic subgroups emerged (Figure 2B). The predicted severe subgroup
27 ($n = 134$, 38.1%) included missense variants affecting Essential3D residues, context-
28 dependent severe variants, and biallelic *SPAST* variants including modifiers p.(Ser44Leu)
29 and p.(Pro45Gln). Consistent with established genotype-phenotype correlations in SPG4,
30 individuals carrying biallelic *SPAST* variants were classified within the predicted severe
31 subgroup based on their known association with severe clinical presentations, which in the
32 present cohort closely paralleled the trajectories observed in patients with predicted
33 severe monoallelic missense variants. The predicted moderate subgroup ($n = 218$, 61.9%)

1 comprised missense variants at neutral residues, context-dependent moderate variants,
2 monoallelic truncating/splice variants, and variants with uncertain classification.

3

4 **Quality of life is predicted by essentiality-based stratification**

5 To test whether this stratification captured patient-relevant differences, we analyzed
6 CPCHILD scores (128 observations) from 73 patients. The CPCHILD quantifies disease
7 impact on health-related quality of life as perceived by affected families⁷³.

8 Linear mixed-effects modeling with age-by-subgroup interaction, adjusted for inheritance
9 and sex, revealed significant trajectory differences between subgroups (age×subgroup
10 interaction $p = 0.025$) (Figure 2C). Estimated marginal trend (EMT) contrast analysis
11 revealed significantly different slopes between the two subgroups ($p = 0.026$). To interpret
12 age effects given the significant interaction, separate models were fitted for each
13 subgroup, yielding an annual decline of -0.77 points ($p = 0.027$, $R_m^2 = 0.15$) in the severe
14 subgroup, while the moderate subgroup showed no significant change over time ($p = 0.16$)
15 (Figure 2D).

16 Cross-sectional analysis using estimated marginal means (EMM) demonstrated significant
17 differences at multiple timepoints (Figure 2C). While no difference was evident at age 3
18 years, the severe subgroup scored an estimated 12.7 points lower at age 5 years ($p =$
19 0.023), with differences persisting at ages 10 years (18.3 points, $p = 4.1e-4$) and 20 years
20 (29.3 points, $p = 7.0e-5$). Estimates beyond age 20 should be interpreted cautiously due to
21 sparse data.

22

23 **Clinical phenotypes diverge between subgroups**

24 Firth's penalized logistic regression adjusted for inheritance, sex, and age revealed the
25 severe subgroup had significantly increased odds for complex features extending well
26 beyond "pure" HSP (Figure 2E).

27 Motor dysfunction was more pronounced, with increased odds for walking aid (OR = 19.4
28 [95%-CI 7.7-48.9], $p_{adj.} = 2.0e-8$) and wheelchair dependence (OR = 27.1 [95%-CI 9.9-74.5],
29 $p_{adj.} = 1.8e-8$). Upper limb involvement included spasticity (OR = 611.5 [95%-CI 6.0-Inf], $p_{adj.}$
30 = 0.046) and weakness (OR = 27.1 [95%-CI 3.2-229.6], $p_{adj.} = 0.024$). Secondary
31 musculoskeletal manifestations including contractures of the lower limbs (OR = 6.7 [95%-
32 CI 2.2-20.0], $p_{adj.} = 0.011$), as well as muscle wasting (OR = 6.8 [95%-CI 2.2-21.1], $p_{adj.} =$
33 0.011) were also more frequent. Neonatal/infantile hypotonia (OR = 8.2 [95%-CI 2.4-28.6],

1 $p_{\text{adj.}} = 0.011$) typically progressed to hypertonia (OR = 9.5 [95%-CI 2.6-34.4], $p_{\text{adj.}} = 0.011$).
2 Neurodevelopmental abnormalities included speech delay (OR = 5.2 [95%-CI 1.7-16.2],
3 $p_{\text{adj.}} = 0.037$) and developmental regression (OR = 20.5 [95%-CI 4.5-92.3], $p_{\text{adj.}} = 2.7e-3$).
4 Additional manifestations encompassed urinary incontinence (OR = 4.5 [95%-CI 1.5-13.2],
5 $p_{\text{adj.}} = 0.047$), dysarthria (OR = 33.0 [95%-CI 3.0-364.3], $p_{\text{adj.}} = 0.037$), history of aspiration
6 (OR = Inf [95%-CI 157.5-Inf], $p_{\text{adj.}} = 0.041$), and drooling (OR = 8.9 [95%-CI 1.9-43.3], $p_{\text{adj.}} =$
7 0.046). Additional nominally significant associations not surviving FDR correction included
8 intellectual disability (OR = 6.2, $p = 0.028$), developmental delay (OR = 7.9, $p = 9.6e-3$),
9 dysphagia (OR = 14.7, $p = 0.028$), and non-verbal status (OR = 14.3, $p = 0.018$).

10 The moderate subgroup was more likely to achieve motor milestones, with higher odds for
11 unsupported standing (OR = 14.3 [95%-CI 2.7-100.0], $p_{\text{adj.}} = 0.016$) and walking (OR = 20.0
12 [95%-CI 5.9-100.0], $p_{\text{adj.}} = 4.6e-5$).

13 Despite the conservative statistical approach that penalizes quasi-complete or complete
14 separation yielding non-significant results for these findings, extrapyramidal movement
15 disorders showed a clear pattern of subgroup specificity (Figure 2F). Both dystonia (19.3%)
16 and parkinsonism (hypokinesia: 4.8%, rigidity: 3.6%) were exclusively observed in the
17 severe subgroup.

18

19 **Divergent motor functional progression**

20 We evaluated the progression of functional motor impairment using SPATAX (451
21 observations, 251 patients) and GMFCS (273 observations, 132 patients) with survival and
22 proportional-hazards analyses (Figure 3 & Supplementary Fig. S4).

23 Kaplan-Meier analysis revealed distinct progression patterns. The moderate subgroup
24 declined gradually, losing running (SPATAX ≥ 3) at median 34.7 years and requiring walking
25 aids (SPATAX ≥ 4) at 40 years; the severe subgroup reached these stages at 4 and 5 years,
26 respectively (Figures 4a-b). Wheelchair dependence (SPATAX ≥ 6) occurred in 12.2% of
27 moderate patients during observation versus 97.5% of severe patients at median 7.4 years.
28 GMFCS results were concordant: Assistive mobility devices (\geq III) were required at median
29 8.2 years in the severe subgroup versus 36.8 years in the moderate subgroup (Figure 4D
30 and E).

31 Cox proportional hazards models adjusted for sex and inheritance confirmed relevant
32 subgroup effects (Figure 4G-I). For SPATAX progression to walking aid (≥ 4) and wheelchair
33 dependence (≥ 6), the severe subgroup showed 20.9-fold (HR = 20.89 [95%-CI 10.12-43.12],
34 $p = 2.1e-16$) and 43.5-fold (HR = 43.46 [95%-CI 15.85-119.17], $p = 2.3e-13$) increased

1 hazards. For GMFCS progression to requiring an assistive mobility device (\geq III) and severe
2 mobility limitation (\geq IV), hazard ratios were 17.8-fold (HR = 17.77 [95%-CI 4.85-65.13], $p =$
3 $1.4e-5$) and 27.1-fold (HR = 27.08 [95%-CI 5.39-136.03], $p = 6.2e-5$).

4 Both scales inversely correlated with quality of life: each one-point increase in SPATAX and
5 GMFCS corresponded to 5.9- ($p = 2.7e-4$) and 6.5-point ($p = 1.4e-6$) decreases in CPCHILD
6 (Figure 4C and F).

7

8 Progression of spasticity differs by subgroup

9 To characterize spasticity progression, we analyzed SPRS total scores and spasticity
10 subscores using longitudinal mixed-effects modeling (Figure 5). Generalized additive mixed
11 models revealed distinct age-score relationships between subgroups for both SPRS total
12 and spasticity subscores. The moderate subgroup exhibited non-linearity (EDF = 2.54) for
13 total scores, while the severe subgroup showed linear relationships. Automated breakpoint
14 detection identified an inflection point at 13.7 years (95%-CI 8.9-18.5) for the moderate
15 subgroup, which was incorporated into all subsequent analyses.

16 Linear mixed-effects models with age-by-subgroup interactions revealed significant
17 trajectory differences. For SPRS total scores (444 observations, 271 patients), the
18 interaction was significant ($p = 5.1e-3$) with different pre-breakpoint slopes ($p = 2.5e-3$) but
19 convergent post-breakpoint progression ($p = 0.26$). Subgroup-specific modeling showed
20 the severe subgroup had a consistent annual progression of +0.43 points ($p = 4.7e-5$, $R_m^2 =$
21 0.1), while the moderate subgroup exhibited biphasic progression: minimal pre-breakpoint
22 change ($p = 0.18$) followed by acceleration after 13.7 years (+0.4 points/year, $p = 6.7e-3$, $R_m^2 =$
23 0.14).

24 Cross-sectional differences were substantial, with the severe subgroup scoring an
25 estimated 19.4 points higher at age 3 ($p = 5.9e-13$), 23.1 points at age 7 ($p = 1.3e-26$), and
26 stabilizing around 28.1 points at age 20 ($p = 1.1e-34$).

27 SPRS spasticity subscore analysis (338 observations, 177 patients) confirmed these
28 patterns with significant interaction ($p = 2.8e-8$) and slope differences ($p = 1.6e-8$). Annual
29 progression rates were +0.27 ($p = 1.2e-11$, $R_m^2 = 0.34$) and +0.04 points ($p = 7.8e-3$, $R_m^2 =$
30 0.1) in severe and moderate subgroups, respectively. Although non-linearity testing did not
31 support delayed acceleration for the moderate subgroup's spasticity subscore, limited
32 data may account for this and the overall small slope; applying the 13.7-year breakpoint
33 did not improve fit.

1 Importantly, both measures correlated with declining quality-of-life, with each one-point
2 increase decreasing CPCHILD scores by 0.94 ($p = 4.7e-9$) and 2.66 points ($p = 2.2e-6$) for
3 total and spasticity subscores (Figure 5C and F).

4 Given prominent upper limb involvement in the severe subgroup (Figure 3E), we further
5 examined spasticity in this subgroup using the modified Ashworth Scale (Figure 5G-H).
6 Ordinal, non-linear mixed-effects modeling revealed a temporal gradient in the lower
7 limbs: distal muscles showed higher initial grades (2) than proximal muscles (1/+1), with
8 both reaching grade 3 by age 20 and grade 4 by age 30. Upper-limb spasticity emerged later
9 (grade 1/1+ in the second decade) and accelerated in the third decade, typically reaching
10 grades 2 and 3 at ~25 and ~30 years, respectively.

11

12 Anchor-based estimation of clinically important SPRS change

13 To define clinically meaningful SPRS change thresholds for interventional trials, we
14 estimated minimal clinically important differences (MCID) anchored to the Clinical Global
15 Impression-Improvement (CGI-C) scale using longitudinal data from 81 patients (Figure 5I).
16 We evaluated six anchor categories reflecting varying levels of clinician-reported global
17 change. For clinical improvement, any perceived improvement ($CGI-C \leq 3$) corresponded to
18 an estimated SPRS change of -4.51 points (95%-CI -6.67—-2.34; $p = 1.2e-4$; mean change =
19 -2.1), while minimal improvement ($CGI-C = 3$) required -3.94 points (95%-CI -6.35—-1.53; p
20 = $2.0e-3$; mean change = -1.8). For clinical worsening, any perceived deterioration ($CGI-C \geq$
21 5) corresponded to +2.45 points (95%-CI 0.76-4.14; $p = 6.0e-3$; mean change = +2.3), while
22 minimal worsening ($CGI-C = 5$) yielded +1.56 points but did not reach significance (95%-CI
23 -0.23-3.35; $p = 0.09$; mean change = +2.0).

24

25 Severe subgroup presents with global developmental difficulties

26 Building on cross-sectional findings (Figure 3E), we analyzed motor milestone attainment
27 and developmental quotients longitudinally. Kaplan-Meier analysis of gross motor
28 milestones revealed substantial developmental delays in the severe subgroup (Figure 6A),
29 with crawling achieved by 71.3% at median 14 months and walking with assistance by
30 77.4% at median 24 months. Subgroup comparisons showed significant differences for
31 unsupported sitting (moderate: 100% at median 8 months vs. severe: 87.7% at median 9
32 months, $p = 0.031$, Figure S5), and more pronounced divergence for unsupported standing
33 (moderate: 93.3% at median 14 months vs. severe: 40.1%, $p = 3.2e-8$, Figure 6B) and

1 walking (moderate: 91.3% at median 18 months vs. severe: 33.4%, $p = 1.8e-12$, Figure 6C).
2 The moderate subgroup generally achieved milestones, albeit with modest delays.
3 Generalized additive mixed models for developmental quotients (DQ) suggested deviation
4 from normal trajectories across domains (Figure 6D). Gross motor DQ started low and
5 declined linearly; fine motor, expressive language, and social domains diverged during
6 infancy/early childhood, deviated rapidly until ~8-10 years, then declined more slowly.
7 Interpretation is, however, limited by small sample size.

8

9 Plasma neurofilament light is differentially elevated between subgroups

10 Blood NfL levels are moderately elevated in SPG4 with substantial variability⁷⁴⁻⁷⁶ and limited
11 data for younger patients. We measured plasma NfL (pNfL) using SiMoA in 26 SPG4
12 patients (median age: 10.4 years, IQR: 2.6–21.1; 46.7% female) and 101 age-matched
13 controls (median age: 5.7 years, IQR: 0.6–24.8; 45.6% female), with longitudinal data
14 available for four patients.

15 Generalized additive mixed modeling revealed distinct age-related trajectories (Figure 6E).
16 The severe subgroup showed significant non-linear decrease over time (EDF = 1.89, $p < 1e-$
17 16), the moderate subgroup linear progression (EDF = 1.00, $p = 0.006$), while controls
18 showed minimal variation (EDF = 1.62, $p = 0.18$).

19 Cross-sectional differences were most pronounced at younger ages. At age 3, the severe
20 subgroup had elevated estimated pNfL levels (59.8 pg/mL, 95%-CI 53.7–65.9) compared to
21 the moderate subgroup (22.0 pg/mL, 95%-CI 16.7–27.4, $p = 4.1e-15$) and controls (5.8
22 pg/mL, 95%-CI 4.4–7.3, $p = 9.3e-33$). Differences remained significant through age 10
23 (severe: 28.7 pg/mL [95%-CI 25.1–32.4] vs. moderate: 15.7 pg/mL [95%-CI 12.3–19.1], $p =$
24 1.0e-6) but converged by age 15 ($p = 0.15$). Both subgroups (severe: 15.5 pg/mL [95%-CI
25 12.2–18.9] $p = 2.5e-8$, moderate: 11.1 pg/mL [95%-CI 6.2–16.1], $p = 6.4e-3$) remained
26 elevated compared to controls (3.3 pg/mL [95%-CI 1.3–5.3]) until age 15.

27 Comparison with large, age-matched reference cohorts confirmed the relevance of these
28 elevations^{77,78}. Only 1 of 19 samples (5.3%) in the severe subgroup fell within the 99th
29 percentile of age-matched reference values, compared to 2 of 11 samples (18.2%) in the
30 moderate subgroup.

31 Furthermore, we tested for the association between pNfL levels and SPRS total scores
32 using a linear mixed effects model adjusted for subgroup, age and sex (Figure 6F). While
33 the overall variance in SPRS scores explained by pNfL levels was modest ($R_m^2 = 0.12$), pNfL

1 was significantly associated with SPRS ($p = 0.013$), with each 4.76 pg/mL pNfL increase
2 corresponding to a one-point SPRS increase.

3 These findings indicate a moderate but consistent, subgroup-dependent elevation of pNfL
4 in early disease stages, particularly pronounced in severe disease trajectories, and
5 detectable in the moderate subgroup before overt acceleration of clinical progression.
6 Given the limited sample size and longitudinal coverage, these observations should be
7 interpreted as exploratory.

8

9 Discussion

10 SPG4 is the most common form of HSP and exhibits marked phenotypic pleiotropy.
11 Although historically framed as a late-onset “pure” HSP, accumulating evidence
12 documents severe early-onset presentations with profound neurodevelopmental
13 impairment and multisystem involvement⁴⁷⁻⁵⁰. Prior work has implicated inheritance
14 pattern, genetic anticipation, variant class, and sex as contributors to variability⁴⁷⁻⁴⁹;
15 however, as our exploratory analyses confirm, these factors explain only part of the clinical
16 heterogeneity. Advances in understanding spastin biology, particularly its microtubule-
17 severing function^{23,64} and the distinct roles of the M1 and M87 isoforms¹⁹, have informed
18 emerging therapeutic strategies^{44,54,56,79}. Yet a central challenge remains: as preclinical
19 candidates advance to trials, biologically grounded patient stratification will be essential to
20 detect treatment effects in small cohorts over short follow-up periods typical of trials for
21 rare diseases⁵⁷.

22 We investigated genotype-phenotype correlations in a well-characterized cohort of 206
23 patients with pathogenic *SPAST* variants, complemented by high-quality literature-derived
24 cases, focusing on individuals under 40 years to capture the window of greatest phenotypic
25 variability and to maximize sensitivity for detecting divergent disease trajectories prior to
26 late-stage convergence.

27 Longitudinal analyses revealed two distinct clinical trajectories: a subgroup with rapid early
28 progression enriched for missense and *de novo* variants, and a second subgroup with
29 biphasic progression enriched for inherited and truncating variants. However, considerable
30 overlap between clinical and genetic categories indicates that neither inheritance nor
31 variant class alone fully accounts for disease severity. Notably, missense variants showed
32 near-complete segregation by inheritance, consistent with the hypothesis that *de novo* and
33 inherited variants often confer distinct molecular effects on spastin function - an

1 observation paralleling other neurogenetic disorders in which *de novo* variants are linked to
2 extreme phenotypes and reduced transmission^{2,80-83}.

3 Biologically informed stratification framework

4 To classify missense variants, we adapted the Essential3D framework developed by Iqbal,
5 Brünger, and colleagues^{60,61}. Structural mapping revealed that predicted essential residues
6 were enriched at functionally critical sites, including those involved in ATP hydrolysis and
7 microtubule binding, whereas residues at protomer interfaces - expected to impair
8 hexamerization if altered - were not enriched. Extending on the original framework, we
9 introduced a “context-dependent” category for residues with high substitution-specific
10 variability in predicted pathogenicity. This category is exemplified by recurrent variants at
11 Arg499: predicted severe substitutions (p.Arg499His, p.Arg499Leu) associated with earlier
12 onset and rapid progression, whereas predicted moderate substitutions showed later
13 onset; longitudinal data for p.Arg499Cys supported a milder, more slowly progressive
14 course. Based on these findings, we propose a biologically informed patient stratification
15 that integrates essentiality-based missense variant classification with established
16 genotype-phenotype rules to delineate predicted moderate and severe subgroups. We
17 provide comprehensive essentiality predictions for all possible missense variants within
18 residues 318–610 (Table S3), forming the basis for subsequent trajectory analyses and
19 offering practical applicability to clinical stratification.

20 Divergent disease trajectories

21 The two identified subgroups exhibited different patterns of disease progression across
22 multiple domains. The severe subgroup exhibited early, severe motor dysfunction with
23 extensive neurodevelopmental impairments including delayed or absent attainment of
24 motor milestones, speech delay, and intellectual disability. Only about a third of patients
25 learned to walk independently, with development stagnating followed by developmental
26 regression. Patients progressed rapidly to wheelchair dependence (median 7.4 years) and
27 near-maximum disability by the second to third decade, with spasticity affecting both lower
28 and upper limbs. This severe presentation was associated with consistently lower and
29 deteriorating health-related quality of life as reported by caregivers.

30 The moderate subgroup followed a distinctive biphasic course with inflection during the
31 second to third decade. Initially, patients achieved milestones with only modest delays,
32 and maintained motor function (SPRS < 10), and stable quality of life. However, clinical
33 acceleration during adolescence led to progressive functional decline, with patients
34 typically losing the ability to run in the fourth decade and requiring walking aids at a median
35 age of 40 years.

1 The wide phenotypic spectrum in this subgroup during the initial phase, including
2 subclinical presentations during childhood, likely contributes to under-recognition,
3 delayed diagnosis and bias in retrospectively reported age at onset. Modest but consistent
4 pNfL elevation and slightly delayed milestones in childhood suggest that subclinical
5 disease precedes overt motor impairment by years, consistent with findings from the
6 recent preSPG4 study⁷⁶. Importantly, pNfL is interpreted here as a marker of ongoing axonal
7 injury reflecting disease burden rather than as a validated prognostic or stratification
8 biomarker, given the limited sample size and longitudinal coverage. At the level of
9 directionality, pNfL elevations were concordant with the essentiality-based stratification
10 framework, in that the subgroup predicted to have higher disease burden also exhibited
11 higher pNfL levels in early disease stages. Interpretation of these observations, however,
12 requires consideration of physiological pNfL dynamics during early neurodevelopment.
13 Circulating NfL concentrations are relatively elevated during infancy and early childhood
14 and gradually decline as neurodevelopmental processes stabilize^{77,84,85}. Against this
15 background, the elevated pNfL levels observed in the severe subgroup during early disease
16 stages may reflect disease-related axonal injury superimposed on this developmental
17 baseline. Importantly, the reference trajectories shown in Figure 5E correspond to age-
18 dependent physiological pNfL ranges derived from a large previously published pediatric
19 cohort^{77,84,85}, allowing subgroup trajectories to be interpreted relative to established
20 developmental reference values. Within this context, the observed subgroup differences in
21 early disease stages are consistent with increased axonal injury in the severe trajectory
22 while remaining exploratory given the limited sample size.

23 For comparisons between subgroups several of the reported effect size estimates were
24 large and accompanied by wide or unbounded confidence intervals. This reflects strong
25 subgroup separation, with some features occurring almost exclusively in one subgroup, as
26 well as low event counts for certain outcomes. Under these conditions, inflated effect sizes
27 and wide confidence intervals are expected despite appropriate penalized or adjusted
28 modeling. Accordingly, these estimates should be interpreted as indicators of substantial
29 relative differences between subgroups rather than precise quantitative risk measures.

30 Pathomechanistic considerations

31 Our clinically defined subgroups are broadly consistent with recent experimental results
32 using human neural organoids⁵⁵. In this model, which captures selected aspects of human
33 cortical neuron biology but does not fully recapitulate corticospinal motor neuron identity
34 or circuit-level vulnerability, truncating *SPAST* variants were associated with delayed
35 neurodegeneration compared with a pathogenic missense variant affecting an Essential3D
36 residue. This was accompanied by dysregulation of spastin isoform balance and

1 subsequent excessive activation of histone deacetylase 6 (HDAC6) mediated via the M1
2 isoform. While our clinical data do not allow mechanistic inference, these findings are
3 generally compatible with the hypothesis that certain missense variants may precipitate
4 earlier dysregulation of spastin isoform balance, whereas truncating variants may initially
5 permit partial compensation under haploinsufficient conditions.

6 Limitations

7 This study has several limitations that merit consideration. First, despite being among the
8 largest systematically characterized SPG4 cohorts, the disorder's rarity and marked
9 phenotypic pleiotropy limited statistical power for some subgroup analyses. Future studies
10 incorporating outcome measures that more specifically quantify developmental, bulbar,
11 upper limb, and autonomic involvement – underappreciated features of severe SPG4 – will
12 enable more robust modeling of additional clinically relevant domains.

13 A second limitation relates to the multicenter design of the study, which encompassed
14 seven academic centers across four continents and enabled recruitment of a large, deeply
15 phenotyped cohort. However, despite the use of standardized clinical scales and expert
16 assessment, some inter-site variability in the weighting and interpretation of symptoms
17 and clinical scores cannot be fully excluded in the absence of a harmonized prospective
18 assessment protocol or formal inter-site calibration.

19 Third, the Essential3D framework has several general and gene-specific limitations in the
20 context of SPG4. It relies partly on *in silico* predictions and is constrained by the availability
21 of high-quality structural data, which for spastin currently cover only residues 318–610,
22 precluding assessment of N-terminal variants potentially affecting isoform-specific
23 functions of M1-spastin. In addition, the framework is based on static structural
24 representations and therefore does not capture conformational dynamics. Finally, reliance
25 on variant and population databases introduces potential vulnerability to incompleteness
26 and inaccuracies within these resources.

27 Finally, while exploration of plasma neurofilament light as a biomarker yielded relevant
28 data extending previous findings⁷⁴⁻⁷⁶ to the pediatric population, the small sample size,
29 particularly for longitudinal data, does not allow for any definitive conclusions. The present
30 data are insufficient to establish pNfL as a prognostic or stratification biomarker at the
31 individual patient level, and subgroup-dependent differences should be interpreted
32 cautiously. Validation in larger longitudinal cohorts, ideally incorporating additional
33 biomarkers specifically assessing *SPAST*-associated dysregulation^{86,87}, will be essential.

34 Given the reverse translational approach that underlies the present study, the proposed
35 stratification framework should be viewed as iterative and subject to refinement. As

1 additional genotype–phenotype correlations or modifier loci are identified, future iterations
2 may incorporate genetic configurations beyond biallelic *SPAST* variants and missense
3 variants affecting functionally essential residues.

4

5 Clinical implications

6 Despite these limitations, our study provides important insights. First, it offers a
7 biologically grounded stratification framework, informed by protein structure and variant-
8 specific context, that can be applied prospectively once a pathogenic *SPAST* variant is
9 identified. Importantly, this framework is not intended as a deterministic classification
10 system, but rather as a probabilistic tool that integrates variant class, inheritance pattern,
11 and for missense variants Essential3D-based classification – ideally interpreted in the
12 context of genetic precedence – to support early risk stratification without requiring
13 longitudinal clinical data. From a clinical care perspective, early trajectory prediction may
14 support anticipatory guidance. Patients predicted to follow an early severe trajectory may
15 benefit from closer developmental surveillance, earlier multidisciplinary involvement, and
16 proactive assessment of domains that are frequently under-recognized in SPG4, including
17 upper limb, bulbar, and autonomic involvement. Conversely, patients predicted to follow
18 the moderate biphasic trajectory may initially present with subtle or subclinical features;
19 awareness of a later acceleration may reduce diagnostic delay and inform longitudinal
20 monitoring strategies and timely initiation of supportive interventions, including early
21 physiotherapy, which has demonstrated benefit in HSP⁸⁸. From a clinical trial perspective,
22 the framework provides a biologically grounded approach to cohort enrichment and
23 stratified randomization. Grouping patients according to predicted disease trajectory rather
24 than age or variant class alone may reduce phenotypic heterogeneity, improve statistical
25 power to detect treatment effects over shorter follow-up intervals, and inform the selection
26 of therapeutic windows and outcome measures appropriate to subgroup and disease
27 stage.

28 Moreover, the study delivers the most detailed natural history characterization of SPG4 to
29 date, providing clinically meaningful progression rates, age thresholds across multiple
30 functional domains, and CGI-C-anchored MCID estimates for the SPRS.

31 Together, these data can support genotype-aware clinical care and facilitate future
32 research efforts through enhanced clinical trial design, stratification, and variant
33 interpretation.

34

1 Data availability

2 Anonymized data and the code supporting the findings of this study are available from the
3 corresponding author upon reasonable request.

4

5 Acknowledgements

6 The authors thank the patients and patients' families for supporting research on SPG4. The
7 contributions of the NIH authors (E.A.Z & A.R. -M.) were made as part of their official duties
8 as NIH federal employees, are in compliance with agency policy requirements, and are
9 considered Works of the United States Government. However, the findings and
10 conclusions presented in this paper are those of the authors and do not necessarily reflect
11 the views of the NIH or the U.S. Department of Health and Human.

12

13 Funding

14 Research in the Ebrahimi-Fakhari Laboratory is supported by the National Institute of
15 Neurological Disorders and Stroke (K08NS123552, 1U54NS148312-01), the Spastic
16 Paraplegia Foundation, the CureAP4 Foundation, the Lilly & Blair Foundation, the
17 CureSPG4 Foundation, the New England Epilepsy Foundation, the Dystonia Medical
18 Research Foundation, the Boston Children's Hospital Translational Research Program, the
19 Boston Children's Hospital TIDO Accelerator Award. V.Q. received a fellowship from the
20 International Parkinson and Movement Disorder Society. L.M.S. received a fellowship from
21 the Deutsche Forschungsgemeinschaft (German Research Foundation, 536105452). D.S.
22 was supported by the Artan Foundation of the University of Zürich. A.R. -M. and E.A.Z were
23 supported by the Intramural Research Program of the National Institute of Neurological
24 Disorders and Stroke and the National Heart, Lung and Blood Institute of the National
25 Institutes of Health (NIH). Research in the Schüle Laboratory is supported by the Federal
26 Ministry of Research, Technology and Space (BMFTR) through funding for the TreatHSP
27 network (grant 01GM2209A), by the European Health and Digital Executive Agency (HADEA)
28 through funding for the European Rare Disease Research Alliance (ERDERA) (grant
29 agreement 101156595) and the MSCA Doctoral Network Medicine Made to Measure (grant
30 agreement #101120256), by the Else Kröner-Fresenius-Stiftung through funding for the
31 Clinician Scientist Programme PRECISE.net, the 'Deutsche Forschungsgemeinschaft'
32 (SCHU 2598/6-1) and the National Institute of Neurological Disorders and Stroke (NINDS)

1 and the National Institutes of Health (NIH) under Award Number R01NS072248. Several
2 authors are members of the European Reference Network for Rare Neurological Diseases –
3 Project ID 739510 (J.R.R., T.W.R., R.S., L.S.).

4

5 Competing interests

6 Darius Ebrahimi-Fakhari has served as a consultant to Guidepoint LLC and BlackFinBio
7 Ltd. and received speaker honoraria from the Movement Disorder Society, publishing
8 royalties from Cambridge University Press, and reports research funding through a joint
9 research agreement with Astellas Pharmaceuticals Inc and a clinical trial with Neurocrine.
10 The other authors declare no potential conflicts of interest.

11

12 Supplementary material

13 Supplementary material is available at *Brain* online.

14

15 References

16 1. Shribman S, Reid E, Crosby AH, Houlden H, Warner TT. Hereditary spastic paraplegia:
17 from diagnosis to emerging therapeutic approaches. *Lancet Neurol*. Dec 2019;18(12):1136-1146.
18 doi:10.1016/S1474-4422(19)30235-2

19 2. Alecu JE, Saffari A, Jordan C, Srivastava S, Blackstone C, Ebrahimi-Fakhari D. De novo
20 variants cause complex symptoms in HSP-ATL1 (SPG3A) and uncover genotype-phenotype
21 correlations. *Hum Mol Genet*. Jan 1 2023;32(1):93-103. doi:10.1093/hmg/ddac182

22 3. Alecu JE, Saffari A, Jumo H, *et al*. Novel CAPN1 missense variants in complex hereditary
23 spastic paraplegia with early-onset psychosis. *Ann Clin Transl Neurol*. Apr 2022;9(4):570-576.
24 doi:10.1002/acn3.51531

25 4. Coutinho P, Ruano L, Loureiro JL, *et al*. Hereditary ataxia and spastic paraplegia in
26 Portugal: a population-based prevalence study. *JAMA Neurol*. Jun 2013;70(6):746-55.
27 doi:10.1001/jamaneurol.2013.1707

- 1 5. Erichsen AK, Koht J, Stray-Pedersen A, Abdelnoor M, Tallaksen CM. Prevalence of
2 hereditary ataxia and spastic paraplegia in southeast Norway: a population-based study. *Brain*. Jun
3 2009;132(Pt 6):1577-88. doi:10.1093/brain/awp056
- 4 6. Ruano L, Melo C, Silva MC, Coutinho P. The global epidemiology of hereditary ataxia
5 and spastic paraplegia: a systematic review of prevalence studies. *Neuroepidemiology*.
6 2014;42(3):174-83. doi:10.1159/000358801
- 7 7. Cao Y, Zheng H, Zhu Z, Yao L, Tian W, Cao L. Clinical and Genetic Spectrum in a Large
8 Cohort of Hereditary Spastic Paraplegia. *Mov Disord*. Apr 2024;39(4):651-662.
9 doi:10.1002/mds.29728
- 10 8. Cunha IA, Ribeiro JA, Santos MC. Hereditary spastic paraparesis: The real-world
11 experience from a Neurogenetics outpatient clinic. *Eur J Med Genet*. Mar 2022;65(3):104430.
12 doi:10.1016/j.ejmg.2022.104430
- 13 9. Giordani GM, Diniz F, Fussiger H, *et al*. Clinical and molecular characterization of a large
14 cohort of childhood onset hereditary spastic paraplegias. *Sci Rep*. Nov 15 2021;11(1):22248.
15 doi:10.1038/s41598-021-01635-2
- 16 10. Schule R, Wiethoff S, Martus P, *et al*. Hereditary spastic paraplegia: Clinicogenetic lessons
17 from 608 patients. *Ann Neurol*. Apr 2016;79(4):646-58. doi:10.1002/ana.24611
- 18 11. Yang JO, Yoon JY, Sung DH, *et al*. The emerging genetic diversity of hereditary spastic
19 paraplegia in Korean patients. *Genomics*. Nov 2021;113(6):4136-4148.
20 doi:10.1016/j.ygeno.2021.10.014
- 21 12. Yu W, He J, Liu X, *et al*. Clinical features and genetic spectrum of Chinese patients with
22 hereditary spastic paraplegia: A 14-year study. *Front Genet*. 2023;14:1085442.
23 doi:10.3389/fgene.2023.1085442
- 24 13. Erfanian Omidvar M, Torkamandi S, Rezaei S, *et al*. Genotype-phenotype associations in
25 hereditary spastic paraplegia: a systematic review and meta-analysis on 13,570 patients. *J Neurol*.
26 Jun 2021;268(6):2065-2082. doi:10.1007/s00415-019-09633-1
- 27 14. Dong EL, Wang C, Wu S, *et al*. Clinical spectrum and genetic landscape for hereditary
28 spastic paraplegias in China. *Mol Neurodegener*. Jul 6 2018;13(1):36. doi:10.1186/s13024-018-
29 0269-1

- 1 15. Ishiura H, Takahashi Y, Hayashi T, *et al.* Molecular epidemiology and clinical spectrum
2 of hereditary spastic paraplegia in the Japanese population based on comprehensive mutational
3 analyses. *J Hum Genet.* Mar 2014;59(3):163-72. doi:10.1038/jhg.2013.139
- 4 16. Mereaux JL, Banneau G, Papin M, *et al.* Clinical and genetic spectra of 1550 index patients
5 with hereditary spastic paraplegia. *Brain.* Apr 29 2022;145(3):1029-1037.
6 doi:10.1093/brain/awab386
- 7 17. Boutry M, Morais S, Stevanin G. Update on the Genetics of Spastic Paraplegias. *Curr*
8 *Neurol Neurosci Rep.* Feb 28 2019;19(4):18. doi:10.1007/s11910-019-0930-2
- 9 18. Lo Giudice T, Lombardi F, Santorelli FM, Kawarai T, Orlacchio A. Hereditary spastic
10 paraplegia: clinical-genetic characteristics and evolving molecular mechanisms. *Exp Neurol.* Nov
11 2014;261:518-39. doi:10.1016/j.expneurol.2014.06.011
- 12 19. Ramakrishnan S, Mohan N, Dong Z, Liu M, Qiang L. Unraveling Isoform Complexity:
13 The Roles of M1- and M87-Spastin in Spastic Paraplegia 4 (SPG4). *Mov Disord.* Mar
14 2025;40(3):420-430. doi:10.1002/mds.30072
- 15 20. Claudiani P, Riano E, Errico A, Andolfi G, Rugarli EI. Spastin subcellular localization is
16 regulated through usage of different translation start sites and active export from the nucleus. *Exp*
17 *Cell Res.* Oct 1 2005;309(2):358-69. doi:10.1016/j.yexcr.2005.06.009
- 18 21. Solowska JM, Baas PW. Hereditary spastic paraplegia SPG4: what is known and not
19 known about the disease. *Brain.* Sep 2015;138(Pt 9):2471-84. doi:10.1093/brain/awv178
- 20 22. Han H, Schubert HL, McCullough J, *et al.* Structure of spastin bound to a glutamate-rich
21 peptide implies a hand-over-hand mechanism of substrate translocation. *J Biol Chem.* Jan 10
22 2020;295(2):435-443. doi:10.1074/jbc.AC119.009890
- 23 23. Sandate CR, Szyk A, Zehr EA, Lander GC, Roll-Mecak A. An allosteric network in spastin
24 couples multiple activities required for microtubule severing. *Nat Struct Mol Biol.* Aug
25 2019;26(8):671-678. doi:10.1038/s41594-019-0257-3
- 26 24. Roll-Mecak A, Vale RD. Structural basis of microtubule severing by the hereditary spastic
27 paraplegia protein spastin. *Nature.* Jan 17 2008;451(7176):363-7. doi:10.1038/nature06482

- 1 25. Solowska JM, Morfini G, Falnikar A, *et al.* Quantitative and functional analyses of spastin
2 in the nervous system: implications for hereditary spastic paraplegia. *J Neurosci.* Feb 27
3 2008;28(9):2147-57. doi:10.1523/JNEUROSCI.3159-07.2008
- 4 26. Evans KJ, Gomes ER, Reisenweber SM, Gundersen GG, Lauring BP. Linking axonal
5 degeneration to microtubule remodeling by Spastin-mediated microtubule severing. *J Cell Biol.*
6 Feb 14 2005;168(4):599-606. doi:10.1083/jcb.200409058
- 7 27. Roll-Mecak A, Vale RD. The Drosophila homologue of the hereditary spastic paraplegia
8 protein, spastin, severs and disassembles microtubules. *Curr Biol.* Apr 12 2005;15(7):650-5.
9 doi:10.1016/j.cub.2005.02.029
- 10 28. Kuo YW, Trottier O, Mahamdeh M, Howard J. Spastin is a dual-function enzyme that
11 severs microtubules and promotes their regrowth to increase the number and mass of microtubules.
12 *Proc Natl Acad Sci U S A.* Mar 19 2019;116(12):5533-5541. doi:10.1073/pnas.1818824116
- 13 29. Vemu A, Szczesna E, Zehr EA, *et al.* Severing enzymes amplify microtubule arrays
14 through lattice GTP-tubulin incorporation. *Science.* Aug 24
15 2018;361(6404)doi:10.1126/science.aau1504
- 16 30. Valenstein ML, Roll-Mecak A. Graded Control of Microtubule Severing by Tubulin
17 Glutamylation. *Cell.* Feb 25 2016;164(5):911-21. doi:10.1016/j.cell.2016.01.019
- 18 31. Stone MC, Rao K, Gheres KW, *et al.* Normal spastin gene dosage is specifically required
19 for axon regeneration. *Cell Rep.* Nov 29 2012;2(5):1340-50. doi:10.1016/j.celrep.2012.09.032
- 20 32. Aiken J, Holzbaaur ELF. Spastin locally amplifies microtubule dynamics to pattern the axon
21 for presynaptic cargo delivery. *Curr Biol.* Apr 22 2024;34(8):1687-1704 e8.
22 doi:10.1016/j.cub.2024.03.010
- 23 33. Brill MS, Kleele T, Ruschkies L, *et al.* Branch-Specific Microtubule Destabilization
24 Mediates Axon Branch Loss during Neuromuscular Synapse Elimination. *Neuron.* Nov 23
25 2016;92(4):845-856. doi:10.1016/j.neuron.2016.09.049
- 26 34. Butler R, Wood JD, Landers JA, Cunliffe VT. Genetic and chemical modulation of spastin-
27 dependent axon outgrowth in zebrafish embryos indicates a role for impaired microtubule
28 dynamics in hereditary spastic paraplegia. *Dis Model Mech.* Nov-Dec 2010;3(11-12):743-51.
29 doi:10.1242/dmm.004002

- 1 35. Costa AC, Sousa MM. The Role of Spastin in Axon Biology. *Front Cell Dev Biol.*
2 2022;10:934522. doi:10.3389/fcell.2022.934522
- 3 36. Wood JD, Landers JA, Bingley M, *et al.* The microtubule-severing protein Spastin is
4 essential for axon outgrowth in the zebrafish embryo. *Hum Mol Genet.* Sep 15 2006;15(18):2763-
5 71. doi:10.1093/hmg/ddl212
- 6 37. Arribat Y, Grepper D, Lagarrigue S, Qi T, Cohen S, Amati F. Spastin mutations impair
7 coordination between lipid droplet dispersion and reticulum. *PLoS Genet.* Apr
8 2020;16(4):e1008665. doi:10.1371/journal.pgen.1008665
- 9 38. Chang CL, Weigel AV, Ioannou MS, *et al.* Spastin tethers lipid droplets to peroxisomes
10 and directs fatty acid trafficking through ESCRT-III. *J Cell Biol.* Aug 5 2019;218(8):2583-2599.
11 doi:10.1083/jcb.201902061
- 12 39. Papadopoulos C, Orso G, Mancuso G, *et al.* Spastin binds to lipid droplets and affects lipid
13 metabolism. *PLoS Genet.* Apr 2015;11(4):e1005149. doi:10.1371/journal.pgen.1005149
- 14 40. Park SH, Zhu PP, Parker RL, Blackstone C. Hereditary spastic paraplegia proteins REEP1,
15 spastin, and atlastin-1 coordinate microtubule interactions with the tubular ER network. *J Clin*
16 *Invest.* Apr 2010;120(4):1097-110. doi:10.1172/JCI40979
- 17 41. Rizo T, Gebhardt L, Riedlberger J, *et al.* Store-operated calcium entry is reduced in spastin-
18 linked hereditary spastic paraplegia. *Brain.* Sep 14 2022;145(9):3131-3146.
19 doi:10.1093/brain/awac122
- 20 42. Cai Z, Wu H, Jiang T, *et al.* The Cullin3-Ring E3 ubiquitin ligase complex and USP14
21 regulate spastin-mediated microtubule severing and promotion of neurite outgrowth. *Neural Regen*
22 *Res.* Apr 1 2026;21(4):1641-1651. doi:10.4103/NRR.NRR-D-25-00037
- 23 43. Kang HM, Kim DH, Kim M, *et al.* FBXL17/spastin axis as a novel therapeutic target of
24 hereditary spastic paraplegia. *Cell Biosci.* Jul 22 2022;12(1):110. doi:10.1186/s13578-022-00851-
25 1
- 26 44. Sardina F, Carsetti C, Giorgini L, Fattorini G, Cestra G, Rinaldo C. Cul-4 inhibition rescues
27 spastin levels and reduces defects in hereditary spastic paraplegia models. *Brain.* Oct 3
28 2024;147(10):3534-3546. doi:10.1093/brain/awae095

- 1 45. Harding AE. Classification of the hereditary ataxias and paraplegias. *Lancet*. May 21
2 1983;1(8334):1151-5. doi:10.1016/s0140-6736(83)92879-9
- 3 46. de Bot ST, van den Elzen RT, Mensenkamp AR, *et al*. Hereditary spastic paraplegia due
4 to SPAST mutations in 151 Dutch patients: new clinical aspects and 27 novel mutations. *J Neurol*
5 *Neurosurg Psychiatry*. Oct 2010;81(10):1073-8. doi:10.1136/jnnp.2009.201103
- 6 47. Mo A, Saffari A, Kellner M, *et al*. Early-Onset and Severe Complex Hereditary Spastic
7 Paraplegia Caused by De Novo Variants in SPAST. *Mov Disord*. Dec 2022;37(12):2440-2446.
8 doi:10.1002/mds.29225
- 9 48. Parodi L, Fenu S, Barbier M, *et al*. Spastic paraplegia due to SPAST mutations is modified
10 by the underlying mutation and sex. *Brain*. Dec 1 2018;141(12):3331-3342.
11 doi:10.1093/brain/awy285
- 12 49. Schieving JH, de Bot ST, van de Pol LA, *et al*. De novo SPAST mutations may cause a
13 complex SPG4 phenotype. *Brain*. Jul 1 2019;142(7):e31. doi:10.1093/brain/awz140
- 14 50. Varghaei P, Estiar MA, Ashtiani S, *et al*. Genetic, structural and clinical analysis of spastic
15 paraplegia 4. *Parkinsonism Relat Disord*. May 2022;98:62-69.
16 doi:10.1016/j.parkreldis.2022.03.019
- 17 51. Chelban V, Tucci A, Lynch DS, *et al*. Truncating mutations in SPAST patients are
18 associated with a high rate of psychiatric comorbidities in hereditary spastic paraplegia. *J Neurol*
19 *Neurosurg Psychiatry*. Aug 2017;88(8):681-687. doi:10.1136/jnnp-2017-315796
- 20 52. Degoutin M, Angelini C, Bar C, *et al*. From spastic paraplegia to infantile
21 neurodegenerative disorder: Expanding the phenotypic spectrum associated with biallelic SPAST
22 variants. *Eur J Neurol*. Jan 2025;32(1):e70025. doi:10.1111/ene.70025
- 23 53. Nolasco GA, Roldan M, Jamshidi Y, *et al*. Expanding Hereditary Spastic Paraplegias
24 Limits: Biallelic SPAST Variants in Cerebral Palsy Mimics. *Ann Clin Transl Neurol*. Sep 26
25 2025;doi:10.1002/acn3.70206
- 26 54. Havlicek S, Kohl Z, Mishra HK, *et al*. Gene dosage-dependent rescue of HSP neurite
27 defects in SPG4 patients' neurons. *Hum Mol Genet*. May 15 2014;23(10):2527-41.
28 doi:10.1093/hmg/ddt644

- 1 55. Mohan N, Ramakrishnan S, Sun X, *et al.* Genotype-Phenotype Distinctions in Spastic
2 Paraplegia 4 Reveal HDAC6 as a Therapeutic Target. *bioRxiv.* Jul 18
3 2025;doi:10.1101/2025.07.15.664947
- 4 56. Nakazeki F, Tsuge I, Horie T, *et al.* MiR-33a is a therapeutic target in SPG4-related
5 hereditary spastic paraplegia human neurons. *Clin Sci (Lond).* Feb 28 2019;133(4):583-595.
6 doi:10.1042/CS20180980
- 7 57. Pizzamiglio C, Vernon HJ, Hanna MG, Pitceathly RDS. Designing clinical trials for rare
8 diseases: unique challenges and opportunities. *Nat Rev Methods Primers.* Dec
9 2022;2(1)doi:10.1038/s43586-022-00100-2
- 10 58. Schule R, Holland-Letz T, Klimpe S, *et al.* The Spastic Paraplegia Rating Scale (SPRS): a
11 reliable and valid measure of disease severity. *Neurology.* Aug 8 2006;67(3):430-4.
12 doi:10.1212/01.wnl.0000228242.53336.90
- 13 59. Svenstrup K, Bross P, Koefoed P, *et al.* Sequence variants in SPAST, SPG3A and HSPD1
14 in hereditary spastic paraplegia. *J Neurol Sci.* Sep 15 2009;284(1-2):90-5.
15 doi:10.1016/j.jns.2009.04.024
- 16 60. Iqbal S, Brunger T, Perez-Palma E, *et al.* Delineation of functionally essential protein
17 regions for 242 neurodevelopmental genes. *Brain.* Feb 13 2023;146(2):519-533.
18 doi:10.1093/brain/awac381
- 19 61. Iqbal S, Perez-Palma E, Jespersen JB, *et al.* Comprehensive characterization of amino acid
20 positions in protein structures reveals molecular effect of missense variants. *Proc Natl Acad Sci U*
21 *S A.* Nov 10 2020;117(45):28201-28211. doi:10.1073/pnas.2002660117
- 22 62. Alecu JE, Saffari A, Ziegler M, *et al.* Plasma Neurofilament Light Chain Is Elevated in
23 Adaptor Protein Complex 4-Related Hereditary Spastic Paraplegia. *Mov Disord.* Sep
24 2023;38(9):1742-1750. doi:10.1002/mds.29524
- 25 63. Zehr E, Szyk A, Piszczek G, Szczesna E, Zuo X, Roll-Mecak A. Katanin spiral and ring
26 structures shed light on power stroke for microtubule severing. *Nat Struct Mol Biol.* Sep
27 2017;24(9):717-725. doi:10.1038/nsmb.3448

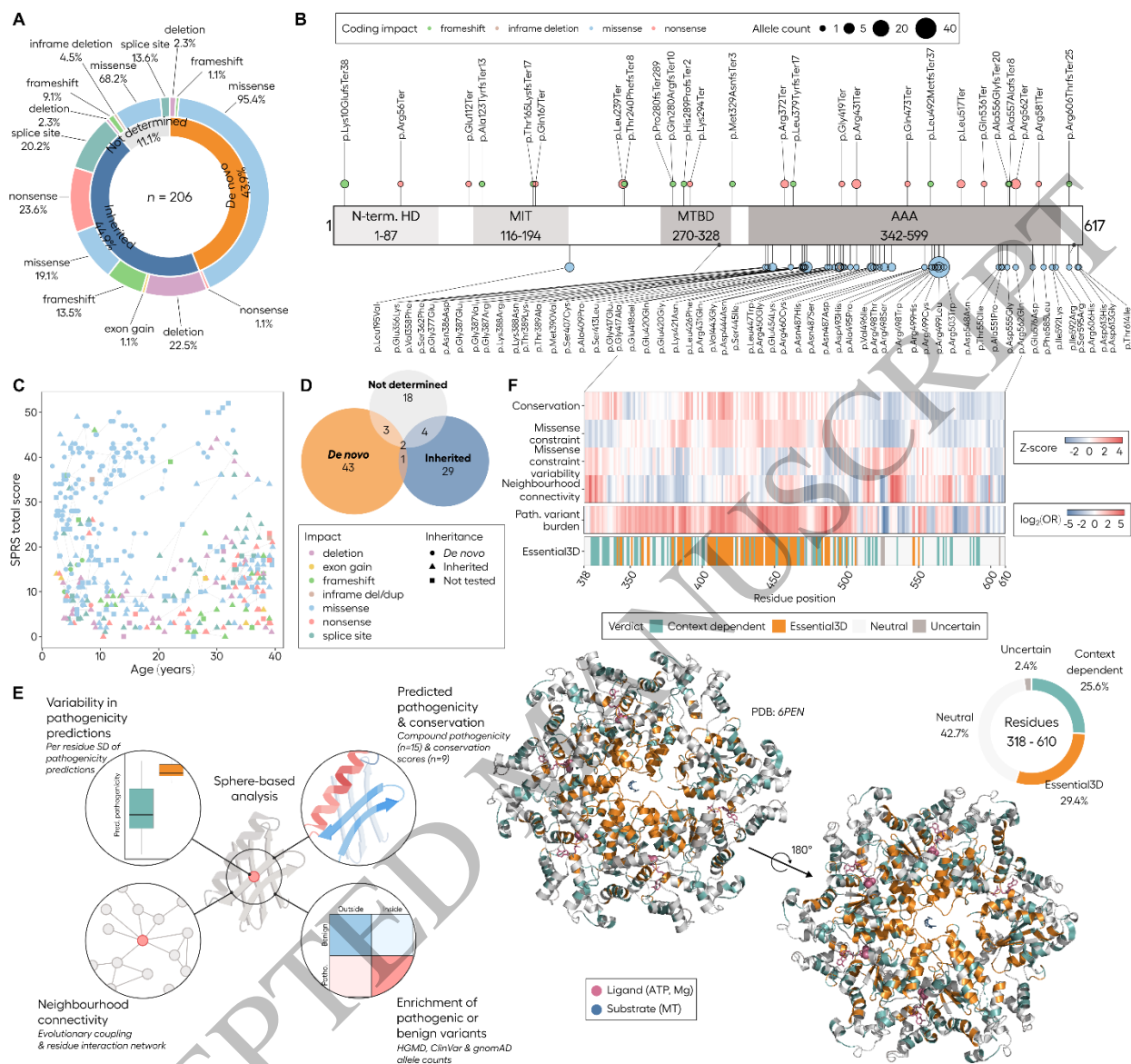
- 1 64. Eckert T, Link S, Le DT, *et al.* Subunit Interactions and cooperativity in the microtubule-
2 severing AAA ATPase spastin. *J Biol Chem.* Jul 27 2012;287(31):26278-90.
3 doi:10.1074/jbc.M111.291898
- 4 65. Fan X, Lin Z, Fan G, *et al.* The AAA protein spastin possesses two levels of basal ATPase
5 activity. *FEBS Lett.* May 2018;592(10):1625-1633. doi:10.1002/1873-3468.13075
- 6 66. Wen M, Wang C. The nucleotide cycle of spastin correlates with its microtubule-binding
7 properties. *FEBS J.* Aug 2013;280(16):3868-77. doi:10.1111/febs.12385
- 8 67. Gillespie MK, Humphreys P, McMillan HJ, Boycott KM. Association of Early-Onset
9 Spasticity and Risk for Cognitive Impairment With Mutations at Amino Acid 499 in SPAST. *J*
10 *Child Neurol.* Apr 2018;33(5):329-332. doi:10.1177/0883073818756680
- 11 68. Nan H, Shiraku H, Mizuno T, Takiyama Y. A p.Arg499His mutation in SPAST is
12 associated with infantile-onset complicated spastic paraplegia: a case report and review of the
13 literature. *BMC Neurol.* Nov 9 2021;21(1):439. doi:10.1186/s12883-021-02478-0
- 14 69. Ogasawara M, Saito T, Koshimizu E, Akasaka N, Sasaki M. A p.Arg499His Mutation in
15 SPAST Is Associated with Infantile Onset Ascending Spastic Paralysis Complicated with
16 Dysarthria and Anarthria. *Neuropediatrics.* Dec 2019;50(6):391-394. doi:10.1055/s-0039-
17 1694973
- 18 70. Ribai P, Depienne C, Fedirko E, *et al.* Mental deficiency in three families with SPG4
19 spastic paraplegia. *Eur J Hum Genet.* Jan 2008;16(1):97-104. doi:10.1038/sj.ejhg.5201922
- 20 71. Xu L, Peng Z, Zhou C, *et al.* A Chinese Patient with Spastic Paraplegia Type 4 with a De
21 Novo Mutation in the SPAST Gene. *Case Rep Genet.* 2021;2021:6636855.
22 doi:10.1155/2021/6636855
- 23 72. Parodi L, Barbier M, Jacoupy M, *et al.* The mitochondrial seryl-tRNA synthetase SARS2
24 modifies onset in spastic paraplegia type 4. *Genet Med.* Nov 2022;24(11):2308-2317.
25 doi:10.1016/j.gim.2022.07.023
- 26 73. Narayanan UG, Fehlings D, Weir S, Knights S, Kiran S, Campbell K. Initial development
27 and validation of the Caregiver Priorities and Child Health Index of Life with Disabilities
28 (CPCHILD). *Dev Med Child Neurol.* Oct 2006;48(10):804-12. doi:10.1017/S0012162206001745

- 1 74. Kessler C, Ruschil C, Abdelhak A, *et al.* Serum Neurofilament Light Chain and Glial
2 Fibrillary Acidic Protein as Biomarkers in Primary Progressive Multiple Sclerosis and Hereditary
3 Spastic Paraplegia Type 4. *Int J Mol Sci.* Nov 3 2022;23(21)doi:10.3390/ijms232113466
- 4 75. Kessler C, Serna-Higuita LM, Wilke C, *et al.* Characteristics of serum neurofilament light
5 chain as a biomarker in hereditary spastic paraplegia type 4. *Ann Clin Transl Neurol.* Mar
6 2022;9(3):326-338. doi:10.1002/acn3.51518
- 7 76. Rattay TW, Volker M, Rautenberg M, *et al.* The prodromal phase of hereditary spastic
8 paraplegia type 4: the preSPG4 cohort study. *Brain.* Mar 1 2023;146(3):1093-1102.
9 doi:10.1093/brain/awac155
- 10 77. Abdelhak A, Petermeier F, Benkert P, *et al.* Serum neurofilament light chain reference
11 database for individual application in paediatric care: a retrospective modelling and validation
12 study. *Lancet Neurol.* Sep 2023;22(9):826-833. doi:10.1016/S1474-4422(23)00210-7
- 13 78. Benkert P, Meier S, Schaedelin S, *et al.* Serum neurofilament light chain for individual
14 prognostication of disease activity in people with multiple sclerosis: a retrospective modelling and
15 validation study. *Lancet Neurol.* Mar 2022;21(3):246-257. doi:10.1016/S1474-4422(22)00009-6
- 16 79. Mohan N, Qiang L, Morfini G, Baas PW. Therapeutic Strategies for Mutant SPAST-Based
17 Hereditary Spastic Paraplegia. *Brain Sci.* Aug 18 2021;11(8)doi:10.3390/brainsci11081081
- 18 80. Acuna-Hidalgo R, Veltman JA, Hoischen A. New insights into the generation and role of
19 de novo mutations in health and disease. *Genome Biol.* Nov 28 2016;17(1):241.
20 doi:10.1186/s13059-016-1110-1
- 21 81. Coe BP, Stessman HAF, Sulovari A, *et al.* Neurodevelopmental disease genes implicated
22 by de novo mutation and copy number variation morbidity. *Nat Genet.* Jan 2019;51(1):106-116.
23 doi:10.1038/s41588-018-0288-4
- 24 82. Erickson RP. The importance of de novo mutations for pediatric neurological disease--It is
25 not all in utero or birth trauma. *Mutat Res Rev Mutat Res.* Jan-Mar 2016;767:42-58.
26 doi:10.1016/j.mrrev.2015.12.002
- 27 83. Wilfert AB, Sulovari A, Turner TN, Coe BP, Eichler EE. Recurrent de novo mutations in
28 neurodevelopmental disorders: properties and clinical implications. *Genome Med.* Nov 27
29 2017;9(1):101. doi:10.1186/s13073-017-0498-x

- 1 84. Geis T, Gutzeit S, Fouzas S, *et al.* Serum Neurofilament light chain (NfL) levels in children
 2 with and without neurologic diseases. *Eur J Paediatr Neurol.* Jul 2023;45:9-13.
 3 doi:10.1016/j.ejpn.2023.05.003
- 4 85. Stukas S, Cooper J, Higgins V, Holmes D, Adeli K, Wellington CL. Pediatric reference
 5 intervals for serum neurofilament light and glial fibrillary acidic protein using the Canadian
 6 Laboratory Initiative on Pediatric Reference Intervals (CALIPER) cohort. *Clin Chem Lab Med.*
 7 Mar 25 2024;62(4):698-705. doi:10.1515/cclm-2023-0660
- 8 86. Sardina F, Valente D, Fattorini G, *et al.* New cellular imaging-based method to distinguish
 9 the SPG4 subtype of hereditary spastic paraplegia. *Eur J Neurol.* Jun 2023;30(6):1734-1744.
 10 doi:10.1111/ene.15756
- 11 87. Wali G, Siow SF, Liyanage E, Kumar KR, Mackay-Sim A, Sue CM. Reduced acetylated
 12 alpha-tubulin in SPAST hereditary spastic paraplegia patient PBMCs. *Front Neurosci.*
 13 2023;17:1073516. doi:10.3389/fnins.2023.1073516
- 14 88. Di Ludovico A, Ciarelli F, La Bella S, Scorrano G, Chiarelli F, Farello G. The therapeutic
 15 effects of physical treatment for patients with hereditary spastic paraplegia: a narrative review.
 16 *Front Neurol.* 2023;14:1292527. doi:10.3389/fneur.2023.1292527
 17

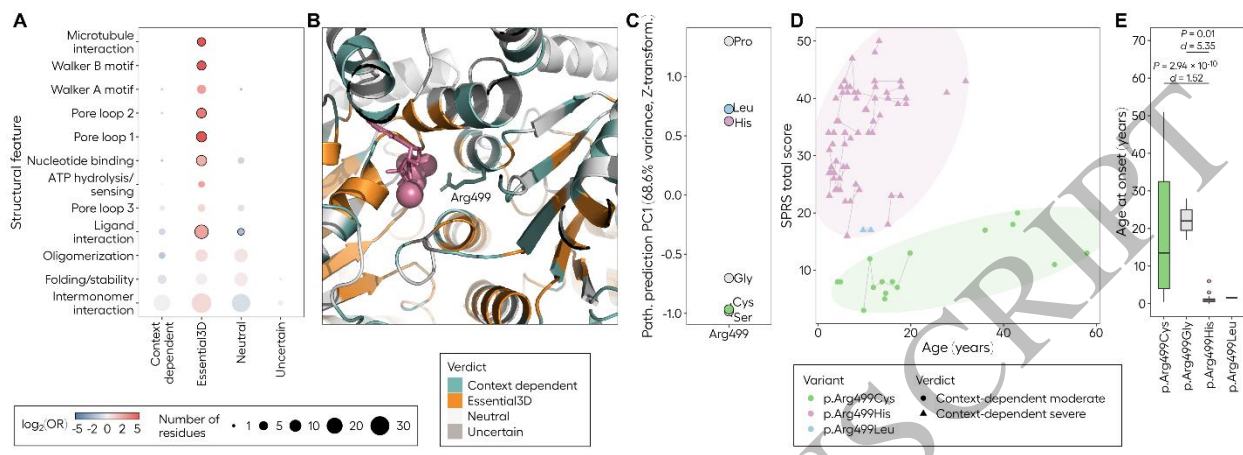
18 Figure legends

19 **Figure 1 Genetic spectrum and essentiality mapping of spastin residues.** (A) Genetic
 20 spectrum of the cohort with inner track representing proportions of inheritance patterns
 21 and outer track representing coding impacts, with proportions nested within the respective
 22 mode of inheritance. (B) Pathogenic variants identified in the cohort, annotated along the
 23 M1-spastin primary structure. Protein-truncating variants are depicted toward the top and
 24 nontruncating variants downward. (C) SPRS scores in the combined literature- and study-
 25 derived dataset. Longitudinal observations from the same patient are connected by lines.
 26 (D) Venn diagram depicting overlaps of distinct missense variants according to inheritance
 27 pattern in the combined dataset. (E) Schematic of the modified Essential3D classification
 28 approach. (F) *Top:* Heatmap depicting variables used for essentiality mapping, including
 29 distribution of essentiality categories. *Bottom:* human wildtype spastin hexamer structure
 30 (PDB: 6PEN) with residues colored by essentiality verdict.

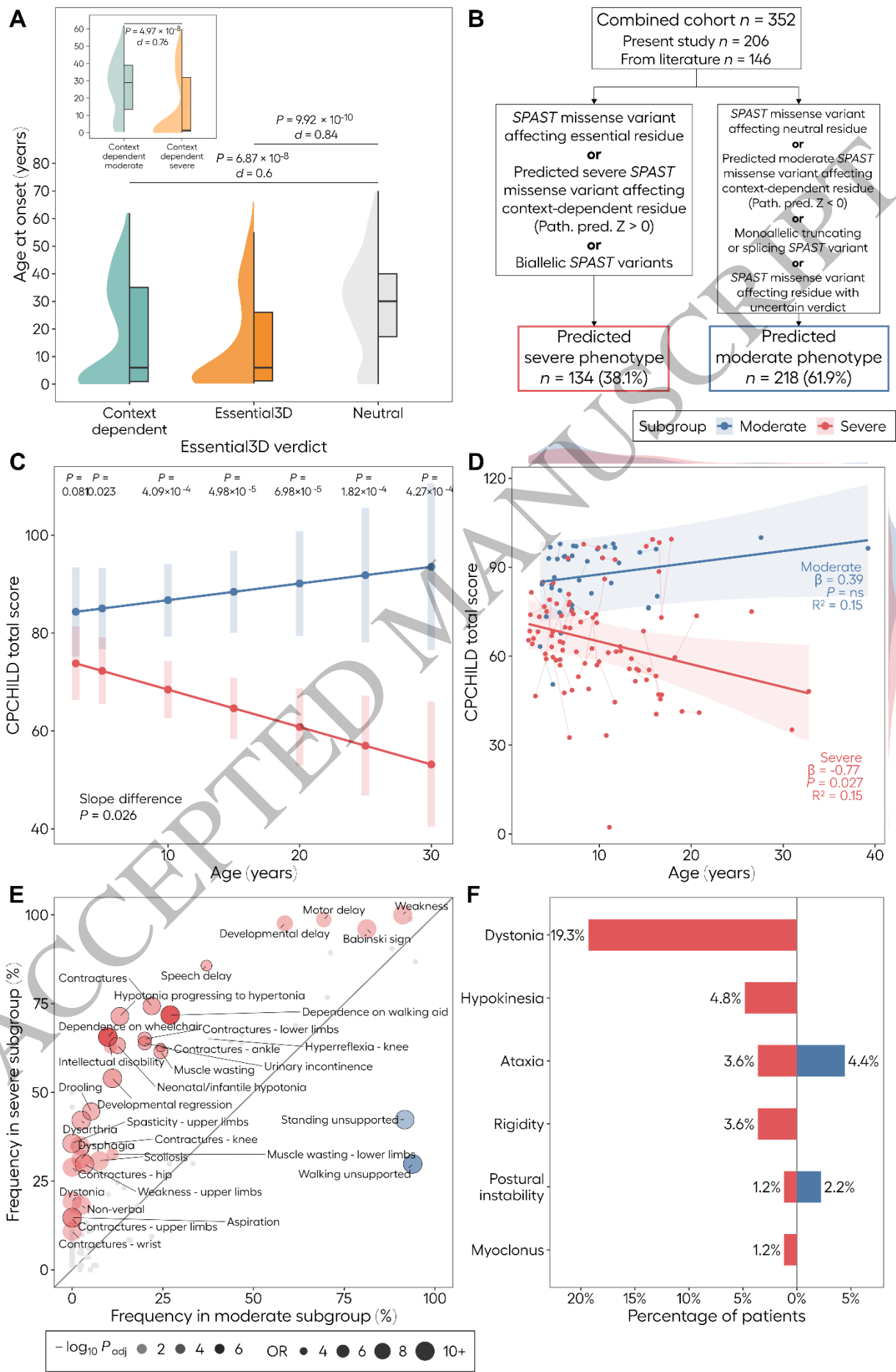


1
 2 **Figure 2 Arg499 exemplifies context-dependent essentiality.** (A) Enrichment analysis of
 3 structural and functional features across essentiality verdicts. Black circles surrounding
 4 dots indicate $p_{adj.} < 0.05$. (B) Zoom-in view on the Arg499 residue neighborhood with
 5 residues colored according to essentiality verdict. Ligands (ATP, BeF, and Mg^{2+}) shown in
 6 purple. (C) Pathogenicity predictions for Arg499 amino acid substitutions. For
 7 subclassification into moderate and severe variants, pathogenicity scores were Z-
 8 transformed for each residue separately. (D) SPRS scores in patients with Arg499 variants
 9 in the combined dataset. Longitudinal observations from the same patient are connected
 10 by lines. (E) Age at onset comparison between Arg499 missense variants. Kruskal-Wallis, p
 11 = $2.6e-10$; p values of pairwise comparisons shown, using Dunn's post-hoc test; d

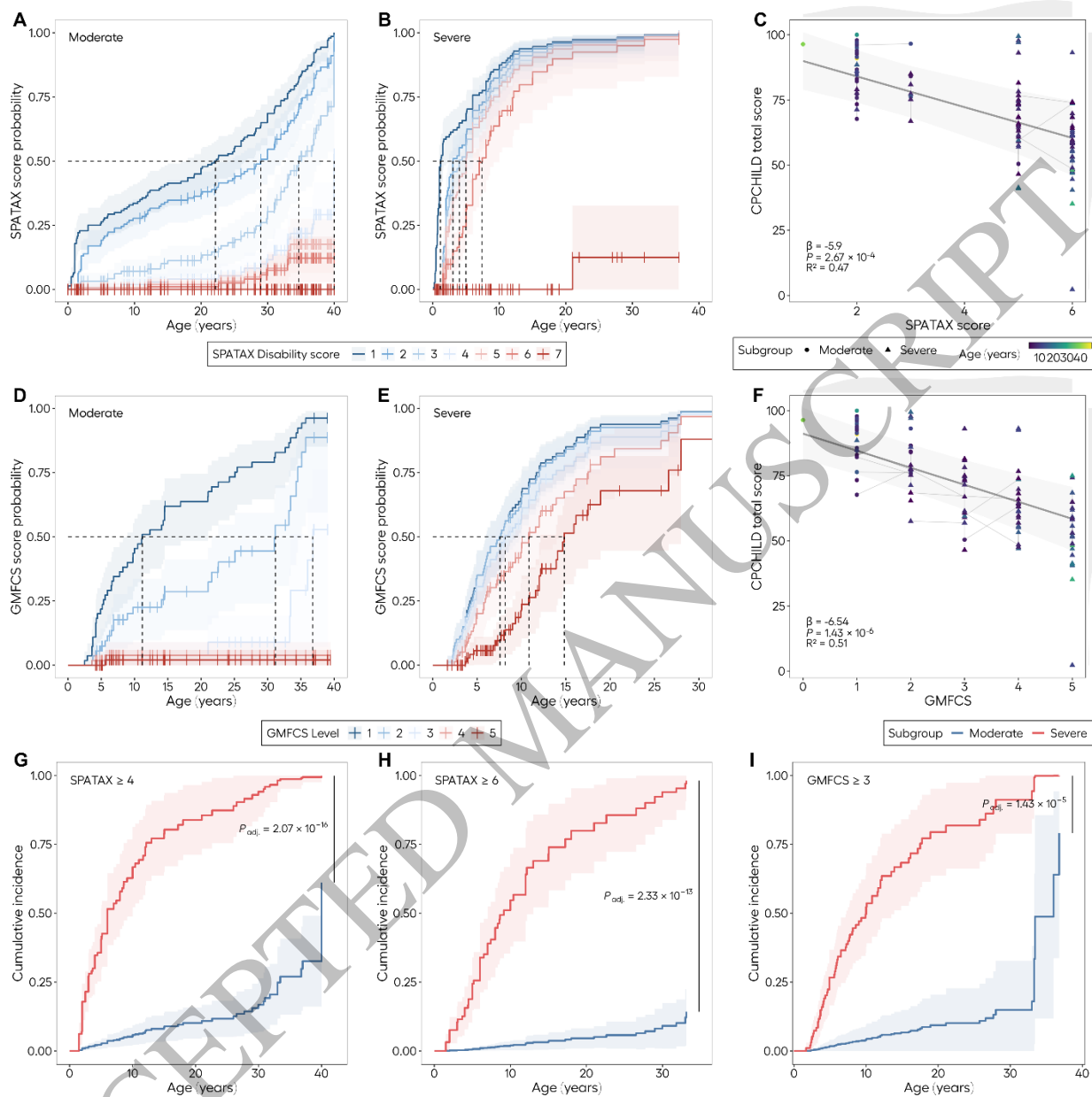
1 represents Cohen's d. Box-and-whisker plots display the median (center line), interquartile
 2 range (box), and minimum and maximum values (whiskers).



3
 4 **Figure 3 Stratification framework, quality of life and phenotypic spectra. (A)**
 5 Comparison of reported ages at onset in the combined dataset stratified by essentiality
 6 verdict, excluding uncertain classifications. Main plot: Kruskal-Wallis, $p < 1e-16$, p values of
 7 pairwise comparisons shown, using Dunn's post-hoc test; subplot: Mann-Whitney U test;
 8 d represents Cohen's d . Half violin and box plots show the distribution of the data as a
 9 kernel density estimate on the left and a box-and-whisker summary on the right, indicating
 10 the median (center line), interquartile range (box), and minimum and maximum values
 11 (whiskers). **(B)** Schematic of the proposed stratification framework. **(C)** Interaction plot,
 12 with dots representing estimated marginal means (EMM) at selected timepoints (3, 5, 10,
 13 13.7, 20, 25, 30 years). Vertical ribbons indicating EMM 95%-CIs. Top row p values for
 14 cross-sectional EMM contrasts at selected timepoints. **(D)** Separate subgroup models with
 15 kernel density estimates. Longitudinal observations from the same patient are connected
 16 by lines. **(E)** Results from phenotypic enrichment analysis using Firth's logistic regression.
 17 Labelled dots represent clinical findings that were nominally enriched, and black circles
 18 surrounding dots indicate $q < 0.05$. **(F)** Frequency of extrapyramidal movement disorders
 19 stratified by subgroup.



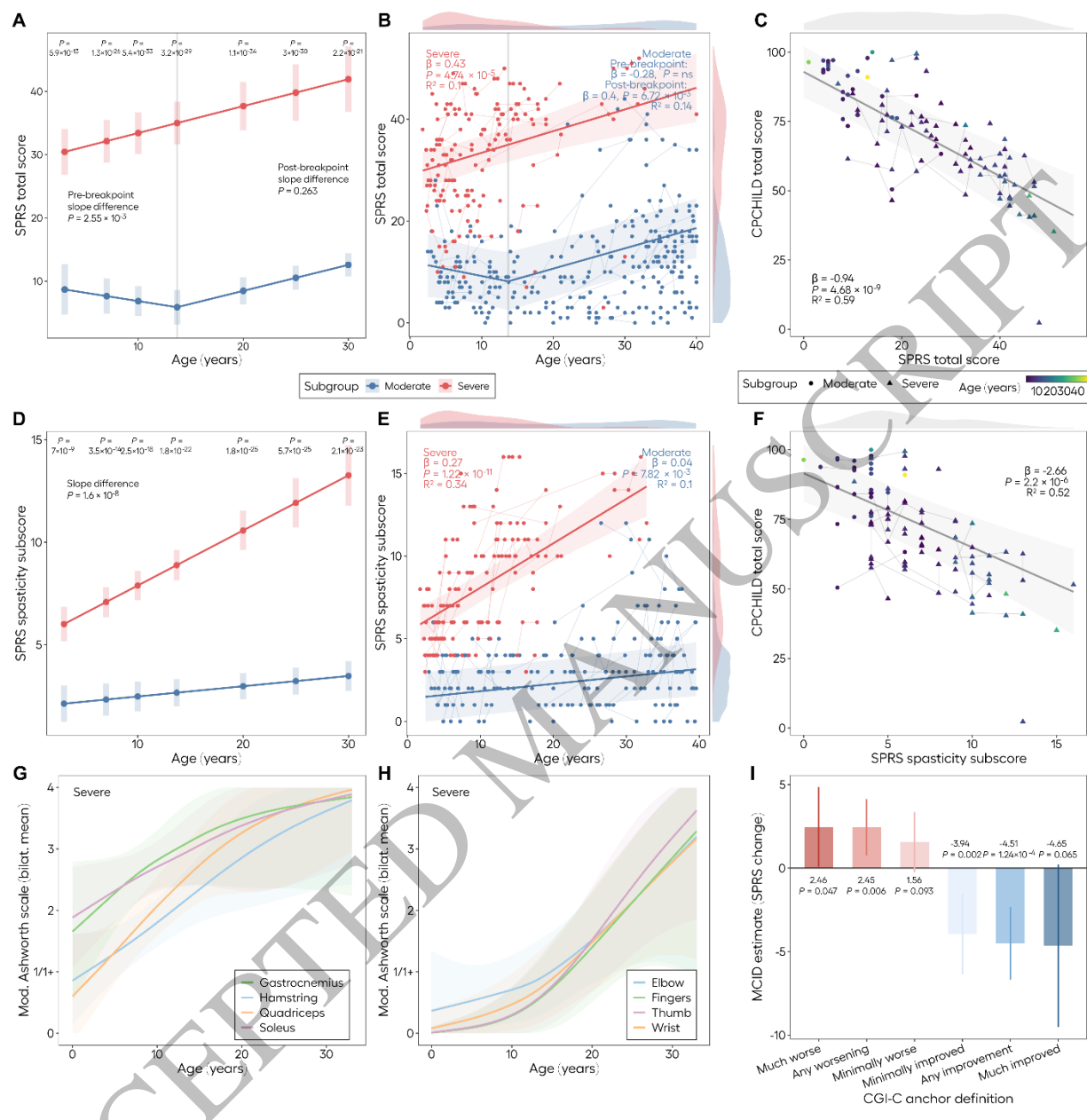
1 **Figure 4 Divergent functional motor impairments between subgroups.** Kaplan-Meier
2 estimators for SPATAX disability stages in the moderate (**A**) and severe (**B**) subgroup. 95%-
3 CIs are shown as shaded ribbons. (**C**) Association between SPATAX and CPCHILD. Linear-
4 mixed effects model adjusted for subgroup, age, and sex. Longitudinal observations from
5 the same patient are connected by lines. (**D & E**) Kaplan-Meier estimators for GMFCS
6 scores in the moderate (**D**) and severe (**E**) subgroup. 95%-CIs are shown as shaded
7 ribbons. (**F**) Association between GMFCS and CPCHILD. Linear-mixed effects model
8 adjusted for subgroup, age, and sex. Longitudinal observations from the same patient are
9 connected by lines. (**G, H & I**) Covariate-adjusted survival curves derived from Cox
10 proportional hazards models for SPATAX threshold severe gait impairment, requiring
11 unilateral support (≥ 4) (**G**) and wheelchair dependence (≥ 6) (**H**) and GMFCS threshold $\geq III$
12 (requiring assistive mobility device) (**I**). Covariates included subgroup, sex, and inheritance
13 pattern.



1
 2 **Figure 5 Differential progression of spasticity between subgroups.** (A) Interaction plot
 3 for segmented linear mixed-effects regression of the SPRS total score, with dots
 4 representing EMM at selected timepoints (3, 5, 10, 13.7, 20, 25, 30 years). Vertical ribbons
 5 indicating EMM 95%-CIs. Top row p values for cross-sectional EMM contrasts at selected
 6 timepoints. Gray vertical ribbon indicates estimated breakpoint for the moderate subgroup.
 7 (B) Separate subgroup models for SPRS total scores with kernel density estimates. (C)
 8 Association between SPRS total score and CPCHILD. Linear-mixed effects model adjusted
 9 for subgroup, age, and sex. (D) Interaction plot for linear mixed-effects regression of the
 10 SPRS spasticity subscore (SPRS items 7-10), with dots representing EMM at selected
 11 timepoints (3, 5, 10, 13.7, 20, 25, 30 years). Vertical ribbons indicating EMM 95%-CIs. Top

1 row p values for cross-sectional EMM contrasts at selected timepoints. **(E)** Separate
2 subgroup models for SPRS spasticity subscores with kernel density estimates. **(F)**
3 Association between SPRS spasticity subscore and CPCHILD. Linear-mixed effects model
4 adjusted for subgroup, age, and sex. **(G & H)** Ordinal generalized additive mixed-effects
5 models for modified Ashworth scale values stratified by lower (G) and upper (H) limb
6 muscle groups in the severe subgroup. Shaded ribbons indicate 95% CIs. **(I)** MCID
7 estimations estimates for SPRS total scores stratified by CGI-C anchor definition. Bars
8 representing SPRS change EMM derived from a linear-mixed-effects model adjusting for
9 baseline SPRS, age at anchor assessment, interval between visits, subgroup and sex as
10 covariates. Vertical lines indicate EMM 95% CIs. **(B, C, E & F)** Longitudinal observations
11 from the same patient are connected by lines.

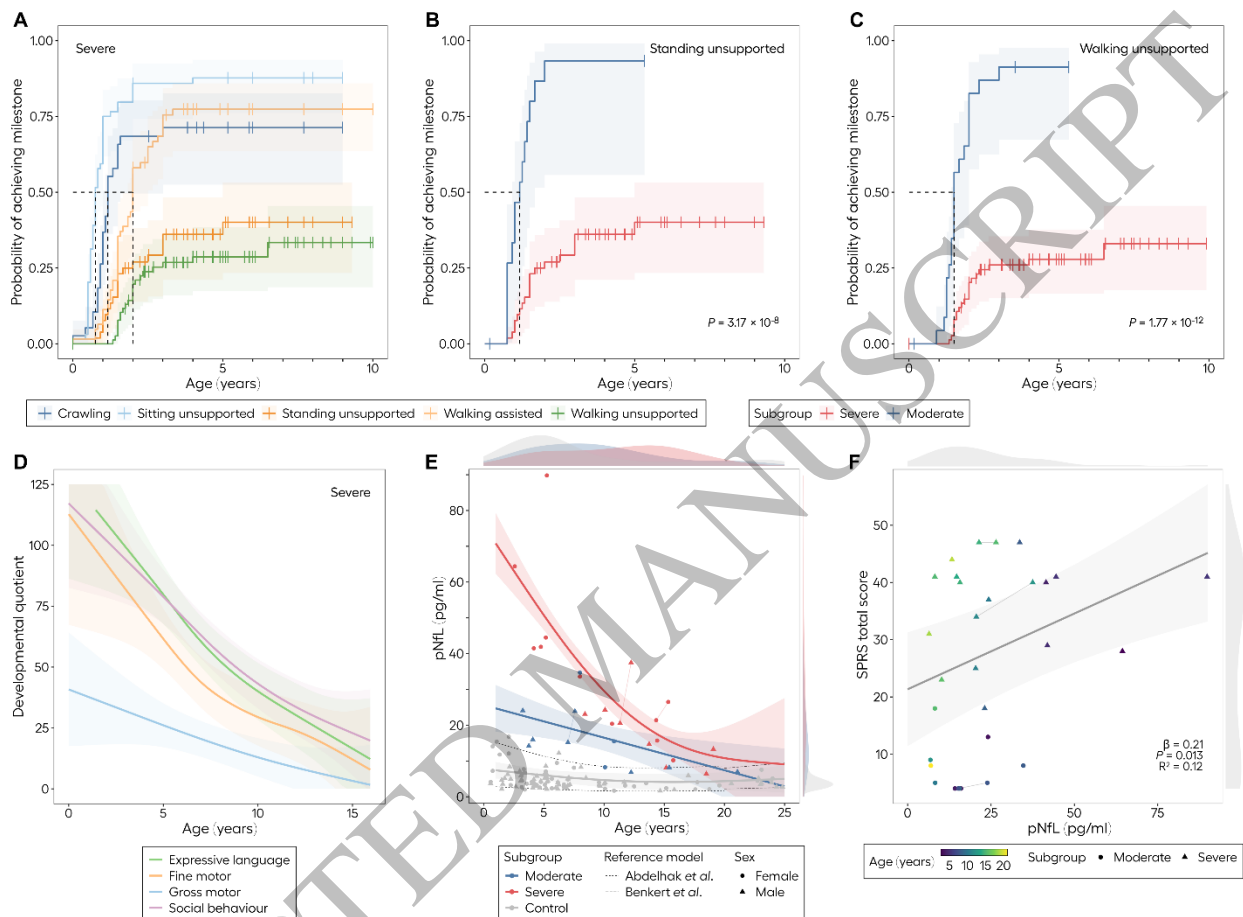
ACCEPTED MANUSCRIPT



1
 2 **Figure 6 Developmental trajectories and plasma NfL.** (A) Kaplan-Meier estimators for
 3 time-to-milestone attainment in the severe subgroup. Shaded ribbons indicate 95% CIs. (B
 4 & C) Kaplan-Meier estimators for time-to-unsupported standing (B) and time-to-
 5 unsupported walking (C) stratified by subgroup. Curves were compared using log-rank test.
 6 (D) Generalized additive mixed-effects modeling of developmental quotients stratified by
 7 domain in the severe subgroup. Shaded ribbons indicate 95% CIs. (E) Plasma NfL over time
 8 stratified by subgroup with kernel density estimates. Dotted lines indicate 1st and 99th
 9 percentile of pNfL reference models in healthy individuals derived from previously
 10 published studies. (F) Association between pNfL and SPRS total scores. Linear-mixed

1 effects model adjusted for subgroup, age, and sex. (E & F) Longitudinal observations from
 2 the same patient are connected by lines.

3



4

5

6